Intracellular Microenvironment Responsive Nanogels for Gene Delivery

Bingyang Zhang

A thesis submitted for the degree of Master of Philosophy



School of Chemical Engineering
The University of Adelaide
Adelaide, Australia
October, 2015

Declaration

I certify that this work contains no material which has been accepted for the award of any

other degree or diploma in my name, in any university or other tertiary institution and, to the

best of my knowledge and belief, contains no material previously published or written by

another person, except where due reference has been made in the text. In addition, I certify

that no part of this work will, in the future, be used in a submission in my name, for any other

degree or diploma in any university or other tertiary institution without the prior approval of

the University of Adelaide and where applicable, any partner institution responsible for the

joint-award of this degree.

I give consent to this copy of my thesis when deposited in the University Library, being made

available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis

resides with the copyright holder(s) of those works.

I also give permission for the digital version of my thesis to be made available on the web, via

the University's digital research repository, the Library Search and also through web search

engines, unless permission has been granted by the University to restrict access for a period of

time.

Signature:_____

Date:

i

Acknowledgements

I would like to express my gratitude to all those who helped me during my study period at the University of Adelaide as a Master candidate.

First and foremost, my deepest gratitude goes to my supervisors Associate Prof. Jingxiu Bi and Associate Prof. Sheng Dai for the constant encouragement and guidance. A. Prof. Bi has walked me through all the stages of the experiments and the writing of this thesis. She has always been willing to discuss with me on academic problems and personal matters. A. Prof. Dai has helped a lot with my project design and management with his broad theoretical and experimental knowledge. The knowledge I learnt from him is not only benefiting my Master project but will benefit my whole academic career.

Many thanks go to my friends and all lab members and staffs of the School of Chemical Engineering for their help and support.

Finally, my thanks would go to my beloved family for their loving considerations and great confidence in me all through these years.

Abstract

Gene therapy is a modern technology applied by using therapeutic nucleic acids to treat or prevent diseases. The therapeutic nucleic acids can either be expressed to recombinant proteins for disease treatments or correct the genetic mutations in gene therapy for the treatments of various diseases. However, the development of gene therapy is greatly impeded by the limitation of gene delivery systems. Delivery of gene to eukaryotic cells is a multi-step process. Several barriers including cell membrane, nuclear pore complex (NPC) and intracellular enzymes have inhibited the transfection efficiency of gene. Therefore, the exploring of safe and high efficient gene carriers is essential for the development of gene therapy.

In this thesis, one of the widely studied cationic polymers, polyehtylenimine (PEI), was employed for modifications in the gene delivery application. High molecular weight PEI (HMW PEI) is able to perform high efficient gene transfection to eukaryotic cells due to its good buffering capacity. However, it also has high cytotoxicity to cells owing to its non-biodegradability and its molecular weight related high positive charge density. On the other hand, low molecular weight PEI (LMW PEI) such as PEI800 shows negligible cytotoxicity to cells, but has low transfection efficiency.

In order to reduce the cytotoxicity of PEI-based gene carrier while keep or improve the transfection efficiency, two biodegradable PEI-based gene carriers have been designed and successfully synthesized. The physical and chemical properties of both synthetic carriers were measured. The gene delivery performances of both carriers were also evaluated against Hela and HEK 293 cells. The synthetic biodegradable gene carriers display good gene binding ability, low cytotoxicity and good buffering capacity. In addition, both carriers are able to perform gene transfection with high efficiency.

Declaration	i
Acknowledgements	ii
Abstract	iii
Chapter 1 Introduction	1
1.1 Background	1
1.2 Aims and Objectives	3
1.3 Thesis outline	4
Reference	5
Chapter 2 Literature review	6
2.1 Introduction	6
2.2 Progress of gene delivery polymers	8
2.2.1 Condensing nucleic acid	9
2.2.2 Cellular uptake	10
2.2.3 Endosomal escape	11
2.2.4 Nuclear trafficking	12
2.3 Considerations for gene carrier design	12
2.4 PEI-based gene carriers	14
2.4.1 General modifications of PEI-based gene carriers	16
2.4.2 Biodegradable PEI-based gene carriers	19
2.4.2.1 Ester linkage	19
2.4.2.2 Imine linkage	21
2.4.2.3 Carbamate linkage	21
2.4.2.4 Disulfide linkage	21
2.5 Cell penetrating peptides (CPPs)	23
2.5.1 HIV-1 Trans-Activator of Transcription (TAT) peptide and its applications	25
2.6 Summary	26

References	28
Chapter 3 Disulfide Cross-linked Cationic Nanogels as Robust Gene carriers	38
Abstract	40
3.1 Introduction	41
3.2. Materials and methods	43
3.2.1 Materials	43
3.2.2 Methods	43
3.2.2.1 Preparation of plasmid DNA	43
3.2.2.2 Synthesis of PEI-DA	44
3.2.2.3 Characterization of PEI-DA	44
3.2.2.4 Buffering capacity	44
3.2.2.5 Gene condensing ability of PEI-DA	45
3.2.2.6 Particle size and zeta potential	45
3.2.2.7 Cell cytotoxicity	45
3.2.2.8 Cellular uptake by confocal laser scanning microscopy (CLSM)	46
3.2.2.9 Gene transfection	46
3.2.2.10 Green fluorescent protein (GFP) expression measurement by flow	•
3.2.2.11 Statistical analysis	
3.3 Results and discussion	
3.3.1 Synthesis and characterization of PEI-DA	48
3.3.2 Buffer capacity	49
3.3.3 Gene binding ability of PEI-DA	49
3.3.4 Particle size and zeta potential	50
3.3.5 Cell viability of PEI-DA	51
3.3.6 Cellular uptake of PEI-DA/pDNA complex	52
3 3 7 Cell transfection	53

3.4 Conclusions	53
References	55
Chapter 4 Cell Penetrating Peptide Labelled Intracellular Microenvironment Romanogels for Advanced Gene Delivery	_
Abstract	70
4.1 Introduction	71
4.2 Materials and Methods	73
4.2.1 Materials	73
4.2.2 Methods	74
4.2.2.1 Preparation of plasmid DNA	74
4.2.2.2 Synthesis of PEI-DA-TAT	74
4.2.2.3 Fourier transform infrared spectroscopy (FTIR)	75
4.2.2.4 Acid–Base Titration Assays	75
4.2.2.5 DNA condensing ability of PEI-DA-TAT	76
4.2.2.6 Particle sizes and zeta potentials	76
4.2.2.7 Evaluation of cytotoxicity	76
4.2.2.8 Cellular uptake by confocal laser scanning microscopy (CLSM)	77
4.2.2.9 Gene transfection	78
4.2.2.10 Green fluorescent protein (GFP) expression	78
4.2.2.11 Statistical analysis	79
4.3 Results and discussion	79
4.3.1 Synthesis and characterization of PEI-DA-TAT	79
4.3.2 Buffering capacity	80
4.3.3 Characterization of PEI-DA-TAT/pDNA complexes	81
4.3.4 Cell toxicity of PEI-DA-TAT	83
4.3.5 Cellular uptake of PEI-DA-TAT/pDNA complexes	84
4.3.6 Cell transfection	85

4.4 Conclusions	86
References	87
Chapter 5 Conclusions and future directions	99
5.1 Conclusions	99
5.2 Future directions	100

List of Tables

Table 2.1 The intracellular and extracellular barriers in polymer mediated gene delivery	7
Table 2.2 Considerations of gene carrier design	13
Table 2.3 CPPs and their sequence	23
Table 2.4 List of molecules conjugated with TAT for efficient intracellular uptake	25
List of Schemes	
Scheme 3.1 Schematic description on the synthesis of PEI-DA	58
Scheme 4.1 Schematic description on the synthesis of PEI-DA-TAT	91
List of Figures	
Figure 1.1 The design and experimental structure of the Master project	3
Figure 2.1 Polymer based gene delivery system (modified form Chenoweth et.al)	8
Figure 2.2 The self-assembled cationic polymer/gene structure	10
Figure 2.3 Proton sponge effect of PEI.	15
Figure 2.4 Chemical structures of the linear and branched polyethylenimine (PEI)	15
Figure 2.5 Schematic representation of different modifications of PEI	17
Figure 2.6 Schematic description of the synthesis of PEI2-GNPs	18
Figure 2.7 Cytotoxicity (Left) and gene delivery activity (Right) of degradable PEI (A) M MB-231 (B) C2C12 cells. Left: 25-KDa PEI (○), 800-Da PEI (●), polymer 1 (▽), and poly 2 (▼). Right: white bars, 25-KDa PEI; gray bars, polymer 1; black bars, polymer 2	ymeı
Figure 2.8 Localization of fluoresceinyl peptides in MDA-MB-435 tumors Fluoresceingle peptides (green) and blood vessels stained with anti-MECA-32 antibody (red) in tumor tis Nuclei were detected by DAPI staining (blue).	ssue
Figure 3.1 FTIR spectrometry of PEI800, DA and PEI-DA	59
Figure 3.2 NMR spectrometry of PEI-DA.	60

Figure 3.3 Buffer capacity of PEI800, PEI25K and PEI-DA solution at 25 $^{\circ}$ C 1 atm. 150 mM NaCl was used as the control
Figure 3.4 Evaluation of pDNA binding ability of PEI800, PEI25K and PEI-DA at different N/P ratios from 0.5 to 50. Exposure time: 400 ms. The first lane, naked DNA, was used as reference
Figure 3.5 Apparent hydrodynamic sizes (a) and zeta potentials (b) of carriers/pDNA complexes prepared by mixing PEI-DA, PEI800 and PEI25K with pDNA at various N/P ratios from 0.5 to 50 at pH 7.0 and 25 $^{\circ}$ C. For all measurements, the concentration of pDNA was fixed at 5 μ g/ml.
Figure 3.6 Cell viability of Hela cells (a) and HEK 293 cells (b) after exposing to PEI800, PEI25K and PEI-DA
Figure 3.7 Hela cell uptake of YOYO-1-labelled pDNA complexed with different gene delivery carriers: PEI800, PEI25K and PEI-DA at an N/P ratio of 10. Mock cells were used as control, and naked pDNA was used as the reference
Figure 3.8 Flow cytometry of GFP expression by various polymer/pDNA complexes prepared by PEI800, PEI25K and PEI-DA at the N/P ratio of 10 in Hela cells (a) and HEK 293 cells (b). Mock cells were used as control, and naked pDNA was used as the reference for both cells.
Figure 3.9 Transfection efficiency of naked DNA, PEI800, PEI25K and PEI-DA when delivering pDNA to Hela cells (a) and HEK 293 cells (b) at an N/P ratio of 1067
Figure 4.1 FTIR spectra of PEI800, DA, HIV-1 TAT, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 %
Figure 4.2 Buffering capacities of PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 % at 25 °C 1 atm. 150 mM NaCl was used as the control
Figure 4.3 Evaluation of nucleic acid binding and protection capability of PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 % at different N/P ratios (N/P ratios of 0/5, 1, 5, 10 and 50 from lane 1-5). Exposure time: 400 ms. The first lane, naked DNA, was used as reference.
Figure 4.4 Apparent hydrodynamic sizes (a) and zeta potentials (b) of carriers/pDNA complexes prepared by mixing PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-

TAT 10 % with pDNA at different charge ratios (N/P) of carrier to pDNA at pH 7.0 and 25
$^{\circ}\text{C}.$ For all measurements, the pDNA concentration was fixed at 5 $\mu\text{g/ml}.$ 95
Figure 4.5 Cell viability of Hela cells (a) and HEK 293 cells (b) after exposing to PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 %96
Figure 4.6 Hela cell uptake of YOYO-1-labelled pDNA complexed with different gene
delivery vectors: PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 % in Hela cells at an N/P ratio of 10. Mock cells were used as control, and naked pDNA was used
as the reference
Figure 4.7 Flow cytometry of GFP expression by various polymer/pDNA complexes prepared
by PEI800, PEI25K, PEI-DA, PEI-DA-TAT 1 % and PEI-DA-TAT 10 % at an N/P ratio of
10 in Hela cells (a) and HEK 293 cells (b). Mock cells were used as control, and naked pDNA
was used as the reference98