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**A comparison of stomatal traits between contemporary and fossil leaves of
Melaleuca quinquenervia: Do they reflect climate variation?**

Review of Palaeobotany and Palynology, 2019; 271:104109-1-104109-8

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Published at: <http://dx.doi.org/10.1016/j.revpalbo.2019.104109>

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4 May 2020

<http://hdl.handle.net/2440/124510>

Manuscript Details

Manuscript number

PALBO_2019_21

Title

A comparison of stomatal traits between contemporary and sub-fossil leaves of *Melaleuca quinquenervia*: do they reflect climate variation?

Abstract

Stomatal traits have been shown to vary in predictable ways in response to environmental change in many species. As a consequence, stomatal traits in fossil leaves are sometimes used as proxies for past CO₂ and climate. Here we investigate the influence of temperature, rainfall and CO₂ on stomatal traits in *Melaleuca quinquenervia*. We use both modern and sub-fossil leaves to evaluate the effect of CO₂, and modern leaves for climate variables. We found a significant negative relationship between stomatal size and density across both modern and sub-fossil leaves of *M. quinquenervia*. However, we were unable to find any relationship between stomatal traits and CO₂ across a range from 260-380 ppm. Using the modern data set we were unable to find any robust relationships between stomatal traits and either evaporation or temperature. Apogeotropic roots account for the lack of stomatal anatomy correlation to evaporation in a region that experiences inundation. We conclude that stomatal size is a highly plastic trait in this species and changes do not necessarily reflect functional changes in the leaves.

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Data for: A comparison of stomatal traits between contemporary and sub-fossil leaves of *Melaleuca quinquenervia*: do they reflect climate variation?

Stomatal traits have been shown to vary in predictable ways in response to environmental change in many species. As a consequence, stomatal traits in fossil leaves are sometimes used as proxies for past CO₂ and climate. Here we investigate the influence of temperature, rainfall and CO₂ on stomatal traits in *Melaleuca quinquenervia*. We use both modern and sub-fossil leaves to evaluate the effect of CO₂, and modern leaves for climate variables. We found a significant negative relationship between stomatal size and density across both modern and sub-fossil leaves of *M. quinquenervia*. However, we were unable to find any relationship between stomatal traits and CO₂ across a range from 260-380 ppm. Using the modern data set we were unable to find any robust relationships between stomatal traits and either evaporation or temperature. Apogeotropic roots account for the lack of stomatal anatomy correlation to evaporation in a region that experiences inundation. We conclude that stomatal size is a highly plastic trait in this species and changes do not necessarily reflect functional changes in the leaves.

Highlights

- Modern and subfossil leaves of the same species are analysed for stomatal variation
- Stomatal measures were higher and lower than the modern range
- No effect of climate on modern stomatal traits
- Stomatal size is a plastic trait of *Melaleuca quinquenervia*

Abstract

Stomatal traits have been shown to vary in predictable ways in response to environmental change in many species. As a consequence, stomatal traits in fossil leaves are sometimes used as proxies for past CO₂ and climate. Here we investigate the influence of temperature, rainfall and CO₂ on stomatal traits in *Melaleuca quinquenervia*. We use both modern and sub-fossil leaves to evaluate the effect of CO₂, and modern leaves for climate variables. We found a significant negative relationship between stomatal size and density across both modern and sub-fossil leaves of *M. quinquenervia*. However, we were unable to find any relationship between stomatal traits and CO₂ across a range from 260-380 ppm. Using the modern data set we were unable to find any robust relationships between stomatal traits and either evaporation or temperature. Apogeotropic roots account for the lack of stomatal anatomy correlation to evaporation in a region that experiences inundation. We conclude that stomatal size is a highly plastic trait in this species and changes do not necessarily reflect functional changes in the leaves.

1 **A comparison of stomatal traits between contemporary and sub-**
2 **fossil leaves of *Melaleuca quinquenervia*: do they reflect climate**
3 **variation?**

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11 **Abstract**

12 Stomatal traits have been shown to vary in predictable ways in response to environmental
13 change in many species. As a consequence, stomatal traits in fossil leaves are sometimes used
14 as proxies for past CO₂ and climate. Here we investigate the influence of temperature, rainfall
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18 sub-fossil leaves of *M. quinquenervia*. However, we were unable to find any relationship
19 between stomatal traits and CO₂ across a range from 260-380 ppm. Using the modern data set
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21 evaporation or temperature. Apogeotropic roots account for the lack of stomatal anatomy
22 correlation to evaporation in a region that experiences inundation. We conclude that stomatal
23 size is a highly plastic trait in this species and changes do not necessarily reflect functional
24 changes in the leaves.

25 Key words: Stomatal density, stomatal size, climate proxy, Holocene, *Melaleuca*
26 *quinquenervia*.

27 **1. Introduction**

28 Plants are capable of morphological and physiological plasticity in response to a range of
29 conditions. Plasticity maximises survival in the face of environmental variables such as light,
30 rainfall, humidity, temperature, CO₂ and nutrients. Plant plasticity manifests in a number of
31 ways including changes in leaf size and shape, and stomatal traits that can often be measured
32 in fossils. Stomatal changes influence the maximum potential uptake of CO₂ and control the
33 rate of water loss in leaves. Maximum potential water loss through stomata (g_{wmax}) is a
34 function of stomatal size and density, and is directly related to stomatal conductance for a
35 range of plants. For example, Feild et al. (2011) analysed 87 basal angiosperm species from
36 across the world and found a strong, positive relationship between g_{wmax} and measured
37 stomatal conductance. Maximum potential water loss through stomata determines the upper
38 limit to stomatal conductance and therefore limits photosynthesis and whole canopy gas
39 exchange (de Boer et al., 2011). It can vary in response to changing environmental
40 conditions; for example, it has been shown to increase with decreasing CO₂ thereby
41 increasing the capacity for CO₂ uptake as availability declines (Franks et al., 2012). The
42 sensitivity of g_{wmax} to changing atmospheric CO₂ concentration across the last 400 My has
43 been demonstrated for almost all plant groups, from non-vascular plants to angiosperms
44 (Franks and Beerling, 2009). This g_{wmax} response has also been experimentally demonstrated
45 in living specimens of *Commelina communis*, *Vicia faba*, *Osmunda regalis* and *Selaginella*
46 *uncinata* grown in growth chambers with CO₂ concentrations of 760 to 820 ppm (Franks et
47 al., 2012).

48 Stomatal size and density are affected by environmental factors other than CO₂. These
49 include water availability (Fraser et al., 2009), temperature (Beerling and Chaloner, 1993),
50 nutrients (Peñuelas and Matamala, 1990), light (Onwueme and Johnston, 2000), soil salinity
51 (Bray and Reid, 2002) and humidity (Nejad and Van Meeteren, 2005). Thus, stomatal traits
52 are often used as proxies to infer past climate.

53 There have been times in the past when the atmospheric CO₂ concentration has been
54 relatively stable. One of these periods was the Holocene epoch when CO₂ changed by less
55 than 20 ppm over the 11 700 years prior to ~1850 AD (Indermühle et al., 1999). Therefore,
56 changes in stomatal traits over this time may be due to other environmental factors, such as
57 temperature or rainfall variation. Considerable global and regional scale climatic variability
58 occurred through the Holocene (Wanner et al., 2011). For example, the northern Australasian

59 monsoon was enhanced between 7500 and 5000 years before present (BP), resulting in high
60 rainfall, at times above that of the modern range (Shulmeister and Lees, 1995). In a review of
61 eastern Australian mid-Holocene rainfall variability, Reeves et al. (2013) found that the
62 majority of studies indicate a decline in effective precipitation after 5000 BP. These proposed
63 rainfall fluctuations may have influenced stomatal traits across the Holocene.

64 The relative stability of atmospheric CO₂ during the Holocene provides an opportunity to
65 examine how stomatal traits may have varied over this time in response to temperature and
66 class A pan evaporation (hereafter, evaporation). In this study, we investigated responses of
67 *Melaleuca quinquenervia* (Cav.) ST Blake to a range of environmental variables. Leaves of
68 *M. quinquenervia* from core samples obtained from both lake sediments (hereafter referred to
69 as sub-fossil) and modern leaf litter (hereafter referred to as the modern dataset) are tested.

70 This is the first study that we know of that incorporates both a long-term leaf litter collection
71 and sub-fossil specimens of the same species and analyses their response to environmental
72 variables.

73 2. Methods

74 2.1 Study site and species

75 *Melaleuca quinquenervia* is an Australian native tree species that occurs on flood plains,
76 wetlands (Greenway, 1994), and littoral zones of lakes (Lockhart et al., 1999). When
77 inundated, the tree grows roots from epicormic buds in an upward direction thus allowing
78 oxygen uptake into roots (McJannet, 2008). It is categorised as scleromorphic, based on
79 features such as fibrous leaves with a thick cuticle, and an evergreen habit (Hill, 1998). *M.*
80 *quinquenervia* occurs along the east coast of Australia in Queensland and New South Wales
81 (Brophy et al., 2013). It also occurs naturally in Indonesia, Papua New Guinea, and New
82 Caledonia (Brophy et al., 2013), the Hawaiian Islands and has naturalised in southern Florida
83 where it is considered an invasive species (Pratt et al., 2014).

84 We measured stomatal traits from *M. quinquenervia* leaves collected between 1992 and 2003
85 from Carbrook Wetland (27.7°S, 153.2°E), a seasonally inundated wetland approximately 40
86 km south of Brisbane (Fig. 1; Greenway, 1994). We also measured stomatal traits of sub-
87 fossil leaves obtained from a sediment core taken from Swallow Lagoon on North Stradbroke
88 Island, Queensland, Australia (27.5°S, 153.4°E). Swallow Lagoon is a small (0.27 ha),

89 oligotrophic, freshwater lake located high (94 m above sea level) in the dunes of North
90 Stradbroke Island, the World's second largest sand island. The lake is perched, meaning that
91 it is separated from the regional water table. Variation in water depth is therefore a function
92 of the balance between precipitation and evaporation. Extant populations of *M.*
93 *quinquenervia* continue to grow as fringing vegetation around the lake.

94 As the modern specimens all came from Carbrook Wetland and all sub-fossil specimens
95 came from Swallow Lagoon, we assume that nutrient availability and soil salinity were
96 similar for all modern and all sub-fossil leaves collected for this study. We also assume that
97 light availability did not affect leaves as the plants we studied have open canopies and do not
98 have significant self-shading.

99 **2.2 Modern leaves**

100 Modern *M. quinquenervia* leaves were taken from samples collected for a leaf-litter
101 monitoring project, the initial results of which are reported in Greenway (1994). Leaf litter
102 was collected in a litter trap at four-week intervals, between April, 1992 and July, 2003, from
103 Carbrook Wetland (Fig. 1). For our study, one leaf was sampled every second month,
104 beginning in April 1992 with an even spread of sampling conducted over the twelve year
105 time frame (n = 4, 5 or 6 per year). A total of 61 leaves were analysed.

106 **2.3 Sub-fossil samples**

107 Sediment cores were taken from a platform, anchored over the deepest part of Swallow
108 Lagoon, in March, 2011. The record is a composite of two cores. Core SL1 (150 - 375 cm),
109 which was collected using a Livingstone corer, was extruded in the field where it was sealed
110 and stored for transportation back to the laboratory for analysis. Core SLP3 (0 - 250 cm) was
111 collected using a clear perspex soft sediment piston corer. This was extruded in the field at 1
112 cm increments from 0 to 150 cm, and the remainder as a single section. The two cores were
113 correlated stratigraphically, using a distinct 5 cm thick band of sand evident in both cores at a
114 depth of 220 cm.

115 The cores were sampled at contiguous 1 cm intervals and the sediment was washed through a
116 500 µm sieve to obtain macrofossil remains. The retained fraction was rinsed in distilled
117 water and *M. quinquenervia* leaf fragments were selected for analysis. Fragments of *M.*
118 *quinquenervia* were identified using distinctive morphological features, such as thick veins,

119 epidermal colour and anatomy of the spongy mesophyll. A chronology for the core, based on
120 twelve ¹⁴C dates on terrestrial leaf macrofossils, indicates the sediment covers the last 7300
121 years (Tibby et al., 2016). The distribution of leaves varied through the sediment column and
122 specimens were collected where samples of sufficient size and number were available. A total
123 of 93 leaf fragments were analysed.

124 **2.4 Cuticle preparation**

125 **2.4.1 Modern leaves**

126 One cm² pieces were cut from the leaf margin half way along the lamina. These were placed
127 in separate test tubes, covered in 80% ethanol v/v, and left for 24 hours at room temperature.
128 The ethanol was then removed and leaves covered in a 2:1 solution of 35% hydrogen
129 peroxide and 80% ethanol (v/v). The test tubes were then placed in a water bath in a fume
130 cupboard and gently heated until the leaf samples were translucent, indicating that the cuticle
131 had separated from the rest of the leaf. The leaf samples were gently rinsed with reverse
132 osmosis (RO) water, and debris was brushed away from the cuticle with a fine camel hair
133 brush. The cuticles were stained with 0.05% crystal violet (w/v) for 10 seconds. Leaf cuticles
134 were transferred to a slide, and mounted in warm phenol glycerine jelly. A coverslip was
135 placed on top of the sample and left overnight at room temperature. Nail polish was then
136 applied to the coverslip edges to preserve the cuticles from dehydration.

137 **2.4.2 Sub-fossil samples**

138 Leaf fragments were covered in a solution of 10% aqueous chromium trioxide (w/v), and left
139 for between 24 hours and 5 days at room temperature. The resulting leaf cuticles were rinsed
140 in RO water, then stained and then mounted onto slides using the same method as described
141 for the modern leaves. Fossil cuticle slides are stored at The University of Adelaide
142 collection.

143 **2.5 Stomatal measurements**

144 Leaf cuticles were examined with a light microscope (Olympus AX70), and
145 photomicrographs were taken with an Olympus UC50 camera. Micrographs were analysed
146 using imaging software (AnalySIS version 6.0.6001 Service Pack 1 Build 6001, Acer,

147 Australia). In the case of the sub-fossil leaves, three pieces of leaf cuticle per one cm of leaf
148 core section were measured; these pieces of leaf cuticle were often fragmentary.

149 Stomatal size (μm^2) was calculated as the mean of length by width of five guard cell pairs per
150 piece of cuticle. Stomatal density (stomata mm^{-2}) was determined by counting the number of
151 stomata in a minimum 100 x 100 μm area on each cuticle. Where possible, a 200 x 200 μm
152 area was used for stomatal counts with three of these areas counted per cuticle. Three of the
153 one cm core sections had only one leaf fragment available for counting density, and three had
154 only two leaf fragments available for this purpose. Final numbers for stomatal density were
155 $n=62$, and for stomatal size were $n=93$. Stomatal density had fewer measurements as some
156 leaf pieces were too small to determine stomatal density but we were able to measure
157 stomatal size.

158 For the modern leaves we present individual measurements of the abaxial and adaxial leaf
159 surfaces for stomatal density and size and compare these with stomatal traits from sub-fossil
160 samples.

161 **2.6 Environmental data**

162 Environmental data were obtained using the Scientific Information for Land Owners (SILO)
163 database for the location 27.7°S, 153.2°E. SILO is a database compiled and interpolated from
164 observational data collected by the Australian Bureau of Meteorology (Jeffrey et al., 2001)
165 The data obtained from SILO were mean annual values for minimum air temperature ($^{\circ}\text{C}$),
166 and class A pan evaporation (mm year^{-1}).

167 The mean annual minimum temperature ranged from 14.1 $^{\circ}\text{C}$ for the 1994 dataset to 15.7 $^{\circ}\text{C}$
168 for the 1998 dataset. Class A pan evaporation ranged from 3.97 mm year^{-1} in 1999 to 4.66
169 mm year^{-1} in 1994. As leaf longevity for *M. quinquenervia* is 2-4 years (M. Greenway,
170 Griffith University, Australia pers. comm; Van et al., 2002), environmental data from two
171 years prior to leaf litter collection have been compared with stomatal data.

172 **2.7 Statistical analyses**

173 All statistical analyses have been run in RStudio. Locally Weighted Least Squares Regression
174 (loess) shows smoothed relationships between stomatal density and size for both modern and
175 sub-fossil specimens. Loess also shows smoothed relationships between stomatal size and
176 climate variables.

177 ANOVAs have been performed to test significance between the measures of sub-fossil and
178 modern populations of stomatal size and density.

179 **3. Results**

180 **3.1 Stomatal size and density**

181 The difference between sub-fossil and modern stomatal density, size is not statistically
182 significant. Modern and sub-fossil differences are as follows: stomatal density $P = 0.264$;
183 stomatal size $P = 0.327$. Notably, there was a greater range in values for stomatal size and
184 density in the sub-fossil data set than the modern one (Figs. 5 and 6). The sub-fossil leaves
185 contained the largest and smallest stomata, and likewise the highest and lowest stomatal
186 densities. For the combined modern and sub-fossil data sets, there was a negative relationship
187 between stomatal size and density (Fig. 3).

188 **3.2 Correlation of stomatal size to climate**

189 The assumptions for linearity were not satisfied for any relationships discussed here, thus any
190 correlations can only be considered as general trends. Stomatal size of leaves from the
191 modern dataset was weakly, but significantly correlated with class A pan evaporation (Fig.
192 4). Variation in class A pan evaporation explained 8.1% of the variation in stomatal size for
193 *M. quinquenervia*. Stomatal density did not correlate with class A pan evaporation. There
194 were also no correlations between stomatal size or density from the modern dataset with
195 mean daily minimum temperature. There is a weak correlation between minimum
196 temperature and stomatal size; minimum temperature explained 5.6% of the variation in
197 stomatal size.

198 **4. Discussion**

199 This study investigated relationships between stomatal morphology and temperature,
200 evaporation and CO_2 using *M. quinquenervia* leaf litter collected over twelve years from
201 1992 to 2003. We also measured stomatal traits of sub-fossils of the same species collected
202 from a lake covering the period from 7300 years ago to 1975 AD.

203 4.1 *Stomatal density and size*

204 The negative relationship we observed between stomatal density and size of both the modern
205 and sub-fossil *Melaleuca quinquenervia* leaves was similar to observations reported in
206 previous studies on different species. These studies have shown a negative, logarithmic
207 relationship between stomatal size and density (for example Brodribb et al., 2013; Franks and
208 Beerling, 2009). This relationship between stomatal size and density is likely to be due to the
209 limited leaf area that is available for stomata (Brodribb et al., 2013), but it also has functional
210 significance as more, smaller stomata result in a higher maximum potential water loss than
211 fewer, larger stomata with deeper pores (Brodribb et al., 2013; Franks and Beerling, 2009).

212 4.2 *Climate correlations*

213 Minimum temperature was weakly correlated with short-term changes in stomatal size in the
214 modern *M. quinquenervia* leaves (Fig. 3). A larger stoma creates a longer diffusion path for
215 water to travel along when exiting the leaf during transpiration (Nobel, 2009), and thus
216 reduces transpiration rates. Our analyses showed that larger stomata in *M. quinquenervia*
217 leaves formed during periods of warmer minimum temperatures. Temperature is directly
218 proportional to vapour pressure deficit, and warmer minimum temperatures create a larger
219 VPD and thus a greater driving force for water to evaporate through open stomata. For a
220 given VPD, larger stomata would slow evaporation relative to smaller stomata, and thus
221 retain water for longer.

222 There was no correlation between stomatal density and temperature, thus, it could be argued
223 that there was no functional change in potential water loss as only stomatal size changed
224 weakly in response to temperature. The lack of response by stomatal density to temperature
225 leads to the hypothesis that stomatal size is a more plastic phenotype than the former two
226 variables with temperature changes.

227 There is no correlation between stomatal size and evaporation. It is possible, however, that
228 stomatal size in leaves of *M. quinquenervia* is unrelated to evaporation because it frequently
229 occurs in areas where the water table is high (e.g. wetlands and lake edges). It also has
230 apogeotropic roots that are adapted to rising and falling water tables. McJannet (2008)
231 demonstrated that stand transpiration of *M. quinquenervia* was unaffected by variation in
232 water table depth due to these root systems. Thus, in this high water environment,
233 evaporation is unlikely to be a limiting factor.

234 Throughout the 7300 year Holocene leaf accumulation, the stomatal size of the sub-fossil
235 samples regularly fall outside the range of that of the modern dataset. The correlation of
236 stomatal size of the modern dataset to minimum temperature likely reflects the correlation
237 between these variables during the Holocene, although it is important to note the temporal
238 range between these two is quite different, since we are comparing a 12 year data set with a
239 7500 year one and the range of temperatures and evaporation will be different, but we do not
240 know the extent of this. As such, the weak correlation between stomatal size and minimum
241 temperature can not be considered for use as a palaeo-temperature proxy. Change in stomatal
242 anatomy has been shown to correlate with CO₂ changes though we do not consider this to be
243 the case for change in *M. quinquenervia* stomatal size as CO₂ only increased by ~20ppm
244 during the Holocene (Indermühle et al., 1999).

245 During the Holocene, large changes in El Niño-Southern Oscillation (ENSO) caused
246 variability in rainfall events; an intensification of ENSO in the late-Holocene has been noted
247 in a range of proxies from the Eastern Pacific (for example Conroy et al., 2008; Koutavas and
248 Joanides, 2012; Moy et al., 2002) and Australia (for example Barr et al., 2019; Donders et al.,
249 2007; Quigley et al., 2010; Shulmeister and Lees, 1995). However, an intensification of
250 ENSO leading to larger rainfall events mediated by *M. quinquenervia*'s root system is thus
251 not reflected in stomatal size during the Holocene. There is evidence that there were cold
252 periods in other parts of the world apart from Stradbroke Island during the Holocene (Wanner
253 et al., 2011). Barr et al. (2019) used sediments from the same species in the same region of
254 Australia to calculate a rainfall proxy for Stradbroke Island during the Holocene. These
255 authors were able to use carbon isotope values to do this reconstruction and our data are not
256 similar to theirs (Barr et al., 2019) as we were unable to demonstrate a rainfall response in
257 stomatal density or size and thus could not reconstruct a proxy for rainfall. Potential reasons
258 for this include stomatal size and density responses occurring at a longer time scale-years-
259 than carbon isotopes-weeks. It is also possible that *M. quinquenervia* stomata are not that
260 sensitive to environmental changes, whereas carbon isotope composition is sensitive to
261 rainfall variation. Finally, it is always going to be more difficult to obtain stomatal data
262 because of the limited amount of sub-fossil material available, whereas the *M quinquenervia*
263 C isotope data were obtained from sediment cores. Thus, is that carbon isotope composition
264 may be a better proxy than stomata.

265 However, more evidence of temperature and evaporation forcing on stomatal morphology is
266 required before we can use these data to create proxies of Holocene climate. The

267 relationships found here are weak for temperature and evaporation response and more data
268 are needed to detect subtle correlations with climate. These include collection of herbarium
269 data from a wider spatial and temporal range, including invasive *M. quinquenervia* from
270 Florida and an increase in the sample size of sub-fossil leaves.

271 **4.3 Conclusion**

272 This study showed that stomatal size correlated with minimum temperatures indicating a
273 response by leaf anatomy to changes in VPD. Apogeotropic roots account for the lack of
274 stomatal anatomy correlation to evaporation in a region that experiences inundation. The sub-
275 fossil dataset indicates that there may have been climate influences forcing stomatal change,
276 however, we did not find stomatal size or density to be reliable proxies of palaeo-
277 environments. Analysis of a larger dataset of sub-fossil and modern leaves is required to
278 detect any more subtle correlations between stomatal anatomy and climate variables.

279 **5. Acknowledgements**

280 We acknowledge Minjerribah (North Stradbroke Island) and the surrounding waters as
281 Quandamooka Country. This work was possible with an Australian Government APA.
282 Margaret Greenway carried out leaf litter collections of *Melaleuca quinquenervia* from
283 1992—2003 that has been used as material for the modern dataset.

284 6. References

- 285 Barr, C., Tibby, J., Leng, M., Tyler, J., Henderson, A., Overpeck, J., Simpson, G., Cole, J.,
286 Phipps, S., Marshall, J., McGregor, G., Hua, Q., McRobie, F., 2019. Holocene El Niño–
287 Southern Oscillation variability reflected in subtropical Australian precipitation. Scientific
288 Reports In press.
- 289 Beerling, D., Chaloner, W., 1993. The impact of atmospheric CO₂ and temperature change on
290 stomatal density: observations from *Quercus robur* lammas leaves. *Annals of Botany* 71,
291 231-235.
- 292 Bray, S., Reid, D., 2002. The effect of salinity and CO₂ enrichment on the growth and
293 anatomy of the second trifoliate leaf of *Phaseolus vulgaris*. *Canadian Journal of Botany* 80,
294 349-359.
- 295 Brodribb, T.J., Jordan, G.J., Carpenter, R.J., 2013. Unified changes in cell size permit
296 coordinated leaf evolution. *New Phytologist* 199, 559-570.
- 297 Brophy, J.J., Craven, L.A., Doran, J.C., 2013. Melaleucas: their botany, essential oils and
298 uses. Canberra: Australian Centre for International Agricultural Research (ACIAR).
- 299 Conroy, J.L., Overpeck, J.T., Cole, J.E., Shanahan, T.M., Steinitz-Kannan, M., 2008.
300 Holocene changes in eastern tropical Pacific climate inferred from a Galápagos lake sediment
301 record. *Quaternary Science Reviews* 27, 1166-1180.
- 302 de Boer, H.J., Lammertsma, E.I., Wagner-Cremer, F., Dilcher, D.L., Wassen, M.J., Dekker,
303 S.C., 2011. Climate forcing due to optimization of maximal leaf conductance in subtropical
304 vegetation under rising CO₂. *Proceedings of the National Academy of Sciences* 108, 4041-
305 4046.
- 306 Donders, T.H., Haberle, S.G., Hope, G., Wagner, F., Visscher, H., 2007. Pollen evidence for
307 the transition of the Eastern Australian climate system from the post-glacial to the present-
308 day ENSO mode. *Quaternary Science Reviews* 26, 1621-1637.
- 309 Feild, T.S., Upchurch, G.R., Chatelet, D.S., Brodribb, T.J., Grubbs, K.C., Samain, M.-S.,
310 Wanke, S., 2011. Fossil evidence for low gas exchange capacities for Early Cretaceous
311 angiosperm leaves. *Paleobiology* 37, 195-213.
- 312 Franks, P., Beerling, D., 2009. Maximum leaf conductance driven by CO₂ effects on stomatal
313 size and density over geologic time. *Proceedings of the National Academy of Sciences* 106,
314 10343-10347.
- 315 Franks, P.J., Leitch, I.J., Ruszala, E.M., Hetherington, A.M., Beerling, D.J., 2012.
316 Physiological framework for adaptation of stomata to CO₂ from glacial to future
317 concentrations. *Philosophical Transactions of the Royal Society B: Biological Sciences* 367,
318 537-546.
- 319 Fraser, L.H., Greenall, A., Carlyle, C., Turkington, R., Friedman, C.R., 2009. Adaptive
320 phenotypic plasticity of *Pseudoroegneria spicata*: response of stomatal density, leaf area and
321 biomass to changes in water supply and increased temperature. *Annals of Botany* 103, 769-
322 775.
- 323 Greenway, M., 1994. Litter accession and accumulation in a *Melaleuca quinquenervia* (Cav.)
324 S.T. Blake wetland in south-eastern Queensland. *Marine and Freshwater Research* 45, 1509-
325 1519.
- 326 Hill, R., 1998. Fossil evidence for the onset of xeromorphy and scleromorphy in Australian
327 Proteaceae. *Australian Systematic Botany* 11, 391-400.
- 328 Indermühle, A., Stocker, T.F., Joos, F., Fischer, H., Smith, H.J., Wahlen, M., Deck, B.,
329 Mastroianni, D., Tschumi, J., Blunier, T., Meyer, R., Stauffer, B., 1999. Holocene carbon-
330 cycle dynamics based on CO₂ trapped in ice at Taylor Dome, Antarctica. *Nature* 398, 121-
331 126.

332 Jeffrey, S.J., Carter, J.O., Moodie, K.B., Beswick, A.R., 2001. Using spatial interpolation to
333 construct a comprehensive archive of Australian climate data. *Environmental Modelling &*
334 *Software* 16, 309-330.

335 Koutavas, A., Joanides, S., 2012. El Niño–Southern Oscillation extrema in the Holocene and
336 Last Glacial Maximum. *Paleoceanography* 27, PA4208.

337 Lockhart, C., Austin, D.F., Aumen, N.G., 1999. Water level effects on growth of *Melaleuca*
338 seedlings from Lake Okeechobee (Florida, USA) littoral zone. *Environmental Management*
339 23, 507-518.

340 McJannet, D., 2008. Water table and transpiration dynamics in a seasonally inundated
341 *Melaleuca quinquenervia* forest, north Queensland, Australia. *Hydrological Processes* 22,
342 3079-3090.

343 Moy, C.M., Seltzer, G.O., Rodbell, D.T., Anderson, D.M., 2002. Variability of El
344 Niño/Southern Oscillation activity at millennial timescales during the Holocene epoch.
345 *Nature* 420, 162-165.

346 Nejad, A.R., Van Meeteren, U., 2005. Stomatal response characteristics of *Tradescantia*
347 *virginiana* grown at high relative air humidity. *Physiologia Plantarum* 125, 324-332.

348 Nobel, P.S., 2009. *Physicochemical and environmental plant physiology*, Fourth ed. San
349 Diego, CA: Academic Press.

350 Onwueme, I., Johnston, M., 2000. Influence of shade on stomatal density, leaf size and other
351 leaf characteristics in the major tropical root crops, tannia, sweet potato, yam cassava and
352 taro. *Experimental Agriculture* 36, 509-516.

353 Peñuelas, J., Matamala, R., 1990. Changes in N and S leaf content, stomatal density and
354 specific leaf area of 14 plant species during the last three centuries of CO₂ increase. *Journal*
355 *of Experimental Botany* 41, 1119-1124.

356 Pratt, P.D., Rayamajhi, M.B., Brown, B., Purcell, M.F., Center, T.D., 2014. Within-plant
357 distribution of the *Melaleuca quinquenervia* biological control agent *Lophodiplosis trifida*.
358 *Biocontrol Science and Technology* 24, 1073-1076.

359 Quigley, M.C., Horton, T., Hellstrom, J.C., Cupper, M.L., Sandiford, M., 2010. Holocene
360 climate change in arid Australia from speleothem and alluvial records. *Holocene* 20, 1093-
361 1104.

362 Reeves, J.M., Barrows, T.T., Cohen, T.J., Kiem, A.S., Bostock, H.C., Fitzsimmons, K.E.,
363 Jansen, J.D., Kemp, J., Krause, C., Petherick, L., Phipps, S.J., 2013. Climate variability over
364 the last 35,000 years recorded in marine and terrestrial archives in the Australian region: an
365 OZ-INTIMATE compilation. *Quaternary Science Reviews* 74, 21-34.

366 Shulmeister, J., Lees, B.G., 1995. Pollen evidence from tropical Australia for the onset of an
367 ENSO-dominated climate at c. 4000 BP. *The Holocene* 5, 10-18.

368 Tibby, J., Barr, C., McInerney, F.A., Henderson, A.C.G., Leng, M.J., Greenway, M.,
369 Marshall, J.C., McGregor, G.B., Tyler, J.J., McNeil, V., 2016. Carbon isotope discrimination
370 in leaves of the broad-leaved paperbark tree, *Melaleuca quinquenervia*, as a tool for
371 quantifying past tropical and subtropical rainfall. *Global Change Biology* 22, 3474-3486.

372 Van, T.K., Rayachhetry, M.B., Center, T.C., Pratt, P.D., 2002. Litter dynamics and
373 phenology of *Melaleuca quinquenervia* in South Florida. *Journal of Aquatic Plant*
374 *Management* 40, 22-27.

375 Wanner, H., Solomina, O., Grosjean, M., Ritz, S.P., Jetel, M., 2011. Structure and origin of
376 Holocene cold events. *Quaternary Science Reviews* 30, 3109-3123.

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378 **7. List of Figures and Tables**

379 Table 1. Ranges and means of stomatal parameters for modern and sub-fossil datasets
380 of *Melaleuca quinquenervia*. For the modern dataset, both stomatal size and density have the
381 same number of observations, n= 126. For the sub-fossil material stomatal density
382 measurements, n = 177, for the sub-fossil material stomatal size measurements n = 184.

383 Figure 1. Location of field sites where the modern leaves (Carbrook) and sub-fossil
384 leaves (Swallow Lagoon) of *M. quinquenervia* were collected (map modified from Figure 1c
385 of Tibby et al., 2016).

386 Figure 2. The relationship between stomatal density and size for modern (closed circles)
387 and sub-fossil (open circles) leaves of *M. quinquenervia*. The line is a loess smoothed model.

388 Figure 3. Relationship between mean daily minimum temperature and stomatal size is a
389 smoothed loess line for the modern *M. quinquenervia* leaves (Multiple R-squared = 0.08, P =
390 0.001).

391 Figure 4. Relationship between class A pan evaporation and stomatal size is a smoothed
392 loess line for the modern *M. quinquenervia* leaves (Multiple R-squared = 0.056, P = 0.007).

393 Figure 5. Changes in stomatal density during the Holocene for *M. quinquenervia* from
394 sub-fossil samples. The dotted lines indicate the maximum and minimum values of stomatal
395 density, for the modern dataset (Table 1).

396 Figure 6. Changes in stomatal size during the Holocene for *M. quinquenervia* from sub-
397 fossil samples. The dotted lines indicate the maximum and minimum values of stomatal size,
398 for the modern dataset (Table 1).

399

400 **8. Tables**

401 Table 1.

	Modern data set			Sub-fossil data set		
	Min.	Max.	Mean \pm SE	Min.	Max.	Mean \pm SE
Stomatal density (stomata mm ⁻²)	175	525	327 \pm 6	75	900	344 \pm 12
Stomatal size (μm^2)	353	1240	740 \pm 13	139	1699	710 \pm 24

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