# Differential maternal and paternal genome effects on placental and fetal phenotype and gene expression at midgestation

#### **Ruidong Xiang**

Thesis submitted for the degree of

Doctor of Philosophy in Science



School of Animal and Veterinary Sciences
Faculty of Sciences
The University of Adelaide
Adelaide, South Australia, Australia
Februry 2014

Maternal and Paternal Effects on Fetal Development

#### **Table of Contents**

List of figures	VI
List of tables	IX
List of supporting information	X
Declaration	XII
List of publication/prepared manuscripts	XIII
Acknowledgements	XV
Abstract	XVIII
Chapter 1: Literature Analysis	1
Chapter 1	2
1.1 Introduction	2
1.2 Placenta	4
1.2.1 Role of placenta	4
1.2.2 Placentogenesis and lifetime impact	5
1.3 Skeleton	7
1.3.1 Role of skeleton	7
1.3.2 Skeletogenesis and lifetime impact	7
1.4 Skeletal muscle	9
1.4.1 Role of skeletal muscle	9
1.4.2 Myogenesis and lifetime impact	9
1.5 Transcriptome determination of phenotypic development	10
1.6 Mendelian genetic effects on placenta, musculoskeletal system and associated transcript abundance profiles	11
1.7 Non-Mendelian genetic and epigenetic factors in placenta, musculoskeletal system and associated transcript abundance profiles	13
1.7.1 Introduction to non-Mendelian genetics	13

1.7.2 Non-Mendelian genetic and epigenetic effects on placenta and musculoskeletal system	13
1.7.2.1 Cytoplasmic inheritance and sex linkage	
1.7.2.2 Genomic imprinting	14
1.7.2.3 Parent-of-origin effects	16
1.7.3 Non-Mendelian genetic and epigenetic effects on transcription profiles	20
1.8 Research aim	22
References	24
Chapter 2: Novel paternal and maternal genome effects on the placental-fetal system support both conflict-of-interest and maternal-offspring coadaptation	38
Chapter 2	41
2.1 Abstract	41
2.2 Introduction	43
2.3 Results	46
2.3.1 Proportion of variation explained by parental genomes, fetal sex and non-genetic maternal effects	46
2.3.2 Nested regression network between placental and fetal phenotype within parental genomes and fetal sex	50
2.3.3 Specific effects of Bt and Bi genomes, fetal sex and maternal weight	55
2.3.4 Parental genome and fetal sex-specific regressions between placental and fetal phenotype	
2.4 Discussion	68
2.5 Materials and methods	74
2.5.1 Animals	74
2.5.2 Placental, fetal and umbilical cord gross-morphometry	75
2.5.3 Placentome immunohistochemistry	76
2.5.4 Placentome histo-morphometry	77
2.5.5 Statistical analysis	78
Dafarancas:	Q 1

Chapter 3: Widespread differential maternal and paternal genome effects on fetal bone ohenotype at midgestation	86
Chapter 3	90
3.1 Abstract	90
3.2 Introduction	92
3.3 Material and methods	94
3.3.1 Animals	94
3.3.2 Bone parameters	96
3.3.3 Principal component analysis	96
3.3.4 General linear models	97
3.4 Results	98
3.4.1 Principal components	98
3.4.2 Proportion of variation explained by parental genomes, fetal sex and non- genetic maternal effects	99
3.4.3 Specific effects of defined maternal and paternal genomes, fetal sex and non-genetic maternal factors	101
3.5 Discussion	104
3.5.1 Fetal bone phenotype at midgestation	104
3.5.2 Effects of maternal and paternal genomes	104
3.5.3 Fetal sex and non-genetic maternal effects	107
References	109
Chapter 4: Maternal and paternal genomes differentially affect myofibre characteristics and muscle weights of bovine fetuses at midgestation	117
Chapter 4	121
4.1 Abstract	121
4.2 Introduction	123
4.3 Pagults	126

4.3.1 Proportion of variation explained by parental genomes, fetal sex and non- genetic effects	126
4.3.2 Specific effects of Bt and Bi genomes, fetal sex and maternal weight	130
4.3.3 Expression of the H19 lincRNA	137
4.4 Discussion	138
4.5 Materials and Methods.	146
4.5.1 Cattle and fetuses	146
4.5.2 Muscle dissection and weights	147
4.5.3 Muscle immunohistochemistry	147
4.5.4 Myofibre Classification and Morphometry	148
4.5.5 Expression of <i>H19</i> in skeletal muscle	149
4.5.6 Statistical estimation of effects and means	150
References	154
Chapter 5: Differential maternal and paternal genome effects on fetal liver transcriptome expression and coexpression network at midgestation	162
Chapter 5	163
5.1 Abstract	163
5.2 Introduction.	165
5.3 Materials and methods	167
5.3.1 Animals and tissue preparation	167
5.3.2 RNA extraction	167
5.3.3 Microarray procedure	168
5.3.4 Data analysis	169
5.4 Results	
5.4 Results	172

5.4.3 Transcript coexpression differentiated by parental genome effects	184
5.5 Discussion	188
5.5.1 Complex maternal genome effects	188
5.5.2 Paternal genome effects	191
5.5.3 Fetal sex effects	192
References	194
Chapter 6: General discussion	199
Chapter 6	200
General Discussion	200
6.1 Introduction and overview	200
6.2 Parental genome effects	202
6.2.1 Complex maternal genome effects	202
6.2.2 Parental genome and evolutionary basis of parent-of-origin effects	205
6.2.3 Independent effects and interaction effects of parental genomes	208
6.3 Fetal sex effects	211
6.4 Non-genetic maternal effects	212
6.5 Phenotypic and transcriptional modules	212
6.6 General conclusions	215
References	217
Supporting figures and tables	225
Supporting figures:	226
Supporting tables:	240

## List of figures

Figure 1.1 Number of imprinted genes reported to date.	16
Figure 1.2. All possible phenotypic patterns of genomic imprinting according to Wolf et al. (2008).	19
Figure 2.1. Relative contributions of parental genomes, fetal sex and non-genetic maternal effects to explained variation in placental and fetal phenotype	48
Figure 2.2. Relative contributions of maternal and paternal genome to genetic variation in placental and fetal phenotype.	50
Figure 2.3. Regression network for total and maternal placenta weight determined by parental genomes and/or fetal sex.	51
Figure 2.4. Regression network for fetal placenta and fetus weight determined by parental genomes and/or fetal sex.	52
Figure 2.5. Regression network for fetal organ weights determined by parental genomes and/or fetal sex.	54
Figure 2.6. Specific effects of maternal genomes, paternal genomes and fetal sex on placenta and fetus weight at midgestation.	56
Figure 2.7. Specific effects of maternal genomes, paternal genomes and fetal sex on fetal organ and fluids weight at midgestation.	57
Figure 2.8. Specific effects of maternal genomes, paternal genomes and fetal sex on umbilical cord phenotype at midgestation.	58
Figure 2.9. Specific effects of maternal genomes, paternal genomes and fetal sex on placental/umbilical cord efficiency at midgestation.	59
Figure 2.10. Specific effects of maternal genomes, paternal genomes and fetal sex on histomorphological placental phenotype at midgestation.	60
Figure 2.11. Specific linear regressions of gross-placental/fetal phenotype on umbilical cord/histo-placental phenotype nested within maternal genomes.	63
Figure 2.12. Specific regressions of fetal and organ phenotype on umbilical cord/fetal fluid phenotype nested within paternal genomes.	65
Figure 2.13. Specific regressions of gross-placental/fetal phenotype on umbilical cord/histo-placental phenotype nested within fetal sex.	67
Figure 3.1. Post conception maternal daily weight gain and final weight for <i>Bos taurus</i> taurus and <i>Bos taurus indicus</i> dams	95

Figure 3.2. Relative contributions of genetic and non-genetic factors to variation explained in principal components for bone weight and geometry parameters.	100
Figure 3.3. Specific effects of maternal genome, paternal genome and fetal sex on identified and extracted principal components (PC) of measured fetal bone geometry and weight parameters.	102
Figure 3.4. Effects of non-genetic effects of maternal daily weight gain nested within maternal genomes and fetal sex on PC3/limb elongation	103
Figure 4.1 Relative contributions of parental genomes, fetal sex and non-genetic maternal effects to explained variation in fetal myofibre characteristics, absolute and relative muscle weights, and <i>H19</i> transcript abundance.	127
Figure 4.2. Relative contributions of maternal and paternal genome to genetic variation in fetal myofibre characteristics, absolute and relative muscle weights, and H19 transcript abundance.	129
Figure 4.3. Specific effects of maternal genomes, paternal genomes and fetal sex on fetal myofibre characteristics of <i>M. semitendinosus</i> at midgestation.	131
Figure 4.4. Specific effects of maternal genomes, paternal genomes and fetal sex on fetal absolute muscle weights at midgestation.	133
Figure 4.5. Effects of final maternal weight nested within maternal genomes on fetal absolute muscle weights at midgestation.	134
Figure 4.6. Specific effects of maternal genomes, paternal genomes and fetal sex on fetal relative muscle weights at midgestation.	136
Figure 4.7. Effects of interaction of maternal and paternal genomes, fetal sex and final maternal weight nested within maternal genetics on <i>H19</i> transcript abundance in fetal <i>M. semitendinosus</i> at midgestation.	137
Figure 4.8. Regressions of fetal muscle mass at midgestation on <i>H19</i> transcript abundance	138
Figure 5.1. Significant parental genomes and fetal sex effects on fetal liver weight at midgestation.	174
Figure 5.2. Significant parental genome and fetal sex effects on mRNA transcript abundances in fetal liver at midgestation.	176
Figure 5.3. Significant parental genome effects on non-coding RNA transcript abundances in fetal liver at midgestation.	182
Figure 5.4. Biological pathways identified for differentially expressed transcripts in fetal liver at midgestation.	184
Figure 5.5. Transcript coexpression networks differentiated by maternal and paternal genome effects in fetal liver at midgestation.	186

Figure 5.6. Pearson correlation regressions of microarray expression intensity values	
between significant miRNAs and predicted target mRNAs.	.187

### List of tables

Table 1.1. Adult metabolic consequences resulting from altered placental phenotypes	6
Table 1.2. Variation in adult bone traits explained by birthweight.	8
Table 1.3. Heritability $(h^2)$ of placental and postnatal musculoskeletal traits	13
Table 2.1. Summary of the final general models (type III sums of squares) for placental and fetal traits with adjusted $R^2$ values and significance levels ( $p$ -values) of models and variables. Only $p$ -values for factors, interactions and nested effects retained in the final model are shown.	47
Table 3.1. Summary and interpretation of results of principal component (PC) analysis of 51 bone weight and geometry parameters.	99
Table 3.2. Summary of the final general linear models (type III sums of squares) for principal components of bone weight and geometry parameters, with adjusted $R^2$ values and significance levels ( $P$ -values) of models and variables. Only $P$ -values for factors, interactions and nested effects retained in the final model are shown. The model for PC6 was not significant ( $P$ >0.05)	101
Table 4.1. Summary of the final general models (type III sums of squares) for myofibre characteristics, muscle weight parameters and H19 gene expression with adjusted $R^2$ values and significance levels ( $P$ -values) of models and variables.	128
Table 5.1. Annotation of 24 mRNA transcripts for significant maternal genome effects, with log <sub>2</sub> fold change (Log FC) from Bt ( <i>Bos taurus taurus</i> , Angus) to Bi ( <i>Bos taurus indicus</i> , Brahman) and false discovery rate (FDR) adjusted <i>P</i> -values.	177
Table 5.2. Annotation of 47 mRNA transcripts for significant paternal genome effects, with log <sub>2</sub> fold change (Log FC) from Bt ( <i>Bos taurus taurus</i> , Angus) to Bi ( <i>Bos taurus indicus</i> , Brahman) and false discovery rate (FDR) adjusted <i>P</i> -values.	178
Table 5.3. Annotation of 26 mRNA transcripts for significant fetal sex effects, with log <sub>2</sub> fold change (Log FC) from M (male) to F (female) and false discovery rate (FDR) adjusted <i>P</i> -values.	180
Table 5.4. Annotation of 10 miRNA transcripts for significant maternal genome effects, with log <sub>2</sub> fold change (Log FC) from Bt ( <i>Bos taurus taurus</i> , Angus) to Bi ( <i>Bos taurus indicus</i> , Brahman) and false discovery rate (FDR) adjusted <i>P</i> -values	183
Table 5.5. Annotation of 5 miRNA transcripts for significant paternal genome effects, with log <sub>2</sub> fold change (Log FC) from Bt ( <i>Bos taurus taurus</i> , Angus) to Bi ( <i>Bos taurus indicus</i> , Brahman) and false discovery rate (FDR) adjusted <i>P</i> -values	183

## List of supporting information

#### **Supporting figures:**

Figure S2.1. Effects of final maternal weight or daily weight gain nested within maternal genomes on fetal and organ weights weights at midgestation.	226
Figure S2.2. Effects of final maternal weight or daily weight gain nested within maternal genomes on umbilical artery/vein diameter at midgestation.	227
Figure S2.3. Effects of final maternal weight or daily weight gain nested within maternal genomes on gross-morphological placentome phenotype at midgestation.	228
Figure S2.4. Effects of final maternal weight or daily weight gain nested within maternal genomes on histo-morphological placental phenotype at midgestation.	229
Figure S2.5. Effects of final maternal weight or daily weight gain nested within maternal genomes on histo-morphological placental phenotype at midgestation continued	230
Figure S2.6. Specific regressions of gross placental/fetal phenotype on maternal barrier thickness nested within maternal and paternal genome interaction.	231
Figure S2.7. Specific regressions of gross placental/fetal phenotype on umbilical artery/vein diameter nested within paternal genome and fetal sex interaction	233
Figure S2.8. Daily weight gain and final weight for <i>Bos taurus taurus</i> and <i>Bos taurus indicus</i> dams.	234
Figure S2.9. Example of immunohistochemical staining for fetal placentome at midgestation.	235
Figure S3.1. Example of set of Day 153 fetal bones with measurements of bone geometry parameters indicated.	236
Figure S4.1. Example of immunohistochemical staining for fetal slow and fast myofibres in <i>M. semitendinosus</i> at midgestation.	237
Figure S4.2. Fetal carcass weights for the four different combinations of maternal and paternal genomes and fetal sex at midgeststion	238
Figure S4.3. Quadratic effects of final maternal weight nested within maternal genomes on absolute weight of fetal <i>M. quadriceps femoris</i> at midgestation.	239
Figure S4.4. Daily weight gain and final weight for <i>Bos taurus taurus</i> and <i>Bos taurus indicus</i> dams	239

#### **Supporting tables:**

Table S2.1 Summary of adjusted $R^2$ value, significance levels of models and nested regressions between gross-morphological placental and fetal traits (response variables) and umbilical traits and histo-morphological placental traits (explanatory variables). Only $P$ -values for significant nested regressions retained in the final model are shown	.240
Table S2.2. Summary for distribution of maternal and paternal genomes and sex of fetuses	246
Table S3.1. Summary for distribution of maternal and paternal genomes and sex of fetuses	247
Table S3.2. Summary of measurements of bone weight and geometry parameters	248
Table S4.1. Summary for distribution of maternal and paternal genomes and sex of fetuses	249
Table S4.2. Primer sequences used for quantitative real time polymerase chain reaction of <i>H19</i> and housekeeping genes.	249
Table S5.1. Summary of pathway analysis for differentially expressed transcripts, with enrichment score and modified Fisher Exact <i>P</i> -value for each pathway and corresponding transcripts information.	.250
Table S5.2. Summary of transcript coexpression (CE) network one identified between significant mRNA and non-coding RNA transcripts for microarray ANOVA	.252
Table S5.3. Summary of transcript coexpression (CE) network two identified between significant mRNA and non-coding RNA transcripts for microarray ANOVA	254

**Declaration** 

I certify that this work contains no material which has been accepted for the award of any other

degree or diploma in any university or other tertiary institution and, to the best of my knowledge

and belief, contains no material previously published or written by another person, except where

due reference has been made in the text. In addition, I certify that no part of this work will, in the

future, be used in a submission for any other degree or diploma in any university or other tertiary

institution without the prior approval of the University of Adelaide and where applicable, any

partner institution responsible for the join-award of this degree.

I give consent to this copy of my thesis when deposited in the University Library, being made

available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis (as

listed below) resides with the copyright holder(s) of those works. I also give permission for the

digital version of my thesis to be made available on the web, via the University's digital research

repository, the Library catalogue and also through web search engines, unless permission has

been granted by the University to restrict access for a period of time.

February 2014

XII

#### List of publication/prepared manuscripts

# Chapter 2: Novel paternal and maternal genome effects on the placental-fetal system support both conflict-of-interest and maternal-offspring coadaptation

Ruidong Xiang<sup>1,2,5</sup>, Consuelo A. Estrella<sup>1,2,5</sup>, Carolyn J. Fitzsimmons<sup>1,2</sup>, Zbigniew A. Kruk<sup>1,2</sup>, Dana A. Thomsen<sup>1,2</sup>, David L. Rutley<sup>1,3</sup>, Claire T. Roberts<sup>1,4</sup>, Stefan Hiendleder<sup>1,2\*</sup>

#### Intended for submission to PLoS Biology.

# Chapter 3: Widespread differential maternal and paternal genome effects on fetal bone phenotype at midgestation

Ruidong Xiang<sup>1,2</sup>, Tanja Eindorf<sup>2</sup>, Madeleine Gugger<sup>2</sup>, Carolyn J. Fitzsimmons<sup>2</sup>, Zbigniew A. Kruk<sup>2</sup>, Wayne Pitchford<sup>3</sup>, Dana A. Thomsen<sup>2</sup>, Gail I. Anderson<sup>4</sup>, Brian M. Burns<sup>5</sup>, David L. Rutley<sup>6</sup>, Stefan Hiendleder<sup>1,2\*</sup>

#### Submitted to Journal of Bone and Mineral Research.

<sup>&</sup>lt;sup>1</sup> JS Davies Epigenetics and Genetics Group, School of Animal and Veterinary Sciences, Roseworthy Campus, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>2</sup> Robinson Institute, Research Centre for Reproductive Health, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>3</sup> Biometry Hub, Waite Campus, The University of Adelaide, South Australia, Australia

<sup>&</sup>lt;sup>4</sup> School of Paediatrics and Reproductive Health, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>5</sup> These authors contributed equally to this work.

<sup>\*</sup> Corresponding author

<sup>1</sup> JS Davies Epigenetics and Genetics Group, School of Animal and Veterinary Sciences, Roseworthy Campus, The University of Adelaide, South Australia, Australia.

<sup>2</sup> Robinson Institute, Research Centre for Reproductive Health, The University of Adelaide, South Australia, Australia.

<sup>3</sup> School of Animal and Veterinary Sciences, Roseworthy Campus, The University of Adelaide, South Australia, Australia.

<sup>4</sup> Veterinary Health Centre, School of Animal and Veterinary Sciences, Roseworthy Campus, The University of Adelaide, South Australia, Australia.

<sup>5</sup> The University of Queensland, Centre for Animal Science, Rockhampton, Queensland, Australia.

<sup>6</sup> Biometry Hub, Waite Campus, The University of Adelaide, South Australia, Australia

<sup>\*</sup> Corresponding author.

## Chapter 4: Maternal and paternal genomes differentially affect myofibre characteristics and muscle weights of bovine fetuses at midgestation

Ruidong Xiang<sup>1,2</sup>, Mani Ghanipoor-Samami<sup>1,2</sup>, William H. Johns<sup>3</sup>, Tanja Eindorf<sup>1</sup>, David L. Rutley<sup>1</sup>, Zbigniew A. Kruk<sup>1</sup>, Carolyn J. Fitzsimmons<sup>1</sup>, Dana A. Thomsen<sup>1,2</sup>, Claire T. Roberts<sup>2,4</sup>, Brian M. Burns<sup>5</sup>, Gail I. Anderson<sup>1</sup>, Paul L. Greenwood<sup>3</sup>, Stefan Hiendleder<sup>1,2\*</sup>

Published in 2013, PLoS ONE 8(1): e53402.

<sup>&</sup>lt;sup>1</sup> J.S. Davies Epigenetics and Genetics Group, School of Animal and Veterinary Sciences, Roseworthy Campus, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>2</sup> Robinson Institute, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>3</sup> NSW Department of Primary Industries, Beef Industry Centre, Trevenna Rd, University of New England, Armidale, New South Wales, Australia.

<sup>&</sup>lt;sup>4</sup> School of Paediatrics and Reproductive Health, The University of Adelaide, South Australia, Australia.

<sup>&</sup>lt;sup>5</sup> The University of Queensland, Centre for Animal Science, Queensland Alliance for Agriculture and Food Innovation, Rockhampton, Queensland, Australia.

<sup>\*</sup> Corresponding author.

#### Acknowledgements

It seems like only recently that Prof. Stefan Hiendleder, my principlal supervisor, showed me around the lab for the first time and I had no idea what he was talking about. However, I am now completing my PhD and I realise that one's PhD work actually includes contributions from many people, thus, I would like to acknowledge their contributions as follows:

Firstly, I would like to sinerely thank my principal supervisor, Prof. Stefan Hiendleder, for his high standard of academic professionalim, exceptional academic mentorship and a countless amount of selfless help that immensely contributed to my PhD. I am also extremely thankful to his implantation of southern German thoroughness and scientific 'purism' into my work and psyche that I will carry with me for life. Specifically, his meticulous attitude towards graphical presentation has significantly raised my esthetic taste in science. His maddening attention to detail finally drove me to learn to insert non breaking spaces between equal signs and *P*-values. His high standard of beer appreciation has extended my understanding of the application of biological principles. It has been an honour and privilege to have Prof. Stefan Hiendleder as my Ph.D supervisor.

I would like to thank my co-supervisor, Prof. Gail Anderson, for her great help with my bone experiment and a lap steel guitar performance that calmed my nerves.

I thank my external supervisor, Dr. Paul Greenwood, for his significant contribution to my muscle related experiment and work conducted at the University of New England, Armidale. I am specifically grateful to his kind introduction to the method of using French bean cuisine and chilli chocolate with wine to cope with freezing winter temperature in Armidale. This advice ensured successful completion of my experiment.

I am very grateful to my co-supervisor Dr. David Rutley for his encouragement and motivating me to delve deeply into statistical analysis. Indeed, I now consider statistics as a potential career path. I sincerely enjoyed the moments that we discussed "simple" statistical questions to the extent that he asked me to leave his office, or other people came to his office to ask us to stop yelling at each other.

I would like to thank Dr. Dana Thomsen for her incredible and significant contribution to my writing skills. Her unbelievable patience with my writing, and persistent refusal to edit my work using computer track-changes, vastly improved my understanding of authentic English and Australian writing.

I am also thankful to Prof. Claire Roberts for her kind help in my placenta work and paper, and for setting the mysterious thermostats each time we met in the North Wing of the Medical School.

I would also like to thank: Dr. Tanja Eindorf for her direct help with my muscle and bone experiment, during which we proved that a slowcooker can be an important experimental instrument; Dr. Bill Johns for his direct help with the muscle fibre staining experiment, during which we found that a good quality morning tea at 11:00am is critical for a whole day experiment; Madeleine Gugger for her direct help in my bone experiment, during which we found that jelly beans run out faster than clean bench protection during intensive work; Consuelo Estrella for her direct help in my placenta work, during which we found that eating cakes in the tearoom without talking to the sales people who provided them is technically doable; Ali Javadmanesh and Mani Ghanipoor Samami, the Doctors-to-be, for their help in the lab, during which we found that a lunch break is not as long as we think.

I thank my parents for their significant life and emotional support during my PhD. I would like to specially thank Ms. Lesley Menzel for her selfless and remarkable direct help with my social life that exposed me to authentic Australian culture and slang, and for her help with my life difficulties. It is a privilege to have her as a friend in Australia and her contribution to my PhD is countless. I want to specially thank Dr. Rugang Tian for his vast help in life matters and for his authentic Chinese cooking skills that absolutely cured my homesickness. Also I feel so grateful to Dr. Tai-yuan Chen's company and encouragement during those long and dark nights. I also want to thank Dr. Bo Li and Dr. You Li for sharing my good and bad times. I specially thank Ms. Linlin for her asisstance in high resolution graph production and Dr. Jingjing Wei for her generous support during my PhD and insightful suggestions for my career path.

I am very grateful for the J.S. Davies Bequest project funding. My sincere thanks are also extended to the China Scholarship Council and The University of Adelaide for providing the PhD scholarship. I thank the University of Adelaide for providing the opportunity for this project and for the support of university staff, in particular the Graduate Centre.

Lastly, I would like to sincerely thank those people or events that consciously or unconsciously provided opportunities for me to refine survival skills under extreme conditions during my PhD in Australia. The results have demonstrated that this guy is nothing if not adaptable and he is up for a new fun chapter.

Cheers.

#### **Abstract**

Lifelong development is largely programmed prenatally. Genetic and epigenetic factors, such as mitochondrial (mt) DNA variation and parent-of-origin effects, significantly contribute to variation in important prenatal phenotypes that determine lifetime development, including placenta and fetal musculoskeletal system. Such effects initially impact on transcriptome expression levels and eventually give rise to altered phenotypic traits. However, data regarding the overall magnitude and specificity of maternal and paternal genome effects in mammalian prenatal development is lacking.

The present study aimed to dissect and quantify differential maternal and paternal genome effects on specific placental and fetal traits, and associated transcriptomic events which drive prenatal development. A large bovine fetal resource (n=73), consisting of both purebreds and reciprocal hybrids with *Bos taurus taurus* (Angus) and *Bos taurus indicus* (Brahman) (epi)genetics, was used in this study. We examined 41 gross- and histo-morphological placental and fetal traits, 51 fetal bone weight and geometry parameters, and 22 myofibre characteristics and muscle mass parameters using morphometrical and/or immunohistochemical methods. Expression of the long non-coding RNA H19 in fetal muscle was determined by real time quantitative PCR. Profiles of mRNA and microRNA expression were obtained with microarrays that contained 24,027 and 13,133 mammalian probe sets, respectively, to assess transcript abundances in fetal liver. Phenotypic data were analysed by Analysis of Variance (ANOVA) using general linear models with nested effects and transcriptome data were analysed with microarray ANOVA procedures.

The analyses identified 49 significant placental and fetal traits, including five principal components representing 51 bone parameters, and H19 gene expression levels in muscle, with

ANOVA model significance levels (P) ranging from  $3\times10^{-2}$ - $9\times10^{-17}$ . We showed that parental genomes contributed to the largest proportion of variation explained by linear models for a majority of placental and fetal traits. Fetal sex was the next most significant factor to explain variation in these traits and non-genetic maternal effects, such as post-conception weight gain and final maternal weight, explained the least amount of variation. Significant effects of the maternal genome ( $P<5\times10^{-2}$ - $5\times10^{-13}$ ) predominantly contributed to genetic variation in:

- (i) Gross- and histo-morphological placental traits and fetal organ weights (59.6–99.9%,); (ii) most extracted principle components (PCs) representing bone weight and geometry traits, including PC1/bone mass (74%), PC3/limb elongation (73%), PC4/flat bone elongation (74%) and PC5/axial skeletal growth (97%) and (iii) most myofibre characteristics including fast myofibre cross-sectional area (CSA, 93%), total cell CSA (82%), absolute mass of studied muscles (59-88%) and H19 transcript abundance in fetal muscle (76%). Conversely, significant paternal genome (P<4×10<sup>-2</sup>-7×10<sup>-8</sup>) predominantly contributed to genetic variation in:
- (i) Fetal fluids weight (73%), umbilical cord weight and length (73%), maternal placenta (70%) and umbilical cord (83%) efficiencies; (ii) PC2/limb ossification (95%) and (iii) Relative mass of studied muscles to fetal weight (54-97%).

Further, using nested effects in ANOVA, we found that maternal genome strongly determined regressions between placental weights and umbilical cord traits (P<4×10<sup>-2</sup>-2×10<sup>-6</sup>), whereas paternal genome and/or fetal sex determined regressions between weight of fetus, fetal organ and fetal fluid s and umbilical cord traits (P<5×10<sup>-2</sup>-10×10<sup>-8</sup>).

For fetal liver transcription profiles, maternal genome strongly affected expression levels of:

(i) Twenty-four mRNA transcripts (false discovery rate, FDR adjusted  $P < 4 \times 10^{-2} - 10 \times 10^{-6}$ ), 13 of which were located in the mt genome and (ii) ten autosomal non-coding RNA transcripts

including mammalian *SNORD113-9*, small nucleolar (sno)RNA, *MIR187* and *MIR1973* microRNA (FDR adjusted  $P < 5 \times 10^{-2} - 8 \times 10^{-3}$ ).

Paternal genome moderately affected expression levels of:

(i) Forty-seven autosomal mRNA transcripts (FDR adjusted,  $P < 5 \times 10^{-2} - 4 \times 10^{-2}$ ) (ii) MIR184 microRNA transcripts in five mammalian species (FDR adjusted,  $P < 5 \times 10^{-2} - 4 \times 10^{-2}$ ).

Two significant coexpression networks, between 86 significant mRNAs and non-coding RNA transcripts, were also identified for differential maternal and paternal genome effects.

Our results show, for the first time, that a wide range of phenotypic and molecular traits within the placental-fetal system are affected by differential maternal and paternal genome and fetal sex effects. Identified differential maternal and paternal genome effects on specific placental and fetal traits are consistent with expression patterns of parent-of-origin effects predicted by both conflict-of-interest and maternal-offspring coadapdation hypotheses, thereby providing important insights to accommodate both hypotheses that explain the evolutionary basis of genomic imprinting effects. Observed complex, and predominantly maternal genome, effects are suggested to result from interaction between epigenetic factors from nuclear and mt genomes via RNA interference. This is further evidence for complex epigenetic crosstalk and coordination that contributes to mammalian prenatal development. Identified morphological and transcriptional modules within the placental-fetal system help to provide a new level of understanding prenatal development, i.e., systematic integration of omics data. Detailed molecular profiles of all core tissues and organs are now required to elucidate genetic, epigenetic and non-genetic components and interactions that control variation in placental and fetal phenotype. Future studies linking genome and epigenome with phenome data covering the

complete placental-fetal system will provide a new multi-layer picture of understanding coordination for molecular and phenotypic events driving mammalian prenatal development.