# Evaluation of the effects of AtCIPK16 expression on the salt tolerance of barley and wheat

## **Emily Laurina Thoday-Kennedy**

A thesis submitted for the degree of
Master of Philosophy
School of Agriculture, Food and Wine
Faculty of Sciences



May 2016

## **Table of Contents**

Table of Contents	. <b></b> i
List of Figures	iii
List of Tables	v
List of Abbreviations	vi
Abstract	х
Declaration	xi
Acknowledgments	xii
Chapter 1: Literature review	1
1.1 A global problem	1
1.2 Salinity	1
1.2.1 Salt-affected soils	1
1.2.2 Global salinity	
1.2.3 Salt-affected Australia	2
1.3 How salt affects plants	3
1.3.1 Sodium toxicity	4
1.4 Salt tolerance mechanisms	4
1.4.1 Osmotic tolerance	4
1.4.2 Ionic tolerance	4
1.5 CBLs and CIPKs	6
1.5.1 Ca <sup>2+</sup> signalling in plants	6
1.5.2 Calcineurin B-like proteins (CBLs)	
1.5.3 Calcineurin B-like Interacting Proteins Kinases (CIPKs)	8
1.5.4 CBL-CIPK signalling pathways	10
1.5.5 Examples of CBL-CIPK pathways	11
1.6 AtCIPK16	
1.6.1 Arabidopsis thaliana Calcineurin B-like Interacting Protein Kinase 16	
1.6.2 Other CIPK16s	14
1.7 Research Aims	
Chapter 2: Evaluation of 35S:AtCIPK16 Golden Promise barley lines under field conditions	
2013 & 2014	
2.1 Introduction	
2.2 Materials and Methods	
2.2.1 Environmental characterisation of field trial site	
2.2.2 Plant material	
2.2.3 Field trial of transgenic barley	
2.2.4 DNA extraction and genotyping analysis	
2.2.5 Soil analysis of field trial plots	
2.2.6 Ion analysis of leaf tissue	
2.3 Results	
2.3.1 Environmental characterisation of field trial site	
2.3.2 Transgenic <i>AtCIPK16</i> barley show variations in plant growth	
2.3.3 Transgenic <i>AtCIPK16</i> expressing barley lines show possible Na <sup>+</sup> exclusion	
2.3.4 Expression of <i>AtCIPK16</i> in barley does not improve yield	
2.4 Discussion	
2.4.1 Transgenic <i>AtCIPK16</i> barley has increased Na <sup>+</sup> and CI <sup>-</sup> exclusion	30
2.4.2 Na <sup>+</sup> and Cl <sup>-</sup> exclusion does not translate to improved biomass or yield in transgenic	24
AtCIPK16 lines	
2.4.3 Variation in results between years linked to environmental factors	
2.5 Conclusions & Future directions	
3.1 Introduction	
3.2 Materials and Methods	30

3.2.1 Plant material	37
3.2.2 Growth conditions	
3.2.3 DNA extraction and genotyping analysis	38
3.2.4 RNA extraction and gene expression analysis	39
3.2.5 Ion analysis of leaf and root tissue	
3.3 Results	
3.3.1 Gene presence and expression analysis of AtCIPK16 transgenic lines	
3.3.2 Transgenic AtCIPK16 lines have varied biomass production	
3.3.3 Transgenic AtCIPK16 lines have varying responses in leaf ion accumulation	
3.3.4 Transgenic AtCIPK16 lines show varied root ion accumulation trends	
3.4 Discussion	
3.4.1 Response of Gladius wheat to NaCl treatment	
3.4.2 One transgenic line, CIPK16-2-2, demonstrates a Na <sup>+</sup> and Cl <sup>-</sup> exclusion phenotype.	
3.4.3 Disruption of transgene expression: hypothesised reason for lack of phenotype	
3.5 Conclusions & Future directions	
Chapter 4: Determination of whether the presence/absence of TATA-box in the AtCIPK16 promoter is responsible for the AtCIPK16 expression differences observed between	)
Arabidopsis ecotypes	61
4.1 Introduction	
4.2 Materials and Methods	
4.2.1 Analysis of promoter regions to identify mutation sites	
4.2.2 Introducing point mutations by PCR mutagenesis	
4.2.3 Restriction digest and DNA ligation reactions	
4.2.4 Generation of amplicon C – pCR8 Gateway® vectors	
4.2.5 Further steps needed to transform final destination vectors into Arabidopsis	
4.3 Results	
4.3.1 Analysis of AtCIPK16 promoters to introduce point mutations and design primers	69
4.3.2. Successful creation of amplicons A, B and C containing the desired point mutation f	for
both alleles	
4.3.3 Creation of pCR8 vector with full AtCIPK16 promoter with point mutation	
4.4 Discussion	
4.4.1 Difficulties in plasmid construction	
4.5 Future work	_
Chapter 5: General Discussion	
5.1 Review of thesis aims	
5.2 Summary of main findings	
5.3 Implications of thesis findings	
5.3.1 Benefits of AtCIPK16 expression in barley and wheat may depend on environment.	
5.3.2 Role of CIPK16 in salt tolerance	
·	
5.4 Future Research	
5.4.2 Further characterisation of transgenic <i>AtCIPK16</i> wheat lines	
5.4.3 What is the AtCIPK16 network pathway in wheat and barley?	
5.4.4 AtCIPK16 expression: which promoter to use?	
5.5 Concluding Remarks	
Chapter 6: Appendices	
Appendix 1	
Appendix 2	
Appendix 3	
References	109

# List of Figures

<b>Figure 1.1:</b> Map showing the regions of Australia affected or potentially affected by transient (yellow)	
and dryland (red) salinity	3
Figure 1.2: General structure of a calcineurin B-like protein (CBL)	7
Figure 1.3: Overall structure of a CIPK showing the N-terminus serine/threonine kinase domain, with	
the activation loop (horizontal lines) and the C-terminus regulatory domain	9
Figure 1.4: Sequence alignment of the region of interest of the AtCIPK16 promoter and gene13	3
Figure 2.1: EM38 map of the field trial site in Kunjin, WA (83 m length × 32 m wide) showing the	
apparent electrical conductivity (EC <sub>a</sub> )17	7
Figure 2.2: Average rainfall (mm) and maximum temperature (°C) at Corrigin, Western Australia for	
the year 2013 and 2014	3
Figure 2.3: Electrophoresis gel showing presence of the native HvVRT2 gene and the AtCIPK16	
transgene in extracted gDNA from wildtype, null segregant and three AtCIPK16 expressing	
barley lines grown at Kunjin, WA23	3
Figure 2.4: Digital images of wildtype and transgenic AtCIPK16 expressing barley plots displaying the	,
range of plant densities in both low and high salt trial sites at Kunjin, Western Australia in 2014.	
$2^{2}$	
Figure 2.5: Shoot biomass and tiller number of wildtype, null segregant and transgenic AtCIPK16	
expressing barley grown at Kunjin, Western Australia25	5
Figure 2.6: Na+, K+ and Cl- concentration and Na+/K+ ratio of wildtype, null segregant and transgenic	
AtCIPK16 barley grown at Kunjin, WA26	
Figure 2.7: Grain yield per plants parameters of wildtype and transgenic AtCIPK16 expressing barley	
grown at Kunjin, Western Australia28	_
Figure 2.8: Grain yield per plot for wildtype and transgenic AtCIPK16 expressing barley lines grown a	t
Kunjin, Western Australia29	
Figure 3.1: Electrophoresis gel showing representative results of genotyping and expression for null	
segregants and three transgenic AtCIPK16 wheat lines	2
Figure 3.2: Photographs of null segregant and three transgenic <i>AtCIPK16</i> wheat lines at 24 days	
grown in 80 L flood-drain hydroponic systems under different salt treatments43	3
Figure 3.3: Whole plant biomass measurements and tiller number of null segregant and three	
transgenic AtCIPK16 wheat lines grown in hydroponic experiments45	5
Figure 3.4: Relative salt tolerance of null segregant and three transgenic AtCIPK16 wheat lines grown	n
under hydroponic experiments46	3
Figure 3.5: Leaf Na+ and Cl- concentration of null segregant and three transgenic AtCIPK16 wheat	
lines grown in hydroponic experiments48	3
Figure 3.6: Leaf K+ concentration of null segregant and three transgenic AtCIPK16 wheat lines grown	1
in hydroponic experiments49	
Figure 3.7: Root Na+, Cl- and K+ concentration of null segregant and three transgenic AtCIPK16	
wheat lines grown in hydroponic experiments5	1
Figure 4.1: Flow diagram outlining the methods undertaken to perform site directed mutagenesis by	
PCR on a reporter construct plasmid	
Figure 4.2: Sequence of the region of the AtCIPK16 promoter in the pCR8 vector and the primers	
involved in the site directed mutagenesis7	1
Figure 4.3: Electrophoresis gel and chromatograph with sequence alignment of amplicons A and B	
from both Shahdara and Bay-0 alleles containing the desired point mutations72	2

igure 4.4: Electrophoresis gel and chromatograph with sequence alignment of amplicon C from b	
Shahdara and Bay-0 alleles containing the desired point mutations	73
<b>igure 4.5:</b> Electrophoresis gel of failed double restriction enzyme digest of Bay-0 and Shahdara	
amplicon Cs	73
igure 4.6: Electrophoresis gel and chromatograph with sequence alignment of amplicon C in pCI	
vector for both Shahdara and Bay-0 alleles containing the desired point mutations	
igure 4.7: Electrophoresis gels of double restriction enzyme digests and results of gel purification	
bands excised from the gel of amplicon Cs in pCR8 vectors and original promoters in pCR8	76

## **List of Tables**

Table 2.1: Fertilisers applied during 2013 and 2014 field at Kunjin, WA	9
Table 2.2: Herbicides, fungicides and insecticides applied during 2013 and 2014 field trials at Kunjin,	
WA1	19
Table 3.1: Components and final concentrations in 80 L hydroponic systems of the standard ACPFG	
growth solution3	38
<b>Table 3.2:</b> Details of gene specific primers and PCR conditions used for the amplification of gDNA and/or cDNA from leaf tissue samples of null segregant and three independent <i>AtCIPK16</i>	
transgenic wheat lines4	11
<b>Table 3.3:</b> Comparison of mean results for biomass and leaf ion concentration for each sibling transgenic line grown in all three hydroponic experiments to the respective null segregants in the	ie 52
<b>Table 3.4:</b> Comparison of mean results for root ion concentration for each sibling transgenic line grown in all three hydroponic experiments to the respective null segregants in the same experiment.	52
<b>Table 4.1:</b> Description of primers designed for site directed mutagenesis of the <i>AtCIPK16</i> promoter by	y 70

### List of Abbreviations

% percentage # number × times

°C degrees Celsius

R registered trademark

-1 per

-ve negative
+ve positive

µL microliter(s)

µmoles micromole(s)

µS microSiemens

3' three prime, of nucleic acid sequence 35S promoter of cauliflower mosaic virus 35S

3D three dimensional

5' five prime, of nucleic acid sequence

aa amino acid ABA abscisic acid

ABARES Australian Bureau of Agricultural and Resource Economics and Sciences

ACPFG Australian Centre for Plant Functional Genomics

AGRF Australian Genome Research Facility

Agrobacterium Agrobacterium tumefaciens

AKT Arabidopsis potassium transporter

At Arabidopsis thaliana
ANOVA analysis of variance

AVP1 Arabidopsis vacuolar pyrophosphatase

Bay-0 Arabidopsis ecotype Bayreuth-0
BLAST basic local alignment search tool

bp base pairs, of nucleic acid

C-terminal carboxyl (COOH)-terminal, of protein

Ca<sup>2+</sup> calcium ion
CaCl<sub>2</sub> calcium chloride
CaM calmodulin

CaSO<sub>4</sub> calcium sulphate Cat. No. catalogue number

CBL calcineurin B-like protein

cDNA complimentary deoxyribonucleic acid

CDPK calcium-dependent protein kinase

CIMMYT International Maize and Wheat Improvement Centre (Centro Internacional de

Mejoramiento de Maíz y Trigo)

CIPK calcineurin B-like (CBL) interacting protein kinase

CI- chloride ion cm centimetre

CML calmodulin-like protein

CO<sub>2</sub> carbon dioxide

Col-0 Arabidopsis ecotype Columbia-0

CRCSLM Cooperative Research Centre for Soil & Land Management

CRISPR/Cas clustered regularly interspersed short palindromic repeats/CRISPR-associated

cv. cultivar

DNA deoxyribonucleic acid

dNTPs deoxynucleotide triphosphates

DREB dehydration-responsive element-binding

dS deciSiemens
DTT dithiothreitol
DW dry weight
E.coli Escherichia coli

EC electrical conductivity

EC<sub>1:5</sub> electrical conductivity of a 1:5 soil to water solution

EC<sub>a</sub> apparent electrical conductivity

EC<sub>e</sub> electrical conductivity of a soil extract EDTA ethylenediaminetetraacetic acid

EF elongation factor
EM electromagnetic

ESP exchangeable sodium percentage

FAO Food and Agricultural Organization of the United Nations

FISH fluorescence in situ hybridization

FW fresh weight g grams(s) g gravity

GC guanine-cytosine, nucleic acid content

gDNA genomic deoxyribonucleic acid

GFP green fluorescent protein
GM genetically modified
GP Golden Promise

GS growth stage, of plant

H+ hydrogen ion

 $H_2O$  water ha hectare

HCL hydrochloric acid

HF high fidelity

HKT high affinity potassium channel

hr hour(s)

Hv Hordeum vulgare

K potassium K+ potassium ion

kb kilobase pairs, of nucleic acid

kg kilogram(s) km kilometre

km<sup>2</sup> square kilometre

L litre

LB luria betani (media or agar)

m metre(s)
M molar
min(s) minute(s)
Mg magnesium

MgCl<sub>2</sub> magnesium chloride

mL millilitre(s)
mm millimetre(s)
mM millimolar

mRNA messenger ribonucleic acid

 $\begin{array}{ll} \text{mS} & \text{milliSiemens} \\ \text{n} & \text{sample size} \\ \text{N} & \text{nitrogen} \\ \text{N}_2 & \text{nitrogen, gas} \end{array}$ 

N-terminal amino (NH<sub>2</sub>)-terminal, of protein

Na+ sodium ion
NaCl sodium chloride

NAF asparagine-alanine-phenylalanine motif (NAF in single amino acid code)

nd not determined ng nanograms

NHX Na+/H+ exchanger

NLWRA National Land & Water Resources Audit

NSCC non-selective cation channel

nt line is not transgenic based on genotyping OGTR Office of the Gene Technology Regulator

Os Oryza sativa
P phosphorus

PIC pre-initiation complex
PCR polymerase chain reaction

PPC2 protein phosphatase 2C-type PPI protein-phosphate interaction

PVC polyvinyl chloride
QTL quantitative trait loci
RNA ribonucleic acid

ROS reactive oxygen species

RT-PCR reverse transcription polymerase chain reaction

S sulphur

s.e.m standard error of the mean SDS sodium dodecyl sulfate

s second(s)

SnRK SNF1 (sucrose non-fermenting 1)-related kinase subgroup

SOS salt overly sensitive

T<sub>1</sub> progeny of the primary transformant containing transgene

TBP TATA-box binding protein(s)

TE tris-EDTA

T<sub>m</sub> melting temperature, of primers

TM unregistered trademark

TGS transgene silencing
TSS transcription start site

U unit(s)

Ubi promoter of maize Ubiquitin-1
UTR untranslated region, of nucleic acid

UV ultraviolet

v/v volume per volume WA Western Australia

### **Abstract**

Soil salinity is a major constraint to crop production in Australia. This has prompted the need to produce salt tolerant cereal cultivars, through the understanding of genes involved in salt tolerance mechanisms and manipulating their expression levels. *Arabidopsis thaliana Calcineurin B-like Interacting Protein Kinase 16 (AtCIPK16)* has been identified as a gene involved in sodium (Na<sup>+</sup>) exclusion. Analysis of *AtCIPK16* alleles from Arabidopsis ecotypes suggests variances in expression are due to differences in the promoters. Experiments in Arabidopsis, barley and wheat (preliminary) have illustrated that *AtCIPK16* overexpression can enhance biomass production through increased Na<sup>+</sup> exclusion, although its full effect in barley and wheat has yet to be properly characterised in both greenhouse and field environments.

The first focus of this project evaluated the salt tolerance of 35S:AtCIPK16 barley (cv. Golden Promise) grown under low and high salinity field conditions in 2013 and 2014 at Kunjin, Western Australia. Comparisons between years were difficult due to waterlogging of the 2013 high salt site and the increased variability in plot establishment in 2014. 35S:AtCIPK16 barley lines had varying responses to high salt conditions depending on the annual rainfall. Results showed Na+ and Clexclusion in certain lines, although this correlated with decreased biomass and yield in high rainfall years. AtCIPK16 expression also increased Na+ and Cl-exclusion in 2012 (a low rainfall year) which instead lead to increasing plant growth and yield.

The second focus of this project aimed to fully characterised the effects of the constitutive expression of *Ubi:AtCIPK16* in wheat (cv. Gladius). Despite conducting three hydroponic experiments, no definitive conclusions about the effects of *AtCIPK16* expression on wheat salt tolerance could be drawn. Although, one sibling transgenic line showed increased Na+ and Cl- exclusion from both root and shoot tissue accompanied by larger biomass under 200 mM salt stress. Despite this finding several factors hinder the analysis of data including the high number of null segregants, considerable variability between siblings of the same transformation event and minimal transgene expression.

The third focus of this project aimed to investigate expression differences between two *AtCIPK16* alleles from the Arabidopsis ecotypes Bay-0 and Shahdara. Since the only differences between the two alleles was a 10 base pair deletion in the Bay-0 promoter, it was hypothesised this deletion was the reason for the increased expression of *AtCIPK16* in Bay-0 as it forms a TATA box (TATATAA). The aim of this project was to alter the expression of each allele by: mutating the last A to a T, removing the TATA box in Bay-0, and mutating the T after the TATATA sequence to an A in Shahdara, forming a TATA box without the deletion. Through PCR mutagenesis the required point mutations were introduced into portions of the two promoter alleles, however due to technical difficulties and time constraints the point mutations were not introduced back into the full promoter constructs driving GFP. It was therefore unable to be determined if the point mutations to the TATA box would indeed affect *AtCIPK16* expression.

### **Declaration**

I certify that this work contains no material which has been accepted for the award of any other degree or diploma in my name, in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

In addition, I certify that no part of this work will, in the future, be used in a submission in my name, for any other degree or diploma in any university or other tertiary institution without the prior approval of the University of Adelaide and where applicable, any partner institution responsible for the joint-award of this degree.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library Search and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.


## **Acknowledgments**

I would like to acknowledge and thank my supervisors Dr. Stuart Roy and Dr. Andrew Jacobs for the guidance and support they have offered throughout my Masters project. It has been an honour and a privilege to have worked with and learnt from you, and I thank you for the patience and understanding you have always shown in your encouragement of my learning.

I am also grateful to the University of Adelaide and the Australian Centre for Plant Functional Genomics (ACPFG) for providing scholarships for the duration of my degree. I would also like to thank the ACPFG and USAID for providing the resources and facilities necessary to undertake my Masters.

I am also grateful to the many people who have helped me during my experiments, especially in conducting field trials. I would like to thank Kalyx Australia (Perth, WA), particularly Dr. Peter Carlton, Mrs. Caris Smith and Mr. Peter Burgess, for their assistance in conducting the GM field trials at Kunjin, WA. I would like also like to acknowledge the work of the ACPFG barley transformation group, ACPFG wheat transformation group and Dr. Parvis Ehsanzadeh for the creation and initial characterisation of the lines used in this project.

I am grateful to for the considerable time and efforts of Ms. Jan Nield who ensured the GM field trials and GM material were compliant to all OGTR licence conditions. I would like to thank Mrs. Ursula Langridge and her glasshouse team as well as The Plant Accelerator for their assistance with the hire of PC2 glasshouses, growth chambers and hydroponic systems. I would like to once again thank my supervisor Dr. Stuart Roy for providing previous years' field data and braving the heat in 2014 to help harvest. I am unendingly grateful to Dr. Rhiannon Schilling for her friendship as well as technical support and for the field material/data provided. Thanks also to Mr. William Heaslip and Ms. Melissa Pickering for their help in harvesting. Considerable thanks to Ms. Melissa Pickering and Ms. Jodie Kretschmer for their never-ending technical support. A final unendingly thank you to Mr. Daniel Menadue for his friendship and support in attempting to keep me sane, especially during long harvest. To all the other members of the ACPFG Salt Focus Group thank you for your advice and support.

Finally I wish to thank my family, particularly my father Paul, step-father Matthew and sister Amethyst, but especially my mother Pam, thank you for your unceasing encouragement and support in believing in me even when I could not.