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**Mechano-reciprocity is maintained between physiological boundaries by tuning signal flux through the Rho-associated protein kinase**

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1       **Mechano-reciprocity is maintained between physiological boundaries by**  
2       **tuning signal flux through the Rho-associated protein kinase**

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1 **Abstract**

2 The mechanical properties of the ECM strongly influence the behavior of all cell  
3 types within a given tissue. Increased matrix tension promotes epithelial cell  
4 proliferation by engaging mitogenic mechanotransduction signaling including the  
5 Salvador/Warts/Hippo, PI 3-kinase, Rho, Wnt and MAP kinase pathways. The Rho  
6 signaling pathways in particular are capable of increasing intra-cellular tension by  
7 elevating the production and contractility of the actomyosin cytoskeleton, which  
8 counteracts tension changes within the matrix in a process termed mechano-  
9 reciprocity. We have discovered that Rho-ROCK signaling increases the production  
10 of ECM through paracrine signaling between the epithelium and fibroblasts and also  
11 the remodeling of the ECM by regulating focal adhesion dynamics in fibroblasts.  
12 These two phenomena together cause increased ECM tension. Enhanced mechano-  
13 reciprocity results in ever-increasing intra- and extra-cellular tension in a vicious  
14 cycle that promotes cell proliferation and tumor progression. These insights reveal  
15 that inhibiting mechano-reciprocity, reducing ECM tension and targeting cancer-  
16 associated fibroblasts in a coordinated fashion has potential as cancer therapy.

## 1 **Introduction**

### 2 *Mechanical forces in biology*

3 The capacity to exert mechanical force upon an object is a fundamental requirement  
4 for physical interaction with the environment. Flowing from this is the need to detect  
5 and measure external forces exerted upon the body. Everyday activities like sitting,  
6 standing and handling objects require not only the exertion of mechanical force, but  
7 the ability to detect mechanical signals and coordinate a timely response. This is  
8 facilitated by the sense of proprioception, which is mediated by mechanoreceptors,  
9 nerve endings specialized for the detection of forces. However, it has been clear for  
10 some time that mechanical force can be detected not only by specialized nerve  
11 receptors, but also by all other cell types. Indeed, the generation and detection of  
12 mechanical force is a key aspect of cell and developmental biology. Cells interact  
13 with their environment by exerting and sensing mechanical forces between  
14 themselves and the extra-cellular matrix (ECM). These mechanical forces greatly  
15 influence cell behavior, by guiding decisions about cell migration, growth, division  
16 and differentiation.

### 17 *Mechanical signaling and cancer*

18 Whereas the biochemical aspects of unrestrained cell growth and proliferation in  
19 cancer have been extensively studied over many decades, an understanding of the  
20 biophysical mechanisms underlying how force and mechanical stress influence  
21 tumor development is only just emerging. However, as early as 1972, reports  
22 appeared that mechanical force has the capacity to influence tumor growth and  
23 metastasis<sup>1</sup>. Throughout the late 70s and early 80s, reports emerged that the  
24 application of mechanical stress upon cells is capable of eliciting distinct phenotypic

1 responses<sup>2-4</sup>. The first clues into how mechanical force may be converted to a  
2 biochemical signal for propagation within the cell were provided by Carvalho et al.<sup>5</sup>,  
3 who demonstrated that integrin expression and subcellular distribution is markedly  
4 altered upon the application of external force on cells. As integrins were well known  
5 to link cells to the extra-cellular matrix<sup>6</sup> and were dysregulated in cancers, they then  
6 became the prime candidates for the mechanosensing (**Figure 1**) receptors  
7 transducing mechanical signals to the intra-cellular biochemical machinery. This  
8 pioneering work caused a flurry of papers providing evidence for a link between  
9 extra-cellular mechanical stresses and changes in actin cytoskeletal structure,  
10 adhesion complex composition and number, cell migration and spreading, and  
11 proliferation via integrin ligation (reviewed in <sup>7</sup>), thereby linking extra-cellular  
12 biophysical signals to intra-cellular biochemical changes. This process became  
13 known as mechanotransduction (**Figure 1**) and several signaling pathways including  
14 those mediated by Salvador/Warts/Hippo<sup>8</sup>, PI 3-kinase<sup>9</sup>, Rho small GTPases<sup>10</sup>,  
15 Wnts<sup>11,12</sup> and MAP kinases<sup>10</sup> have been shown to be activated downstream of  
16 changes in the mechanical properties of the ECM.

17 It therefore follows that changes in the mechanical properties of the ECM have  
18 implications for tumor progression, as mitogenic pathways are directly linked to  
19 changes in ECM stiffness via mechanotransduction. However, the observation that  
20 ECM was increased in many different cancers and directly influenced tumor growth  
21 and spread<sup>13,14</sup> suggested that tumors themselves were capable of strongly  
22 promoting changes in the ECM that facilitated their growth and spread. Several  
23 growth factors of tumor origin, including TGF $\beta$ , CTGF, IL6 and LIF are known to  
24 directly act upon tumor-associated fibroblasts and other stromal cells to increase the  
25 production and remodeling of ECM molecules to increase ECM stiffness<sup>15-18</sup>. While

1 some of the mechanisms underlying mechano-sensation, mechanotransduction and  
2 conversely the direct involvement of tumors in regulating ECM stiffness are  
3 beginning to be uncovered, a precise understanding of how these processes are  
4 integrated and coordinated still remains elusive.

### 5 **The Rho family of small GTPases and the regulation of intracellular tension**

6 Rho (Ras homology) GTPases are monomeric GTP-binding proteins and are a  
7 subset of the Ras superfamily, comprising 22 members of which the best  
8 characterized are Rho, Rac and Cdc42. Rho family members act as molecular  
9 regulators of signal transduction by switching between GDP-bound inactive and  
10 GTP-bound active states, and have numerous molecular targets including kinases,  
11 transcription factors and scaffold proteins that mediate diverse cellular processes  
12 such as migration, proliferation, adhesion and apoptosis in cell type and temporal  
13 context-dependent ways (reviewed in <sup>19</sup>). The regulation of actomyosin cytoskeletal  
14 dynamics by these proteins is the best studied aspect of their biology.

15 Dynamic remodeling of the actin cytoskeleton to generate intra-cellular force is key  
16 to cell migration. The extension of actin filament networks by actin polymerization  
17 generates intracellular forces, as filaments in the leading edge are compressed  
18 between transient associations with the cell membrane and the bulk of the actin  
19 cytoskeletal network behind them. As protrusions grow and retract, individual actin  
20 filaments undergo tension from transient bonds with the membrane, and are bent or  
21 compressed depending on their orientation<sup>20</sup>. Myosin-mediated contraction of the  
22 actin cytoskeleton results in a reduction in plasma membrane surface area and  
23 causes clustering of cell surface integrins, which are responsible for attachment to  
24 the ECM. Integrin-mediated ECM adhesion activates guanine nucleotide exchange

1 factors (GEFs), which enhance RhoA activity<sup>21</sup>. During ameboid cell movement, Rac  
2 interacts with lamellipodin at the leading edge to extend actin filaments and generate  
3 lamellipodia, sheet-like cell protrusions supported by short actin filaments in a  
4 branched network, which adhere to substrates and determine the direction of cell  
5 movement<sup>22</sup>. Cdc42 controls the formation of filopodia, thin protrusions from the cell  
6 membrane that contain parallel actin bundles for mechanosensing the environment  
7 in order to determine cell polarity – another mechanism by which the direction of  
8 movement is determined<sup>23</sup>. In order to direct migration, actin polymerization must be  
9 restricted to a portion of the plasma membrane. Cdc42 directs this polarity, by acting  
10 through the Par polarity complex, localizing Rac activity and stabilizing microtubules  
11 within the cytoskeleton. At the trailing edge of cells exhibiting mesenchymal motility,  
12 and to a lesser extent those exhibiting ameboid motility, RhoA controls retraction  
13 through activation of effector kinases such as Rho-associated kinase (ROCK)<sup>24,25</sup>.  
14 Furthermore, Rho-mediated activation of ROCK is required for the formation of  
15 membrane ruffles and lamellae and in particular membrane blebs. Membrane blebs  
16 are a feature of ameboid motility (also termed blebbing motility), a mode of cell  
17 migration that is independent of lamellipodia and frequently observed in cancer  
18 cells<sup>26</sup>. Rho signaling is also responsible for the generation of intra-cellular tension,  
19 which is required for the morphological changes associated with other cellular  
20 processes such as apoptosis<sup>27,28</sup> and cytokinesis<sup>29</sup>.

21 The effector proteins of Rho signaling, Rho-associated kinases 1 and 2 (ROCK1 and  
22 ROCK2), are serine-threonine kinases containing a Rho-binding domain and are  
23 activated upon interaction with Rho-GTP<sup>30</sup>. Activation of ROCK by Rho induces  
24 stress fiber assembly in a number of ways. ROCK directly phosphorylates and  
25 activates the regulatory myosin light chain (MLC)<sup>31</sup>, it indirectly increases MLC

1 phosphorylation by phosphorylating and inhibiting the myosin targeting subunit of  
2 MLC phosphatase (MYPT)<sup>32</sup> and directly phosphorylating and activating the LIM  
3 kinases (LIMK1 and LIMK2), which subsequently phosphorylate and inhibit cofilin  
4 and stabilizes actin structures<sup>33</sup>. In cultured cells, parallel bundles of actin filaments  
5 form stress fibers that link sites of focal adhesions, permitting tension to be  
6 transmitted between focal adhesions and thereby to the ECM. As well as being the  
7 product of intracellular tension, stress fibers themselves are able to exert force upon  
8 focal adhesions, permitting forces exerted by the external environment to be  
9 counteracted<sup>21</sup>.

## 10 **Rho-ROCK signaling and the regulation of extracellular tension**

11 Not surprisingly, cells are not merely passive reactors to changes in the mechanical  
12 properties of the ECM, but are active players in the remodeling of their  
13 environments. Protein components of the ECM can be degraded by a variety of  
14 proteinases such as the families of matrix metalloproteinases (MMPs)<sup>34</sup> and a  
15 disintegrin and metalloproteinases (ADAMs)<sup>35</sup> that are produced by cancer cells to  
16 remodel the ECM. While most proteinases acting on the ECM are secreted, a subset  
17 of MMPs are membrane tethered, permitting cells to exert a greater degree of spatial  
18 control over their deployment<sup>36</sup>. The ECM may also be remodeled by changes to its  
19 composition and level of cross-linking. Lysyl oxidases and lysyl hydroxylases are  
20 produced by cells to crosslink collagen chains, thereby increasing ECM stiffness<sup>37</sup>.  
21 The mechanical properties of the ECM are also significantly altered in diseased  
22 states, such as chronic wounds, cancer and fibrosis, by changes to ECM  
23 composition<sup>38,39</sup>.



1 Recently, we demonstrated a mechanism by which epithelial cells and tumor cells of  
2 epithelial origin are capable of increasing the mechanical stiffness of the ECM by  
3 elevating the production of ECM components<sup>14,39,40</sup>. Activation of ROCK in the  
4 context of tumor cells or the hyper-proliferation of epidermal cells in the context of  
5 wound healing results in paracrine signals arising from the proliferating epithelia that  
6 act on stromal fibroblasts, increasing their production of collagen, fibronectin,  
7 periostin and tenascin C, components of the ECM, and thereby increasing matrix  
8 stiffness<sup>14,39</sup>. While the nature of the specific paracrine signals caused by activation  
9 of ROCK remains to be uncovered, cytokines and other secreted molecules of tumor  
10 origin have been previously implicated in the recruitment and activation of cancer-  
11 associated fibroblasts, the key regulators of ECM composition and stiffness in the  
12 tumor context<sup>15-17</sup>. Nevertheless, ROCK activation in tumor cells results in increased  
13 ECM stiffness of the magnitude frequently observed in epithelial cancers including  
14 cutaneous squamous cell carcinoma<sup>14</sup>, breast cancer<sup>13</sup> and pancreatic cancer<sup>41</sup> and  
15 promotes tumor progression. We therefore propose that ROCK integrates inputs  
16 from growth factor and mechanotransduction signaling to produce the appropriate  
17 cellular response, be it migration, proliferation, ECM production or remodeling.

### 18 **Biochemical changes arising from increased extra-cellular tension**

19 External forces drive clustering of integrin molecules at the cell membrane,  
20 stabilizing focal adhesions and linking the intracellular actin cytoskeleton to the ECM.  
21 The stiffer the ECM or substrate to which a cell is attached, the greater the size and  
22 strength of those focal adhesions, and the greater the cellular response, as intra-  
23 cellular forces are generated to balance the extra-cellular forces. Upon integrin  
24 activation, talin and vinculin are recruited to the complex, and focal adhesion kinase  
25 (FAK) is auto-phosphorylated at Y397 and/or Y925 and activated. FAK

1 phosphorylation results in the recruitment of Src-homology-2 (SH2) domain-  
2 containing proteins Src and Shc that link FAK to the Ras pathway and Rho-ROCK  
3 pathway and, via PI 3-kinase and Akt activation, to the stabilization of  $\beta$ -catenin and  
4 the transcriptional activity of TCF/LEF transcription factors. Extracellular signal-  
5 regulated kinase (ERK) is also regulated by the FAK-Src complex, and is responsible  
6 for mechanical signal transduction from the microenvironment, to regulate  
7 intracellular processes<sup>10,14,42</sup>.

8 Responses to cell-matrix interactions are governed by a complex interplay between  
9 signaling pathways involving FAK, ERK,  $\beta$ -catenin, Rho and others, and cells are  
10 therefore able to respond to extracellular forces in myriad ways. For example, in the  
11 mammary gland, where development and homeostasis occur within the context of a  
12 pliable ECM, Rho activity in fibroblasts was increased upon heightened matrix  
13 rigidity. This led to an increase in focal adhesion assembly and growth factor-  
14 dependent ERK activation, suggesting that a mechano-regulatory “circuit”  
15 amalgamates stimulatory extracellular tension with focal adhesion formation through  
16 ERK and Rho-dependent cytoskeletal changes, to maintain homeostasis<sup>13</sup>.

### 17 **Mechano-reciprocity**

18 Mechano-reciprocity (**Figure 1**) is the ability of cells to enhance or moderate intra-  
19 cellular tension in order to adapt to increased or reduced extra-cellular stiffness  
20 respectively<sup>43</sup> (termed outside-in signaling), but conversely may also refer to their  
21 ability to remodel the extra-cellular matrix and modify its mechanical properties in  
22 order to offset changes in intra-cellular tension (inside-out signaling). In a situation  
23 where sustained growth factor stimulation or oncogenic mutation causes increased  
24 mechanotransduction signaling such as via the Rho, YAP,  $\beta$ -catenin and/or MAP

1 kinase pathways, the combination of inside-out and outside-in signaling has the  
2 potential to establish a vicious cycle by which ever-escalating ECM stiffness causes  
3 uncontrolled cell proliferation. Indeed, this is observed in mechano-responsive  
4 cancers such as those of the skin, breast and intestine<sup>14,44</sup> and is characterized by  
5 persistent activation of the ROCK protein. We have shown that enhanced mechano-  
6 reciprocity of this kind promotes tumor progression<sup>14</sup>. Targeting runaway mechano-  
7 reciprocity is therefore a novel way in which diseases of cellular homeostasis  
8 characterized by increased ECM stiffness such as cancer or fibrosis may be treated.

### 9 **Mechano-reciprocity in diseases of cellular homeostasis**

10 It is becoming increasingly evident that persistent mechanical signaling can advance  
11 cancer progression. Overexpression of Rho GTPases has been associated with  
12 progression of disease in a number of malignancies, and Rho, Rac, Cdc42 and  
13 ROCK<sup>45</sup> have all been reported to be mutated in various cancers to confer tumor  
14 promoting functions and inhibit tumor suppressive functions (reviewed in <sup>46</sup>).

15 The role of mechanical signaling in driving tumorigenesis has been well  
16 characterized in the mammary gland. A known mechano-responsive tissue<sup>44</sup>, the  
17 mammary epithelium is subjected to a multitude of external forces throughout  
18 development and cycling, such as ductal morphogenesis, lactation, and involution to  
19 remodel the gland and degrade the ECM<sup>47</sup>. A pliable ECM is best suited to these  
20 morphological changes. Compression analysis of normal and tumor tissue of murine  
21 mammary origin demonstrated that although normal mammary tissue was soft,  
22 tumor and peri-tumoral tissue was significantly stiffer<sup>10</sup>. Matrix stiffness is promoted  
23 by a greater amount of collagen cross-linking<sup>48</sup>. This ECM stiffening is sensed by  
24 cellular integrins, which in turn activate RhoA signaling, resulting in generation of

1 focal adhesions and influencing tumor cell invasion, cell proliferation, and changes in  
2 cytoskeletal organization. Thus, a persistent and high level of mechano-reciprocity is  
3 potentially able to create a feed forward mechanism, promoting further cancer  
4 development. This phenotype is often seen in patients with a high mammographic  
5 density and it is becoming clearer that mammographic density in the normal breast is  
6 an important prognostic marker for breast cancer, with a higher density indicating a  
7 four-to-six fold higher likelihood of developing breast cancer<sup>49</sup>.

8 We have found that skin in which ROCK had been hyper-activated exhibited  
9 increased MLC2 phosphorylation and elevated ECM production resulting in greater  
10 tissue stiffness. Increased tissue stiffness gave rise to mechano-reciprocal activation  
11 of the PI 3-kinase signaling pathway which led to the stabilization of  $\beta$ -catenin and  
12 epidermal hyper-proliferation. Interestingly, in the context of the multi-step chemical  
13 carcinogenesis model, mechano-reciprocity cooperated with oncogenic  
14 transformation to promote tumor progression in a manner that was dependent on  
15 ROCK-induced tissue stiffness. Conversely, inhibition of ROCK signaling by  
16 treatment with the inhibitor Y-27632 lowered ECM stiffness and impeded tumor  
17 formation and growth<sup>14,50</sup>. These observations clearly show that tumor cells have a  
18 key role in establishing a stiff ECM that promotes their own proliferation and growth.

19 A number of parallels exist between normal and malignant tissue development and  
20 wound healing. Rapid cell proliferation and mechanisms mediating cytoskeletal  
21 changes during cell migration are similar in acute wound healing to those involved in  
22 tumor progression. At all stages of wound healing, including hemostasis,  
23 inflammation, proliferation and remodeling, examples of mechano-reciprocity exist  
24 between the ECM and a number of cell types including platelets, immune cells,  
25 fibroblasts, endothelial cells and keratinocytes<sup>51</sup>. We have recently reported that

1 enhanced signaling through ROCK, occurring during either cutaneous squamous cell  
2 carcinoma progression or acute skin wound healing and which causes increased  
3 ECM production in both contexts, is negatively regulated by the phospho-serine  
4 binding molecular adaptor protein 14-3-3 $\zeta$ <sup>39</sup>. 14-3-3 $\zeta$  bound to the ROCK antagonist  
5 MYPT1, promoting its MLC phosphatase function and antagonizing ROCK-mediated  
6 phosphorylation of MLC. Mice lacking 14-3-3 $\zeta$  exhibited increased signal flux through  
7 ROCK leading to enhanced mechano-reciprocity, causing rapid wound healing. This  
8 observation was corroborated using a novel pharmacological inhibitor of 14-3-3 $\zeta$ ,  
9 which accelerated acute wound healing in wild-type mice<sup>39</sup>.

10 Interestingly, this negative regulatory mechanism that moderated signaling through  
11 ROCK and thereby limited mechano-reciprocity, was lost in the majority of patient  
12 samples of cutaneous squamous cell carcinoma, which exhibited little or no 14-3-3 $\zeta$   
13 compared to normal skin. Accordingly, 14-3-3 $\zeta$ -deficient mice on the multi-step  
14 chemical carcinogenesis protocol formed larger tumors than wild-type mice of the  
15 same strain background. Taken together, these observations lead us to the  
16 conclusion that 14-3-3 $\zeta$ -mediated promotion of MLC dephosphorylation is a  
17 mechanism by which mechano-reciprocity is maintained within manageable limits  
18 such that normal wound healing is facilitated while tumor formation is prevented  
19 **(Figure 2)**.

20 We do not believe that 14-3-3 $\zeta$  is the only negative regulator of mechano-reciprocity.  
21 Under steady-state conditions, 14-3-3 $\zeta$ -deficient mice exhibit thinner skin than wild-  
22 type mice<sup>39</sup>, suggesting that compensatory mechanisms as yet undiscovered and not  
23 involving the seven other 14-3-3 family members are capable of maintaining normal  
24 cellular homeostasis at near-physiological rates.

## 1 **Targeting mechano-reciprocity and the extra-cellular matrix as therapy**

2 Given the highly tumor-promoting environment caused by increased mechano-  
3 reciprocity, targeting the players that facilitate it (ROCK signaling, fibroblasts and  
4 their activation) and enhancing the activities of its negative regulators (14-3-3 $\zeta$ ) are  
5 potentially novel approaches to cancer therapy. However, an important consideration  
6 is the effect that inhibition of pleiotropic proteins like ROCK or 14-3-3 $\zeta$  may have on  
7 cellular homeostasis and indeed normal development. For instance, tumor capillary  
8 endothelial cells subjected to stress exhibited cytoskeletal rearrangements, and  
9 exerted stronger Rho-ROCK-mediated traction compared to non-cancer cells. Pre-  
10 treatment with the ROCK inhibitor Y-27632 before the application of stress restored  
11 normal actin behavior in tumor cells, but caused significant cytoskeletal disruption in  
12 normal cells<sup>52</sup>. This indicates that normal and tumor cells differ in sensitivity to  
13 external mechanical force as governed by ROCK signaling and illustrates the  
14 challenges accompanying attempts to target tumors via the mechanotransduction  
15 machinery.

16 The most common current therapy regimes involve targeting tumor cells directly via  
17 inhibition of cell-intrinsic functions such as their proliferative capacity or their ability to  
18 evade apoptosis, whereas the therapies that offer the most promise, such as  
19 immunotherapy or hormone therapy, target non-intrinsic functions that require cell to  
20 cell communication. Their need for cell to cell communication could be viewed as a  
21 vulnerability of cancers. The cell to ECM communication mechanisms that enhance  
22 mechano-reciprocity are mediated by communication between cancer cells and  
23 normal (fibroblast) cells lacking oncogenic mutations, a particularly serious  
24 vulnerability. Normalizing the ECM by normalizing tumor-associated fibroblasts  
25 therefore holds out the tantalizing possibility of therapies that halt or even reverse

1 tumor progression. This approach may also provide new opportunities for  
2 combination therapies where the tumor and its ECM are both targeted concurrently.

### 3 **Concluding Remarks**

4 In conclusion, analyzing Rho-ROCK pathway activation in the in vivo context has  
5 revealed its function in augmenting mechano-reciprocity via enhanced ECM  
6 stiffness. These observations have also highlighted the importance of mechano-  
7 reciprocity in normal tissue homeostasis and demonstrated that negative regulators  
8 of this process have significant therapeutic utility.

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24

1 **Figure Legends**

2 Figure 1

3 Mechanosensing through engagement of integrins with the ECM causes  
4 mechanotransduction, by activation of Talin and autophosphorylation of FAK, which  
5 initiates several intra-cellular signal transduction pathways, including PI 3-kinase/Akt  
6 and ROCK signaling. Mechanotransduction gives rise to increased actin  
7 polymerization and actomyosin contractility, establishing mechano-reciprocity, which  
8 in turn leads to paracrine signaling between the parenchyma and the stroma,  
9 increasing ECM production and remodeling.

10 Figure 2

11 ROCK activation enhances intracellular tension in epidermal cells by activating MLC  
12 and LIM kinases. It also increases extracellular tension by elevating ECM production  
13 by dermal fibroblasts. ROCK activation in dermal fibroblasts promotes ECM  
14 remodeling by regulating focal adhesion dynamics and fibroblast migration. In both  
15 contexts, 14-3-3 $\zeta$  limits signal flux through ROCK, thereby maintaining mechano-  
16 reciprocity between normal physiological boundaries, permitting normal wound  
17 healing and protecting against tumor formation. This figure has been adapted from  
18 <sup>50</sup>.

Figure 1

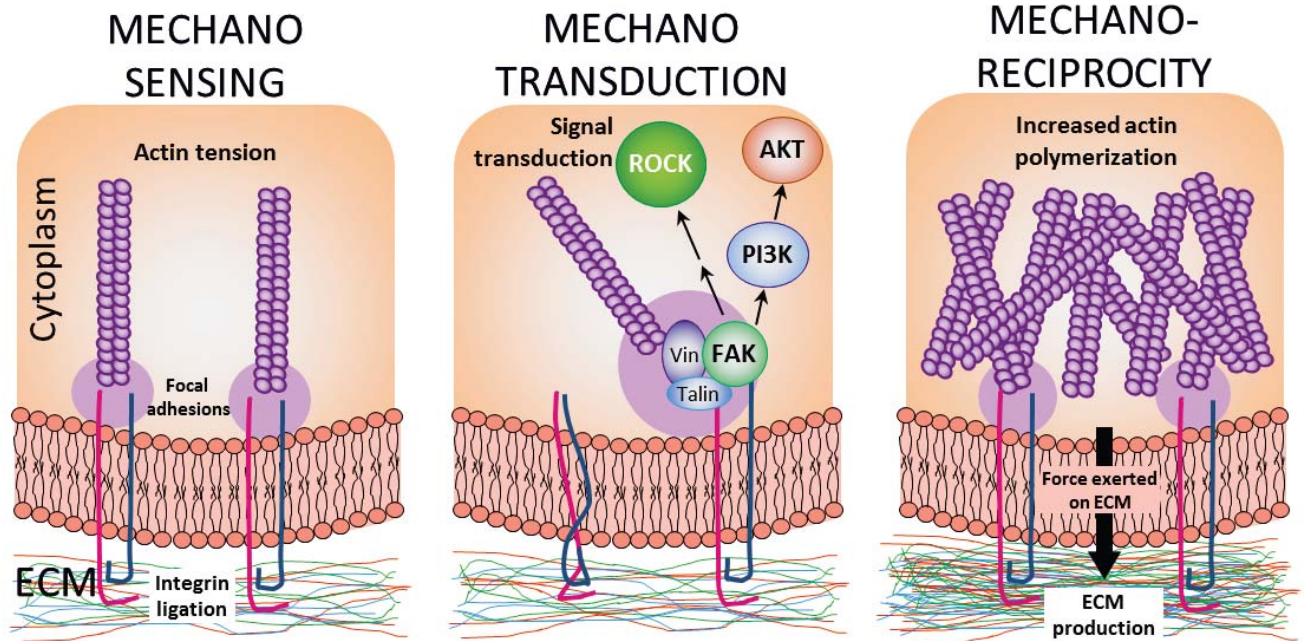


Figure 2

