



# Ontogenetic allometry underlies trophic diversity in sea turtles (Cheloniodea)

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## Abstract

Despite only comprising seven species, extant sea turtles (Cheloniidae and Dermochelyidae) display great ecological diversity, with most species inhabiting a unique dietary niche as adults. This adult diversity is remarkable given that all species share the same dietary niche as juveniles. These ontogenetic shifts in diet, as well as a dramatic increase in body size, make sea turtles an excellent group to examine how morphological diversity arises by allometric processes and life habit specialisation. Using three-dimensional geometric morphometrics, we characterise ontogenetic allometry in the skulls of all seven species and evaluate variation in the context of phylogenetic history and diet. Among the sample, the olive ridley (*Lepidochelys olivacea*) has a seemingly average sea turtle skull shape and generalised diet, whereas the green (*Chelonia mydas*) and hawksbill (*Eretmochelys imbricata*) show different extremes of snout shape associated with their modes of food gathering (grazing vs. grasping, respectively). Our ontogenetic findings corroborate previous suggestions that the skull of the leatherback (*Dermochelys coriacea*) is pedomorphic, having similar skull proportions to hatchlings of other sea turtle species and retaining a hatchling-like diet of relatively soft bodied organisms. The flatback sea turtle (*Natator depressus*) shows a similar but less extreme pattern. By contrast, the loggerhead sea turtle (*Caretta caretta*) shows a peramorphic signal associated with increased jaw muscle volumes that allow predation on hard shelled prey. The Kemp's ridley (*Lepidochelys kempii*) has a peramorphic skull shape compared to its sister species the olive ridley, and a diet that includes harder prey items such as crabs. We suggest that diet may be a significant factor in driving skull shape differences among species. Although the small number of species limits statistical power, differences among skull shape, size, and diet are consistent with the hypothesis that shifts in allometric trajectory facilitated diversification in skull shape as observed in an increasing number of vertebrate groups.

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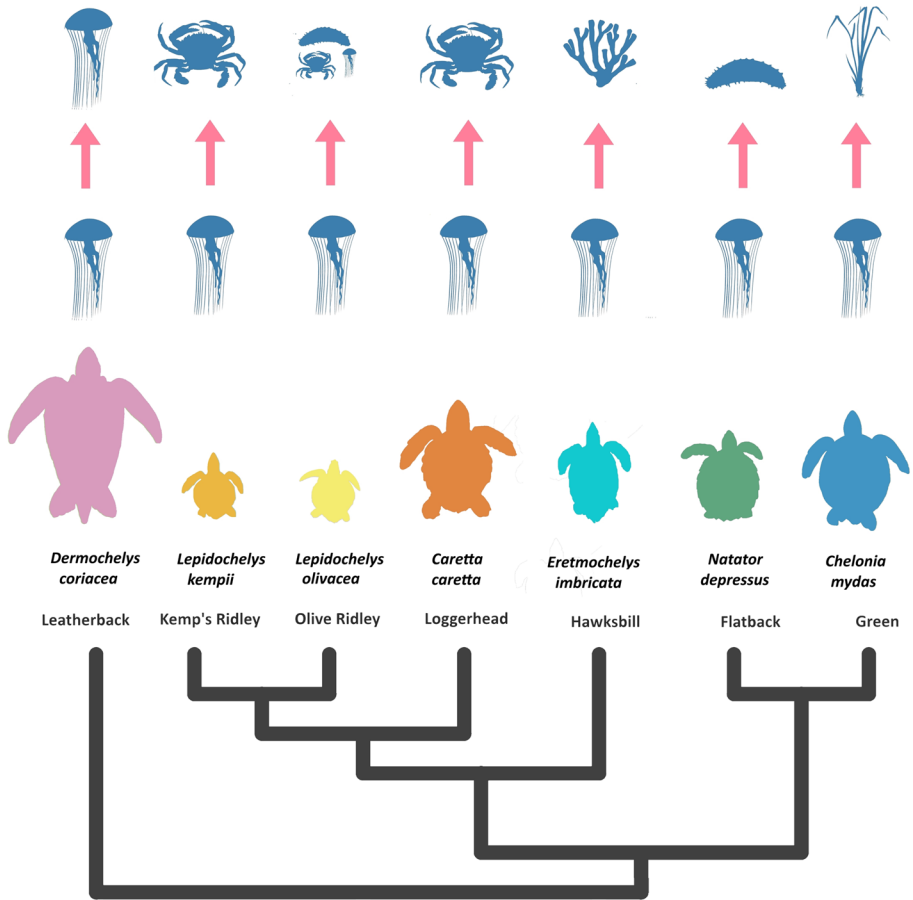
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## Introduction

Among vertebrates, changing shape as a consequence of changing size (allometry) has been shown to be a primary mechanism for generating morphological diversity (Klingenberg 1998; Erickson et al. 2003; Tokita et al. 2017; Gray et al. 2019; Sherratt et al. 2019). Studies of allometry and ontogeny among animals often focus on the effects of changes in growth and body size on ecology (Urošević et al. 2013; Esquerré et al. 2017; Morris et al. 2019; Gray et al. 2019), as well as their evolutionary and functional consequences (Mitteroecker et al. 2005; Wilson and Sanchez-Villagra 2011; Piras et al. 2011; Bhullar et al. 2016; Esquerré et al. 2017; Morris et al. 2019). Changes in ontogenetic allometry through altered developmental timing (heterochrony) have been demonstrated to be an effective mechanism for dietary adaptations (Denoël et al. 2004; Esquerré et al. 2017; Sherratt et al. 2019), often resulting in differences in skull shape that permit access to new feeding niches (Denoël et al. 2004; Frederich et al. 2008; Morris et al. 2019).

Sea turtles represent an excellent group to study the effects of ontogenetic and evolutionary allometry on dietary habits. They are geographically widespread, monophyletic (Evers et al. 2019), exhibit a range of ecological roles and body sizes (Pritchard and Trebbau 1984; Bjorndal et al. 1997), and have a fossil record with the potential to trace macroecological patterns across deep time (Parham and Pyenson 2010). The seven extant species belong to two families, the monotypic Dermochelyidae and the more speciose Cheloniidae (Fig. 1) (Naro-Maciel et al. 2008; Duchene et al. 2012). These families likely diverged during the Late Cretaceous (Duchene et al. 2012; Thomson et al. 2021), with the crown of Cheloniidae diverging during the late Oligocene or early Miocene (Thomson et al. 2021). There is dramatic size variation among species (average adult weights between 35 and 400 kg; Pritchard and Trebbau 1984; Dodd 1988; Zug and Parham 1996) and within species, with *Dermochelys coriacea* (leatherback) increasing in size by three orders of magnitude during growth (Pritchard and Trebbau 1984; Jones et al. 2011). This increase in size is one of the greatest known among extant amniotes, only matched by some of the largest crocodylians (Brandt 1991; Leach et al. 2009). In contrast to other giant testudines such as the giant tortoises of the Galapagos and Aldabra (*Chelonoidis nigra*, *Aldabrachelys gigantea*), most sea turtles undergo significant changes in diet during ontogeny (Gibson 1983; Fowler de Neira and Johnson 1985; Furrer et al. 2004). Among them, heterochrony has been previously inferred for one species, *D. coriacea*, on the basis of postcranial features associated with juveniles found in the adult skeleton, such as cartilaginous epiphyses and a lack of fusion between cervical neural arches and their corresponding centra (Nick 1911; Pritchard and Trebbau 1984).

Despite their low taxonomic diversity, modern sea turtles display a remarkable ecological breadth, with most species inhabiting a unique dietary niche as adults (Bjorndal et al. 1997) (Fig. 1). In contrast, all juvenile sea turtles share a similar diet of plankton and small pelagic cnidarians (Bjorndal et al. 1997; Bolten 2003). Given the functional roles of the vertebrate skull in food acquisition and processing (Pritchard and Trebbau 1984; Claude et al. 2004; Parham and Pyenson 2010; Jones et al. 2012), the relationship between turtle skull structure and diet has been previously investigated in sea turtles but generally with a focus on one or two species. Examinations of skull development of *Chelonia mydas* (green) and *Caretta caretta* (loggerhead) indicate that dietary shifts are associated



**Fig. 1** Cladogram based on Naro-Maciel et al. (2008) and Duchene et al. (2012). Specimens scaled to relative size, based on available literature (Pritchard and Trebbau 1984; Dodd 1988). Top two rows of symbols represent the dietary categories of juveniles (bottom) and adult (top) members of each species: jellyfish=Pelagic; crab=Durophage; sponge=Sponge; sea cucumber=Soft; sea grass=Herbivore; sea cucumber+crab+jellyfish=General. See Table 1 for further details. Silhouettes redrawn from National Aquarium Baltimore(<https://www.aqua.org/blog/2015/April/oceans-seven>)

with morphological differences between ontogenetic stages (Nishizawa et al. 2010; Coelho et al. 2018; Lunardon et al. 2020). However, the relationship between skull shape and size among extant turtles remains poorly known, which limits our ability to distinguish ontogenetic and phylogenetic shape differences (Jones et al. 2012) which could inhibit interpretations of turtle evolution. For example, heterochrony has been suggested to be a significant factor in the diversification of other reptile groups, contributing to recent morphological disparity (Sherratt et al. 2019; Morris et al. 2019; Gray et al. 2019) as well as early divergences (Esquerré et al. 2017; Morris et al. 2019). Therefore, the understanding of differing allometric patterns can potentially give insights into the evolutionary processes underlying sea turtle diversity.

There have been several morphometric analyses on the skulls of sea turtles. Previous morphometric analyses of sea turtle skulls have either involved linear measurements

(Kamezaki and Matsui 1995, 1997; Kamezaki et al. 2003), landmarks (Nishizawa et al. 2010; Coelho et al. 2018), or both (Lunardon et al. 2020). However, most of these studies focused on variation within one species: *Ca. caretta* (Kamezaki and Matsui 1997; Lunardon et al. 2020) or *Ch. mydas* (Kamezaki and Matsui 1995; Nishizawa et al. 2010; Coelho et al. 2018). Although some of these studies use large sample sizes they do not include all stages of ontogeny: a study of *Ca. caretta* used 80 individuals but none were younger than 10 years old (Lunardon et al. 2020), whereas the study of *Ch. mydas* used 145 individuals all between 3 and 5 years old (Kamezaki and Matsui 1995). Only two studies included multiple species and neither examined growth trajectories or ecological differences. Kamezaki et al. (2003) sampled three species and focused on skull characteristics of *Ca. caretta*. Myers (2007) included six extant species but found *Lepidochelys olivacea* (olive ridley) to be notably different from other Cheloniidae, which appears at odds with several qualitative anatomical comparisons (Gaffney 1979; Pritchard and Trebbau 1984; Jones et al. 2012). Other morphometric studies of turtle skull shape have been broader in scope, examining multiple families or Testudinata as a whole (Claude et al. 2004; Foth et al. 2017). As such, the relationship between skull shape, diet, and size among all seven extant sea turtle species remains unknown.

To fill this gap in knowledge, here we use geometric morphometrics to characterise ontogenetic allometry in the skulls of modern sea turtles. We sample all seven species across a wide size range encompassing hatchlings to large adults. While characterising patterns of ontogenetic allometry in each species, we attempt to distinguish among the effects of diet and phylogenetic history on the relationship between skull shape and size. We aim to determine if heterochrony has been a significant mechanism in the evolution of skull shape within Chelonioidea. This study represents the first systematic examination of sea turtle skull shape with three-dimensional landmarks and the most comprehensive study of allometry in the group to date.

## Methods

### Specimens

We sampled 63 specimens from museum collections representing all seven species of extant sea turtle, choosing as broad a size range as possible (Table 1). All hatchlings were ethanol-preserved as well as one large adult specimen of *Natator depressus* (flatback); all other specimens were dry skulls. Immature sea turtles in their pelagic stage are naturally rarer in museums due to their lower frequency of being washed up on beaches and collected. This factor limited specimen availability, although *Caretta caretta* and *Chelonia mydas* have samples representing all stages of ontogeny and total samples exceeding 10 (15 and 11 respectively). The sample of *Lepidochelys kempii* (Kemp's ridley) lacks hatchlings and large adults whereas *Dermochelys coriacea* lacks intermediate size animals. Otherwise, all species have at least one specimen for each size category (see below).

Specimens were scanned using X-ray computed tomography (CT). Some of the larger specimens were scanned in medical CT machines, whereas others were scanned using X-ray micro-CT at various facilities (Appendix 1 Table 1). The reconstructed data sets had voxel dimensions between 9 and 500  $\mu\text{m}$ . The resulting tiff stacks were processed in AVIZO 9.0 Lite software (Visualisation Science Group, SAS). The cranium was isolated in

an associated label file using the threshold and brush tool. Surface models of the skull were exported as PLY files for measurement (details below).

Specimens were divided into three age classes that broadly reflect ontogenetic differences: hatchling, intermediate, and adult (Table 1). Individuals were categorised as hatchlings ( $n=14$ ) if they exhibited fontanelles and lacked ossified basicranial elements, individuals were categorised as adults ( $n=24$ ) if they had a skull length within two standard deviations of the average reported adult skull size for the species (Pritchard and Trebbau 1984; Dodd 1988; Zangerl et al. 1988; Nishizawa et al. 2010; Lunardon et al. 2020). The remaining individuals were categorised as intermediates ( $n=25$ ), as they had closed fontanelles and ossified basicranial elements but were two standard deviations smaller than the skull size reported for an adult of that species.

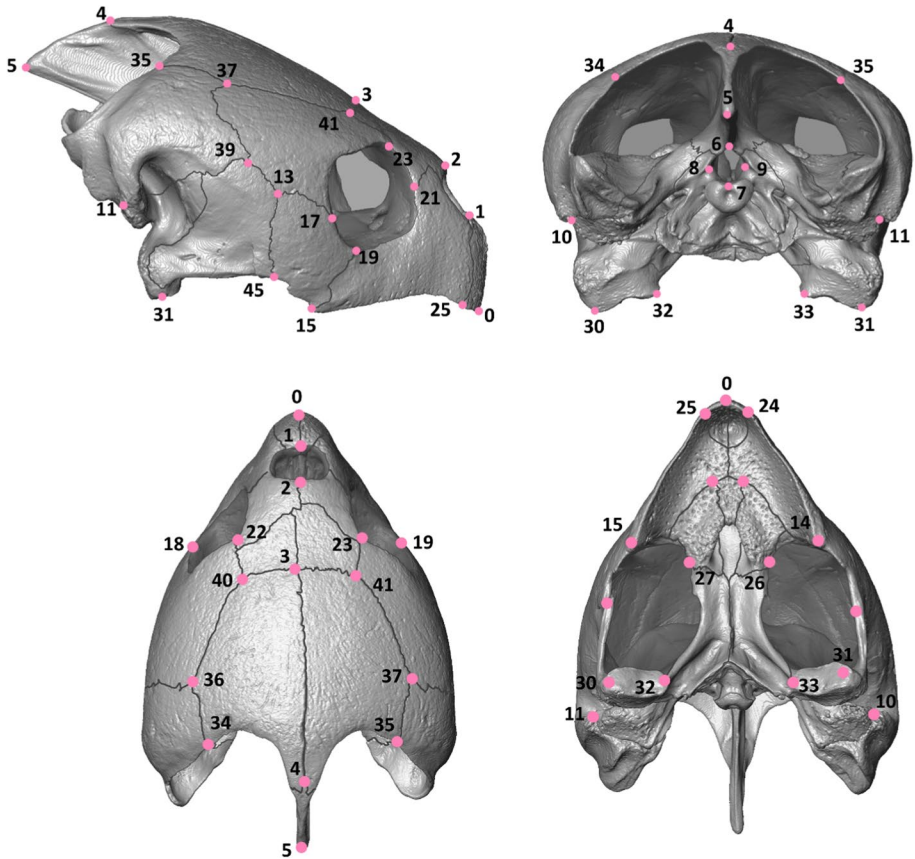
The following age classes and species combinations were used to determine a total of four datasets:

1. *Chelonioides all age classes*: effectively a complete dataset including all seven species and all age classes ( $n=63$ ).
2. *Cheloniidae all age classes*: no *Dermochelys coriacea* ( $n=55$ ).
3. *Chelonioides adults only*: only adult specimens ( $n=21$ ).
4. *Cheloniidae adults only*: only adult specimens and no *D. coriacea* ( $n=17$ ).

Dataset 1 provides a holistic overview of the entire sample, dataset 3 serves to examine evolutionary allometry, whereas datasets 2 and 4 provide an understanding of the samples without potential skew from the highly specialised and taxonomically isolated *D. coriacea*. The four datasets also facilitate comparisons to previous studies that did not include hatchlings or excluded *Dermochelys coriacea* from some analyses (e.g., Myers 2007). Although *D. coriacea* is the sister taxon to Cheloniidae, their lineages diverged in the Cretaceous and have been evolutionarily isolated for over 65 Ma (Duchene et al. 2012).

## Diet classification

Diet was classified into six categories according to food items reportedly consumed in the literature (Bjorndal 1985; Dodd 1988; Bjorndal et al. 1997; Bolten 2003; Limpus 2007; Limpus and Limpus 2007): pelagic (gelatinous invertebrates: jellyfish, crustacean larva, etc.), soft (soft-bodied invertebrates include neritic, benthic, largely non-gelatinous prey), herbivore (plant matter, sea grass, algae), sponge (mainly sponges), durophage (hard-bodied invertebrates: clams, echinoderms, crabs), and general (mixture of jellyfish, fish, crabs, salps/tunicates etc.) (Table 2; Fig. 1). The amount of data for each species is highly variable: some species have been heavily studied such as *Ca. caretta* with studies analysing gut contents (Nierop and Hartog 1984; Seney and Musick 2005), faecal content (Marchiori et al. 2018), and observational data (Babcock 1938; Limpus 1992) whereas other species such as *N. depressus* have received limited study (Limpus 2007). The diet of hatchling specimens is also comparatively understudied due to their pelagic nature, however available data consistently suggests a diet of soft bodied invertebrates that inhabit the upper water column (Bjorndal et al. 1997; Boyle and Limpus 2008).



**Fig. 2** A three-dimensional mesh derived from micro-CT data of *Caretta caretta* (SAMA Unregistered) showing the location of the 46 landmarks used in this analysis (numbered 0–45)

**Table 1** Number of specimens for each species and age class

Species	Hatchlings	Intermediate	Adults	Total
<i>Caretta caretta</i>	4	6	5	15
<i>Chelonia mydas</i>	3	4	4	11
<i>Dermodochelys coriacea</i>	4	0	4	8
<i>Eretmodochelys imbricata</i>	1	4	2	7
<i>Lepidochelys kempii</i>	0	8	0	8
<i>Lepidochelys olivacea</i>	3	3	3	9
<i>Natator depressus</i>	1	1	3	5
Total	16	26	21	63

## Cranial landmarks

The three-dimensional cranial surface models were landmarked in IDAV Landmark Editor

**Table 2** Diet categories and associated references

Species	Age class	Common items	Rare items	Assigned diet category	References
<i>Caretta caretta</i>	Hatchling	Zooplankton, Jellyfish	Sargassm	Pelagic	Dodd (1988) and Boyle and Limpus (2008)
	Intermediate	Jellyfish, Snails, Crabs, Small bivalves; Sea Urchins	Algae; Coral; Salps, fish	General	Dodd (1988) and Burke et al. (1993)
	Adult	Crabs, Snails, Conches, Bivalves, Sea pens, Horseshoe crab, Sea Urchins	Jellyfish, Coral, Algae, fish	Durophage	Dodd (1988) and Seney and Musick (2007)
<i>Chelonia mydas</i>	Hatchling	Zooplankton, Cnidarians	Sargassm	Pelagic	Bjorndal (1985) and Boyle and Limpus (2008)
	Intermediate	Sea grass, Algae	Jellyfish, Coral, Salps	Herbivore	Mortimer (1982), Ross (1985) and Seminoff et al. (2002)
	Adult	Sea grass, Algae	Jellyfish, Coral, Salps	Herbivore	Mortimer (1982), Ross (1985) and Seminoff et al. (2002)
<i>Dermostochelys coriacea</i>	Hatchling	Zooplankton, Cnidarians	N/A	Pelagic	Bleakney (1965)
	Intermediate	Jellyfish	Salps	Pelagic	Bleakney (1965) and Eckert et al. (1989)
	Adult	Jellyfish	Salps	Pelagic	Bleakney (1965), Frazier et al. (1985) and Eckert et al. (1989)
<i>Eretmochelys imbricata</i>	Hatchling	Zooplankton, Cnidarians	N/A	Pelagic	Meylan (1984)
	Intermediate	Sponges, Algae	Coral, Sea Urchins, Anenomes	Sponge	Den Hartog (1979), Meylan (1985), Limpus (1992) and Andres and Uchida (1994)
	Adult	Sponges	Coral, Sea Urchins, Algae, Anenomes	Sponge	Meylan (1985), Limpus (1992) and Andres and Uchida (1994)
<i>Lepidochelys kempii</i>	Hatchling	Zooplankton, Cnidarians	Sargassm	Pelagic	Shaver (1991)
	Intermediate	Crabs, Jellyfish, Sea horses	Algae, Fish	General	Shaver (1991), Burke et al. (1993), Seney and Musick (2005) and Schmid and Tucker (2018)
	Adult	Crabs, Molluscs	Algae, Fish	Durophage	Shaver (1991), Burke et al. (1993) and Burke et al. (1994)

Table 2 (continued)

Species	Age class	Common items	Rare items	Assigned diet category	References
<i>Lepidocheilys olivacea</i>	Hatchling	Zooplankton, Cnidarians	N/A	Pelagic	Bjorndal et al. (1997)
	Intermediate	Salps, molluscs, Crabs, Jellyfish, Snails	Fish, Fish eggs, Algae, Turricates	General	Marquez and Penaflores (1976), Montenegro and Bernal (1982), Frick et al. (2011) and Colman et al. 2014
	Adult	Salps, molluscs, Crabs, Jellyfish, Snails	Fish, Fish eggs, Algae, Turricates	General	Marquez et al. (1976) and Montenegro et al. (1982)
<i>Natator depressus</i>	Hatchling	Zooplankton, Cnidarians	N/A	Pelagic	Limpus (2007)
	Intermediate	N/A	N/A	Soft	Limpus (2007)
	Adult	Sea cucumbers, Jellyfish, Corals, Sea pens	N/A	Soft	Limpus (2007)



v. 3.6 (Wiley 2007). Forty-six landmarks were placed at equivalent locations across each specimen (Fig. 2), representing external suture junctions or distinct anatomical points, e.g., the posterior most tip of the supraoccipital. Landmarks were chosen to best characterise the entirety of cranial shape while still being applicable to every species and ontogenetic stage (Appendix 1). These criteria meant that much of the basicranium was not landmarked because in hatchlings this part of the skull is still represented largely by cartilage without distinct junctions.

The exported landmark coordinates were subjected to a generalised Procrustes superimposition in the R package *geomorph* v. 3.3.3 (Adams et al. 2020), standardising for variation in translation, rotation, and size using the function *gpagen*. Due to lack of body size data for all specimens, centroid size (the square root of the sum of squared distances of the landmarks to their centroid) of each cranium was used as a measure of size.

Missing landmarks were estimated using *estimate.missing* in the package *geomorph*. Only four specimens of *Lepidochelys kempii* were missing landmarks, these were from the anterior portion of the snout (landmarks 0, 1, 24, 25) and the tip of the supraoccipital (landmark 5). This function estimates missing landmarks based on the mean coordinates for each landmark across the entire dataset using the regression method of estimation.

## Shape analysis

Statistical analyses were performed in the R packages *geomorph* v. 3.3.3 (Adams et al. 2020) and *RRPP* v.0.6.2 (Collyer et al. 2020). We performed a principal component analysis (PCA) of the Procrustes aligned coordinates, using the function *gm.prcomp* to determine the main components of shape variation. To interpret the variation described by the major axes we used a combination of thin-plate spline deformation grids, vector analysis, and warping a mesh of the mean shape to the extremes (minimum and maximum PC scores) of each axis. These were implemented using the *warpRefMesh* function.

To test for a relationship between size and shape we used the phylogenetic generalised least squared method (*procD.pgls*). This function performs an ANOVA within a phylogenetic framework assuming a Brownian model of evolution (Adams 2014a; Adams and Collyer 2015). However, this technique normally relies on species being either represented by a single set of coordinates or an aggregate mean for each phylogenetic tip (Prevosti et al. 2011; Püschel and Sellers 2016; Wang et al. 2021). Since our data are multiple individuals aligned along ontogenetic trajectories of individual species, implementations currently available need a small modification to be operable. We created a tree file based on the phylogenetic tree found in Duchene et al. (2012) where each species was represented by a soft polytomy consisting of all their individual specimens, such that the phylogenetic relatedness can be considered while retaining the data structure (e.g., Sanger et al. 2013). It should be noted that the validity of phylogenetic comparative methods on groups with a small number of taxa, as is the case in sea turtles, is still unclear (Blomberg et al. 2003; Adams 2014b). Therefore, we also conducted a Procrustes ANOVA which does not account for phylogeny (using *procD.lm*) for all analyses. In both cases, we evaluated whether a common ontogenetic allometry model or a species unique allometry model better explained the observed patterns for each dataset. This evaluation was achieved by comparing ANOVAs of each model with the null assumption of a common (shared) allometry. The relationship between shape, size, and diet was also assessed using the above methods.

The high number of dietary categories relative to number of species makes it difficult to disentangle differences in shape associated with diet from those associated with

phylogenetic inheritance. However, this problem is somewhat mitigated because dietary overlap does occur between different age classes of some species, e.g. the intermediate age class of *Caretta caretta* and *Lepidochelys kempii* exhibit a general diet like the adults of *Lepidochelys olivacea* (Table 2). Thus, analysing the differences in shape and diet within species may help to inform our understanding of the differences in shape and diet between species.

To assess the strength of evolutionary allometry, the relationship between shape and size of the adult datasets were tested using a PGLS and the original tree (*procD.pgls*). Species pairwise comparisons were performed using the *pairwise* function from the package *RPPP*. This comparison was done to identify differences in slope vector length and orientation, where slope length is magnitude of shape change per unit size and orientation is the direction of shape change per unit size.

To assess whether any of the morphological variation characterised by individual PC axes are significantly correlated with size, we calculated a Pearson's product moment correlation coefficient (*cor.test*) comparing PC scores to centroid size, assuming a normal distribution. This was done towards understanding which particular aspects of morphological variation are most associated with allometry.

We assessed the morphological disparity between ontogenetic groups using the *morphol.disparity* function of *geomorph*. This function estimates disparity of a group (hatchling, intermediate, and adult) as their Procrustes variance using residuals of a linear model fit, in our case using the species unique allometry model.

A residual randomisation procedure with 10,000 iterations was used to assess statistical significance for all tests.

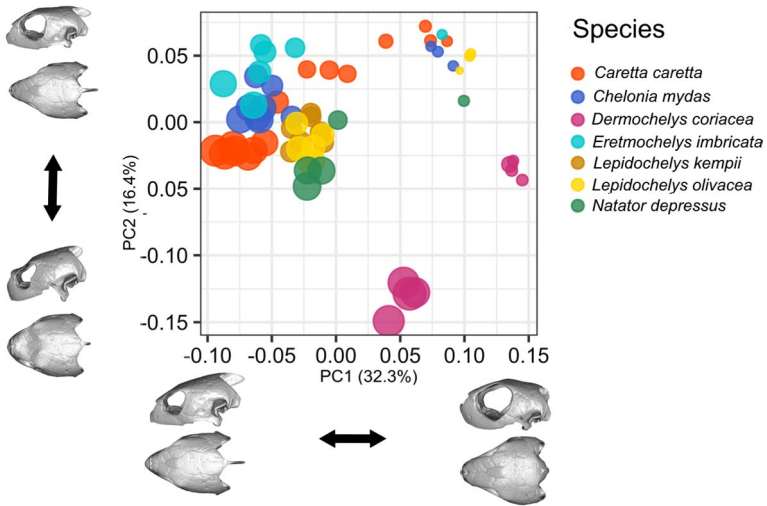
To assess phylogenetic signal, we used the *physignal* function, which assumes a Brownian motion model of evolution, and the phylogenetic tree recovered in Naro-Maciel et al. (2008) and Duchene et al. (2012) (Fig. 1). This function estimates phylogenetic signal using a generalisation of Blomberg's K statistic for high dimensional multivariate data ( $K_{mult}$ ; Adams 2014a). We tested phylogenetic signal for the adults and hatchlings separately, using the mean adult shape and mean hatchling shape for each species, as demonstrated in Gray et al. (2019). As there is a high degree of variability among dated phylogenies of the group (e.g. Duchene et al. 2012; Thomson et al. 2021), we did not use a time calibrated tree. Instead branch length was standardised using the *compute.brLen* function with the "Grafen" method in the R package *ape* v. 5.4 (Paradis and Schlip 2019). This comparison was visualised by projecting the phylogenetic tree into morphospace. Phylogenetic signal was assessed to determine how important phylogenetic relatedness is in determining skull shape among living chelonioidea, as well as to determine if there is a change in phylogenetic signal across ontogeny which might suggest an increased adaptive signal.

## Results

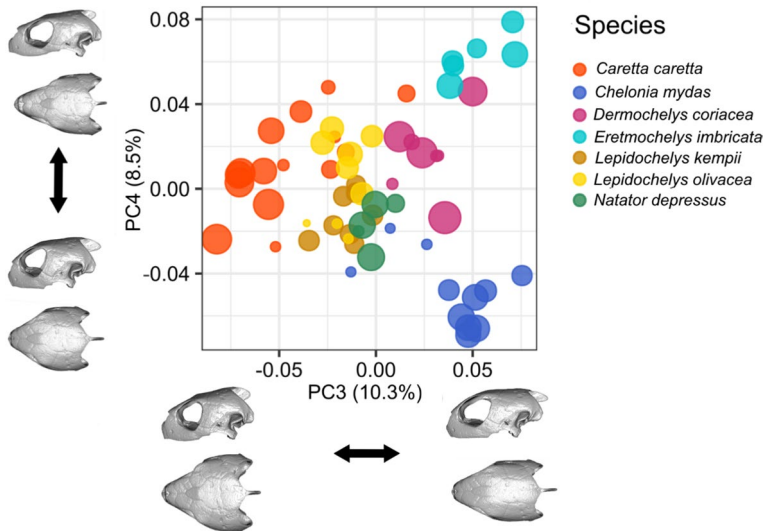
### PCAs

#### Chelonioidea all age classes (total group: n = 63)

The first four PC axes account for 67.9% of the total cranial shape variation and the remaining axes each account for less than 5%. PC scores vary among species (Figs. 3, 4) reflecting differences in skull shape. Overall PC1, and to a lesser extent PC2, are broadly associated



**Fig. 3** PC1 and PC2 of the cranial morphospace of Chelonioidae, with different colours representing different species. Skulls show lateral and dorsal views of the mean shape mesh warped to the coordinates of the extremes for each axis. All points scaled to centroid size



**Fig. 4** PC3 and PC4 of the cranial morphospace of Chelonioidae, with different colours representing different species. Skulls show lateral and dorsal views of the mean shape mesh warped to the coordinates of the extremes for each axis. All points scaled to centroid size

with size and ontogeny whereas the other PC axes appear to reflect species differences and individual variation.

The greatest axis of variation (PC1; 32.3%) represents differences in the relative antero-posterior length of the posterior of the skull, in particular the length of the supraoccipital

crest (Landmarks 5–6), as well as the relative size of the orbit (Fig. 3). High PC1 scores represent relatively large orbits, an anteroposteriorly short posterior part of the skull, and a proportionately short supraoccipital crest whereas low PC1 scores represent relatively small orbits, an anteroposteriorly elongate posterior part of the skull, and a proportionately enlarged supraoccipital crest (Fig. 3). Hatchling skulls have high PC1 scores, juveniles have moderate PC1 scores, and adults tend to have low PC1 scores. However, for *Dermochelys coriacea*, hatchlings have particularly high PC1 scores, and its adults have similar PC1 scores to the hatchlings of other turtles such as *Caretta caretta*. The adult specimens of *Natator depressus* have notably high PC1 scores in comparison to the similarly sized specimens of other cheloniids.

PC2 describes 16.4% of cranial shape variation. High PC2 scores represent a circular orbit, a small ventral projection of the mandibular condyle and a tapering pointed rostrum, whereas low PC2 scores represent an increasing ventral projection of the quadrate and jaw joint, an ovoid orbit, and laterally broader anterior rostrum (Fig. 3). The skulls of larger specimens tend to plot with lower PC2 scores. Skulls with the lowest PC2 scores are exclusively adult *D. coriacea*.

PC3 describes 10.3% of cranial variation (Fig. 4). High PC3 scores represent skulls that have laterally narrow posterior sections of the skull and have a dorsoventrally shallow lateral profile whereas low PC3 scores represent skulls that have laterally wide posterior sections of the skull and are dorsoventrally deep (Fig. 4). High PC3 scores characterise both *Chelonia mydas* and *Eretmochelys imbricata* whereas low PC3 scores characterise the wide cranium of *Ca. caretta*.

PC4 describes 8.5% of cranial variation. High PC4 scores represent skulls that have an elongated rostrum whereas low PC4 scores represent skulls with a shorter, blunt rostrum (Fig. 3B). High PC4 scores characterise the long snouted *E. imbricata* whereas low PC4 scores characterise the blunt snouted *Ch. mydas* (Fig. 4).

### Cheloniidae all age classes (no *Dermochelys coriacea*: n = 55)

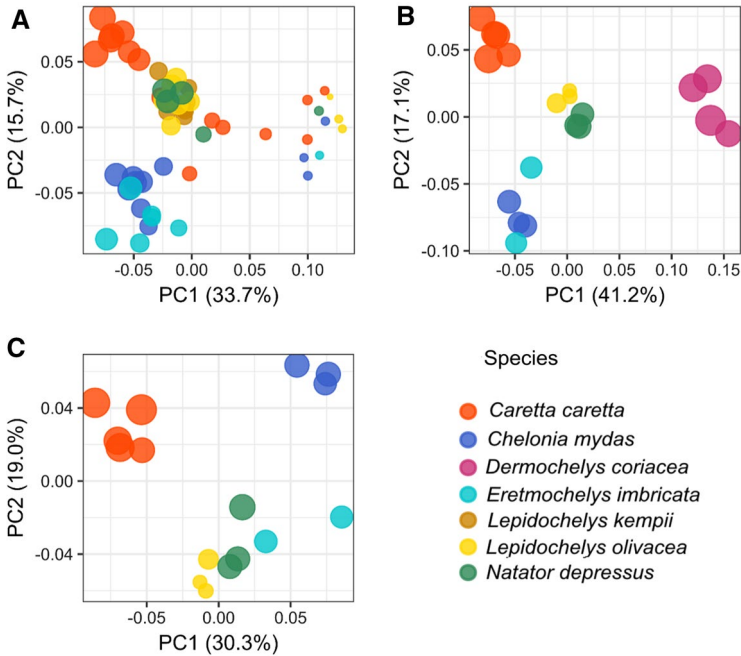
When *D. coriacea* is excluded from the sample, PC1 (33.7%) is still characterised by the same variation seen in the total group dataset (Fig. 5A). High PC1 scores represent relatively large orbits and small posterior part of the skulls whereas low PC1 scores are characterised by relatively small orbits and large posterior part of the skulls. The characterisation of PC2 (15.7%) is similar to that of PC3 of the total group dataset and describes the relative width of the posterior part of the skull with high PC2 scores having relatively wide skulls and low PC2 scores having relatively narrow skulls.

### Chelonioidae adults (n = 21)

PC1 for the total group adult is largely similar to PC1 for the total group, with orbit size and posterior part of the skull size characterising the axis (Fig. 5B). PC2 for the adult subset is similar to PC3 for the total group, characterised by the relative width of the posterior part of the skull.

### Cheloniidae adults (no *Dermochelys coriacea*: n = 17)

PC1 (30.3%) for the cheloniid adult subset is similar to PC3 for the total group, characterised by the relative width of the posterior part of the skull. PC2 (19%) is characterised



**Fig. 5** PC1 versus PC2 of all Cheloniidae (A), adult Cheloniidae (B), and adult Cheloniidae (C). Colours represent different species. All points are scaled by centroid size

by the shape of the squamosal and jugal (Fig. 5C). High PC2 scores are characterised by a more curved squamosal with the ventroposterior contact with the quadrate being more anteriorly located, and a more anteriorly located contact between the jugal and quadratojugal. Low PC2 scores are characterised by a less curved squamosal with the ventroposterior contact with the quadrate being more posteriorly located, and a more posteriorly located contact between the jugal and quadratojugal.

**Table 3** Procrustes ANOVAs for the Chelonioidae all age class dataset assessing species interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsq	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	1.54	1.54	0.40	48.29	4.15	<0.001
Species	6	0.09	0.01	0.02	0.45	-6.05	1
Log (size): species	6	0.65	0.11	0.17	3.39	6.48	<0.001
Residuals	49	1.56	0.03	0.41			
Total	62	3.83					
<i>procD.lm</i>							
Log (size)	1	0.22	0.22	0.25	50.84	8.35	<0.001
Species	6	0.36	0.06	0.41	13.87	12.94	<0.001
Log (size): species	6	0.08	0.01	0.09	3.17	7.47	<0.001
Residuals	49	0.21	0.00	0.24			
Total	62	0.86					

## Ontogenetic allometry

### Chelonioida all age classes (total group: n = 63)

A significant relationship between shape and size is found using both phylogenetic as well as non-phylogenetic comparative methods ( $p < 0.001$ ; Table 3; Fig. 6). Both PC1 and PC2 are significantly related to size (both  $p < 0.001$ ), whereas the other PC axes are not. This result suggests that orbit and posterior skull size both scale allometrically. Species and diet were found to have a significant interaction with size, as well as with each other. For both methods a model using unique species-specific allometries was a significant improvement over a common one ( $p < 0.001$ ). There was no significant difference in model strength between size and diet or size and species, but both are significantly better than size alone ( $p < 0.001$ ; Table 4).

Adults occupy a significantly ( $p < 0.01$ ) more disparate morphospace than the other two ontogenetic groups. There is not a significant difference in disparity between intermediate and hatchling specimens. Adults occupied a larger area of morphospace (Procrustes variance = 0.0131) than hatchlings (Procrustes variance = 0.008) or intermediate specimens (Procrustes variance = 0.007). This result suggests that the crania of hatchling sea turtles more closely resemble each other than do adult crania.

Most species pairs had significantly different slope angles (Table 5). However, *Natator depressus* was only different to either *D. coriacea* or *L. olivacea*. *Lepidochelys kempii* is also not significantly different to any other species in slope angle, but this result may be due to the small size range. *Caretta caretta* and *L. kempii* have the steepest slopes, whereas *L. olivacea* and *N. depressus* having the shallowest ones.

There are few differences in slope length among species (Table 6). *N. depressus* differs significantly from *Ch. mydas*, *L. olivacea*, and *E. imbricata*. Also *D. coriacea* significantly differs in slope length from *Ca. caretta*, *Ch. mydas*, and *E. imbricata* (Table 6). *L. kempii* is not significantly different to any species.

**Table 4** Procrustes ANOVAs for the Chelonioida all age class dataset assessing diet interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsq	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	1.54	1.54	0.40	50.06	4.17	<0.001
Diet	5	0.60	0.12	0.16	3.89	7.31	<0.001
Log (size): diet	5	0.24	0.05	0.06	1.58	2.99	0.001
Log (size): diet: species	8	0.13	0.02	0.03	0.53	− 5.46	1
Residuals	43	1.32	0.03	0.34			
Total	62	3.83					
<i>procD.lm</i>							
Log (size)	1	0.22	0.22	0.25	53.38	8.42	<0.001
Diet	5	0.30	0.06	0.35	14.66	11.91	<0.001
Log (size): diet	5	0.05	0.01	0.06	2.58	5.27	<0.001
Log (size): diet: species	8	0.12	0.02	0.14	3.70	7.54	<0.001
Residuals	43	0.17	0.00	0.20			
Total	62	0.86					

**Table 5** Table showing the pairwise relationships for vector angle of species allometric trajectories between species based on a model of unique species allometries.  $r$  = correlation coefficient

	$r$	angle	UCL (95%)	Z	P value
<i>Caretta caretta</i> : <i>Chelonia mydas</i>	0.753	0.717	0.536	4.562	< <b>0.001</b>
<i>Caretta caretta</i> : <i>Dermochelys coriacea</i>	0.603	0.922	0.518	8.068	< <b>0.001</b>
<i>Caretta caretta</i> : <i>Eretmochelys imbricata</i>	0.640	0.876	0.733	3.31	<b>0.004</b>
<i>Caretta caretta</i> : <i>Lepidochelys kempii</i>	0.407	1.151	1.660	- 1.357	0.91
<i>Caretta caretta</i> : <i>Lepidochelys olivacea</i>	0.809	0.627	0.556	2.815	<b>0.009</b>
<i>Caretta caretta</i> : <i>Natator depressus</i>	0.757	0.710	0.715	1.773	0.053
<i>Chelonia mydas</i> : <i>Dermochelys coriacea</i>	0.509	1.036	0.552	8.806	< <b>0.001</b>
<i>Chelonia mydas</i> : <i>Eretmochelys imbricata</i>	0.719	0.767	0.758	1.946	<b>0.043</b>
<i>Chelonia mydas</i> : <i>Lepidochelys kempii</i>	0.264	1.303	1.670	- 0.475	0.674
<i>Chelonia mydas</i> : <i>Lepidochelys olivacea</i>	0.744	0.731	0.594	3.64	<b>0.002</b>
<i>Chelonia mydas</i> : <i>Natator depressus</i>	0.750	0.721	0.738	1.629	<b>0.065</b>
<i>Dermochelys coriacea</i> : <i>Eretmochelys imbricata</i>	0.466	1.084	0.744	5.338	< <b>0.001</b>
<i>Dermochelys coriacea</i> : <i>Lepidochelys kempii</i>	0.217	1.351	1.668	- 0.183	0.559
<i>Dermochelys coriacea</i> : <i>Lepidochelys olivacea</i>	0.571	0.962	0.571	7.328	< <b>0.001</b>
<i>Dermochelys coriacea</i> : <i>Natator depressus</i>	0.655	0.855	0.724	3.228	<b>0.004</b>
<i>Eretmochelys imbricata</i> : <i>Lepidochelys kempii</i>	0.257	1.310	1.690	- 0.49	0.679
<i>Eretmochelys imbricata</i> : <i>Lepidochelys olivacea</i>	0.838	0.576	0.773	-0.141	0.507
<i>Eretmochelys imbricata</i> : <i>Natator depressus</i>	0.652	0.860	0.876	1.679	0.063
<i>Lepidochelys kempii</i> : <i>Lepidochelys olivacea</i>	0.353	1.208	1.669	- 1.035	0.844
<i>Lepidochelys kempii</i> : <i>Natator depressus</i>	0.332	1.231	1.689	- 0.944	0.822
<i>Lepidochelys olivacea</i> : <i>Natator depressus</i>	0.688	0.811	0.751	2.449	<b>0.018</b>

Angle is the difference in slope angle in radians between species. UCL shows the upper confidence limits of angles from the distributions of pairwise angles

### Cheloniidae (n = 55)

Both phylogenetic and non-phylogenetic comparative methods recover a relatively weak but significant relationship between shape and size ( $p < 0.001$ ; Fig. 7A, B; Table 7). When using phylogenetic comparative methods there was a stronger correlation between size, species, and shape ( $R^2 = 0.13$ ) than when using a non-phylogenetic method ( $R^2 = 0.08$ ). There is a significant relationship (Table 8) between shape and diet using both methods ( $p < 0.001$ ) and both found a significant ( $p < 0.003$ ) but weak interaction between diet, shape ( $R^2 = 0.07$ ). Both phylogenetic and non-phylogenetic comparative methods found that a species unique allometry was significantly stronger than a common allometry model ( $p < 0.001$ ; Table 4). Both methods found that there was no significant difference in model strength between diet and shape or species and shape as possible explanators for shape, but both are significantly better than size alone ( $p < 0.001$ ). The Pearson correlation tests found that only PC1 was significantly related to size ( $p < 0.001$ ).



**Table 6** Table showing the pairwise relationships for vector length of species allometric trajectories based on a model of unique species allometries

		d	UCL (95%)	Z	P value
<i>Caretta caretta</i>	: <i>Chelonia mydas</i>	9.97E–05	1.39E–04	– 0.017	0.499
<i>Caretta caretta</i>	: <i>Dermochelys coriacea</i>	6.03E–05	4.10E–05	3.465	<b>0.002</b>
<i>Caretta caretta</i>	: <i>Eretmochelys imbricata</i>	1.58E–04	1.88E–04	0.995	0.149
<i>Caretta caretta</i>	: <i>Lepidochelys kempii</i>	1.19E–03	1.75E–03	– 0.291	0.568
<i>Caretta caretta</i>	: <i>Lepidochelys olivacea</i>	2.00E–04	2.64E–04	– 0.423	0.652
<i>Caretta caretta</i>	: <i>Natator depressus</i>	1.04E–05	1.24E–04	– 1.961	0.986
<i>Chelonia mydas</i>	: <i>Dermochelys coriacea</i>	1.60E–04	1.50E–04	2.088	<b>0.020</b>
<i>Chelonia mydas</i>	: <i>Eretmochelys imbricata</i>	5.85E–05	9.22E–05	0.82	0.180
<i>Chelonia mydas</i>	: <i>Lepidochelys kempii</i>	1.09E–03	1.65E–03	– 0.289	0.567
<i>Chelonia mydas</i>	: <i>Lepidochelys olivacea</i>	1.00E–04	1.70E–04	– 0.364	0.635
<i>Chelonia mydas</i>	: <i>Natator depressus</i>	1.10E–04	8.54E–05	2.816	<b>0.007</b>
<i>Dermochelys coriacea</i>	: <i>Eretmochelys imbricata</i>	2.19E–04	1.99E–04	2.347	<b>0.021</b>
<i>Dermochelys coriacea</i>	: <i>Lepidochelys kempii</i>	1.25E–03	1.76E–03	– 0.103	0.491
<i>Dermochelys coriacea</i>	: <i>Lepidochelys olivacea</i>	2.60E–04	2.76E–04	1.204	0.114
<i>Dermochelys coriacea</i>	: <i>Natator depressus</i>	4.99E–05	1.36E–04	– 0.997	0.844
<i>Eretmochelys imbricata</i>	: <i>Lepidochelys kempii</i>	1.03E–03	1.63E–03	– 0.424	0.624
<i>Eretmochelys imbricata</i>	: <i>Lepidochelys olivacea</i>	4.17E–05	1.61E–04	– 1.211	0.872
<i>Eretmochelys imbricata</i>	: <i>Natator depressus</i>	1.69E–04	1.28E–04	2.962	<b>0.009</b>
<i>Lepidochelys kempii</i>	: <i>Lepidochelys olivacea</i>	9.85E–04	1.54E–03	– 0.241	0.550
<i>Lepidochelys kempii</i>	: <i>Natator depressus</i>	1.20E–03	1.68E–03	0.014	0.446
<i>Lepidochelys olivacea</i>	: <i>Natator depressus</i>	2.10E–04	2.07E–04	1.713	<b>0.042</b>

d is the distance between least square means of two species. UCL is the upper confidence limit for the distance value. UCL shows the upper confidence limits of distance from the distributions of pairwise distances

## Evolutionary allometry

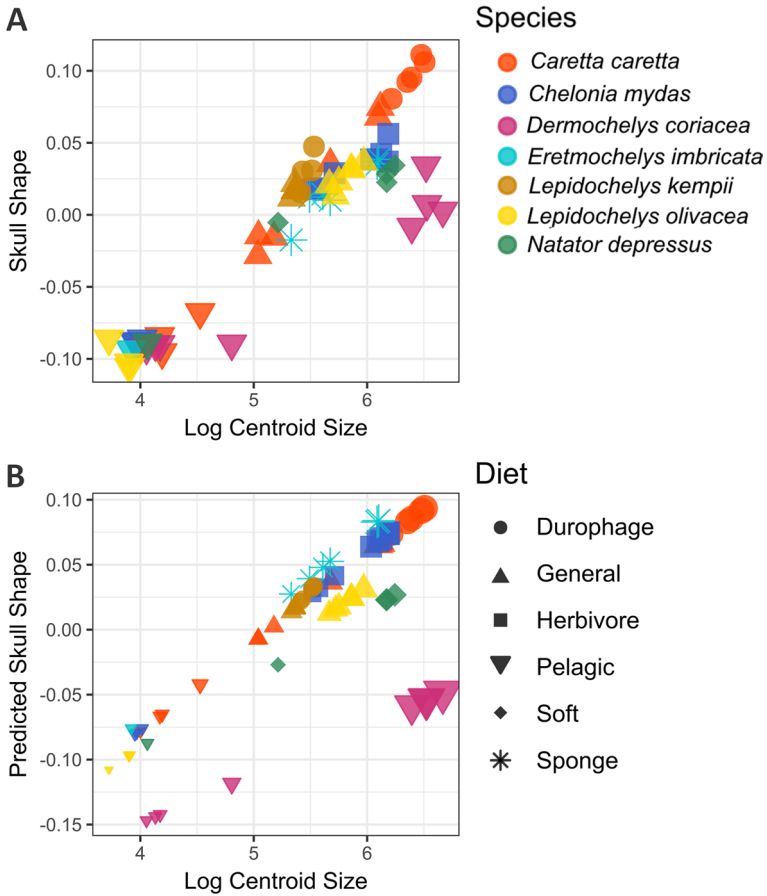
### Chelonioidae adults (n = 21)

Both phylogenetic ( $p=0.015$ ) and non-phylogenetic comparative ( $p<0.001$ ) methods found a significant relationship between size and shape but neither with particularly strong correlations ( $R^2=0.09$  and  $0.12$  respectively; Table 9; Fig. 7C). However, only the phylogenetic comparative ANOVA found a significant interaction between size, shape, and species ( $p=0.005$ ) with a relatively strong correlation ( $R^2=0.42$ ). When using phylogenetic comparative methods a species unique allometry model was significantly stronger than a common allometry model ( $p=0.005$ ). Using the non-phylogenetic methods we found that a species unique model was not significantly stronger than a common allometric model ( $p=0.167$ ). No individual PC axis was significantly correlated with size.

### Cheloniidae adults only (n = 17)

A significant relationship between size and cranial shape was also recovered for both models when *D. coriacea* was removed from the adult sample (Fig. 7D; Table 10). Both





**Fig. 6** **A** Multivariate regression of skull shape based on a common allometry against size plotted on log centroid size. **B** The predicted skull shape based on the regression of skull shape on size of all species based on species allometry. Different colours represent different species, different shapes represent different diets. All points scaled by centroid size

phylogenetic ( $p=0.016$ ) and non-phylogenetic ( $p<0.001$ ) comparative methods found a significant relationship between size and shape. However, only the phylogenetic comparative ANOVA found a significant interaction between size, shape, and species with a relatively strong correlation ( $R^2=0.47$ ,  $p=0.005$ ), with a species unique allometry being the favoured model ( $p=0.005$ ). Using the non-phylogenetic methods, we found that a species unique model was not significantly stronger than a common allometric model ( $p=0.065$ ). The Pearson correlation test found PC1 and PC2 were both significantly related to size (PC1  $p=0.049$ ; PC2  $p=0.009$ ). No other PC axis was found to have a significant relationship with size.

**Phylomorphospace** For the mean shapes of hatchlings (Fig. 8A), PC1 represents 47.9% of the shape variation and PC2 represents 20.4% of the shape variation (68.3% for both combined). For the mean shapes of the adults (Fig. 8B), PC1 represents 50.5% of the shape variation and PC2 represents 21.6% of the shape variation (72.1% for both combined). The phylogenetic

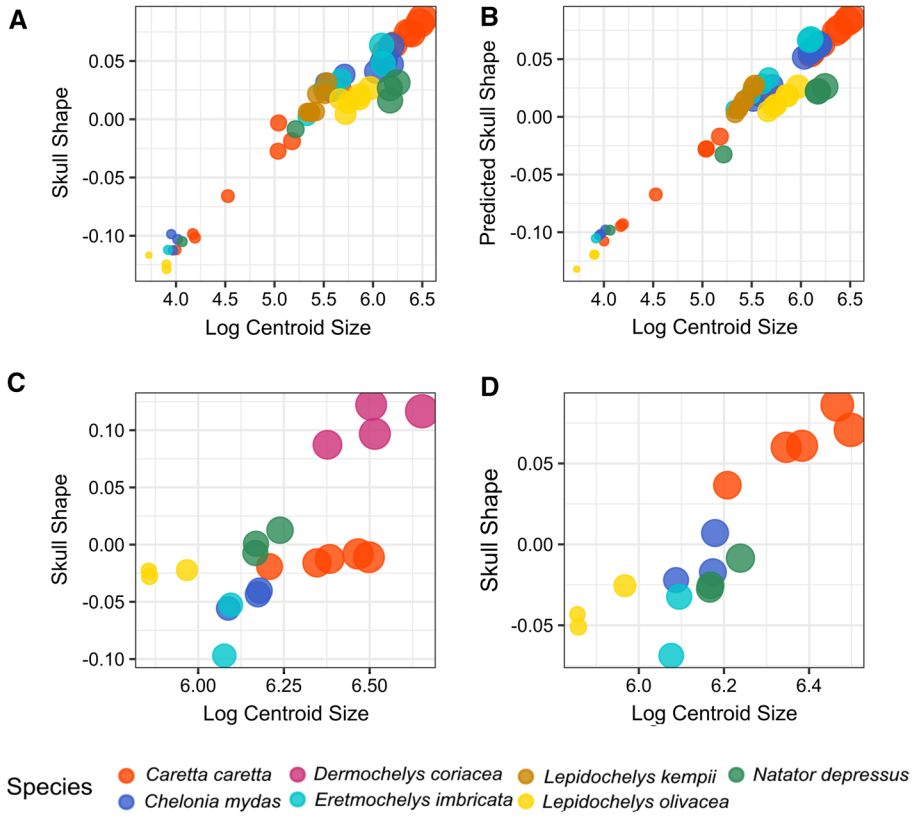
**Table 7** Procrustes ANOVAs for the Cheloniidae all age classes dataset assessing species interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsq	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	1.15	1.15	0.43	44.55	3.93	<0.001
Species	5	0.06	0.01	0.02	0.45	5.55	1
Log (size): species	5	0.34	0.07	0.13	2.65	5.04	<0.001
Residuals	43	1.11	0.03	0.42			
Total	54	2.66					
<i>procD.lm</i>							
Log (size)	1	0.20	0.20	0.32	47.84	5.58	<0.001
Species	5	0.19	0.04	0.31	9.40	8.23	<0.001
Log (size): species	5	0.05	0.01	0.08	2.42	4.67	<0.001
Residuals	43	0.18	0.00	0.29			
Total	54	0.62					

**Table 8** Procrustes ANOVAs for the Cheloniidae all age classes dataset assessing diet interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsq	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	1.15	1.15	0.43	47.30	3.99	<0.001
Diet	5	0.37	0.07	0.14	3.00	6.16	<0.001
Log (size): diet	5	0.19	0.04	0.07	1.55	2.76	0.003
Log (size): diet: species	7	0.08	0.01	0.03	0.48	- 5.54	1
Residuals	36	0.88	0.02	0.33			
Total	54	2.66					
<i>procD.lm</i>							
Log (size)	1	0.20	0.20	0.32	52.32	5.66	<0.001
Diet	5	0.19	0.04	0.30	9.97	8.88	<0.001
Log (size): diet	5	0.04	0.01	0.07	2.20	5.07	<0.001
Log (size): diet: species	7	0.06	0.01	0.09	2.12	6.14	<0.001
Residuals	36	0.14	0.00	0.22			
Total	54	0.62					

signal for the adult specimens was not significant ( $K=0.62$ ;  $p=0.077$ ), however the signal for the hatchlings was ( $K=0.72$ ;  $p=0.011$ ). There is also a visible difference between the shape of the phylomorphospace when plotted separately for hatchlings and adults. The hatchlings plot in a manner consistent with the accepted phylogenetic hypothesis for sea turtles (Fig. 1): *D. coriacea* is relatively isolated in phylomorphospace with a high PC1 score, *N. depressus* and *Ch. mydas* plot close to one another with moderate PC1 scores and low PC2 scores, whereas *E. imbricata*, and *Ca. caretta*+*L. olivacea* plot with high PC2 scores and low PC1 scores. The adults plot with less adherence to the phylogenetic tree. Species which are more distantly related (i.e., *N. depressus* and *L. olivacea*, and *Ch. mydas* and *E. imbricata*) plot with similar scores for PC1 and PC2, whereas the closely related *Ca. caretta* and *L. olivacea*, plot relatively far apart. When only the adults are plotted, PC1 represents a similar aspects of variation to PC1



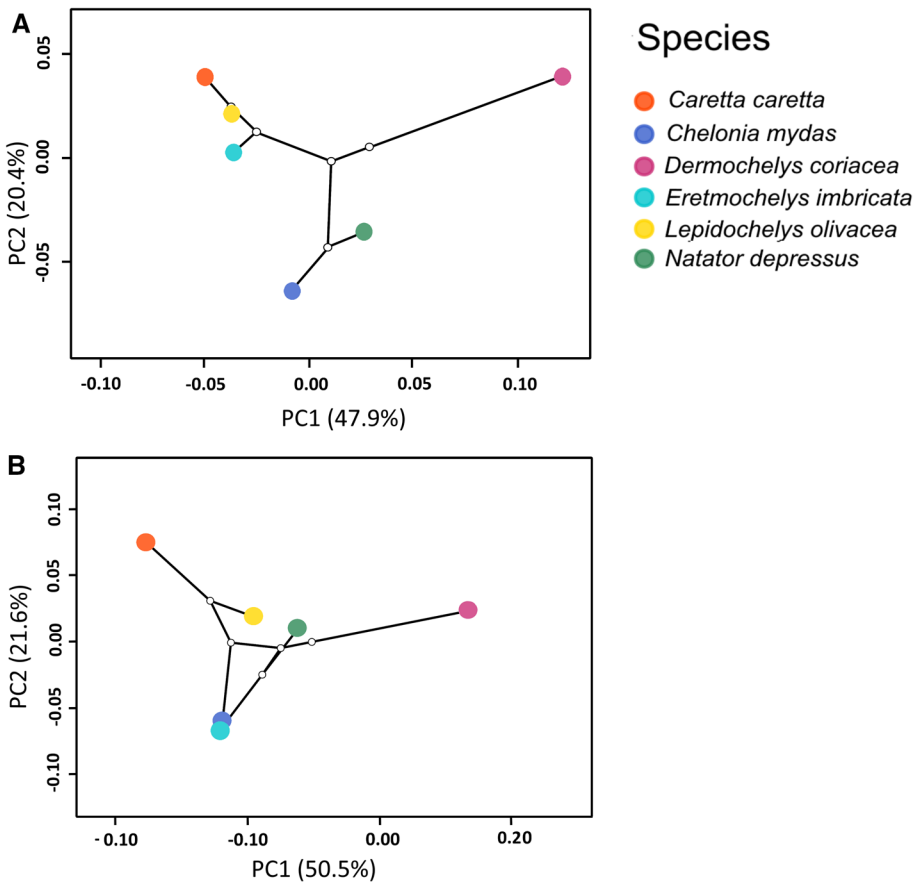
**Fig. 7** Allometric regressions for three of the datasets. A shows log size compared to the regression score for shape for Cheloniidae. B shows the predicted shape values compared to size based on a unique species allometry model for Cheloniidae. C shows the log size compared to the regression score for shape for the adults of Cheloniodea. D shows the log size compared to the regression score for shape for the adults of Cheloniidae all points scaled by centroid size

**Table 9** Procrustes ANOVAs for the Cheloniodea adult dataset assessing species interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsqr	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	0.06	0.06	0.09	2.30	2.16	0.014
Species	5	0.13	0.03	0.19	1.02	0.10	0.460
Log (size): species	5	0.29	0.06	0.42	2.19	2.42	0.005
Residuals	8	0.21	0.03	0.30			
Total	19	0.69					
<i>procD.lm</i>							
Log (size)	1	0.03	0.03	0.12	7.98	3.26	<0.001
Species	5	0.18	0.04	0.67	8.87	5.96	<0.001
Log (size): species	5	0.02	0.00	0.09	1.22	0.95	0.170
Residuals	8	0.03	0.00	0.12			
Total	19	0.27					

**Table 10** Procrustes ANOVAs for the Cheloniidae all age classes dataset assessing species interaction using both phylogenetic (*procD.pgls*) and non-phylogenetic (*procD.lm*) comparative methods

	Df	SS	MS	Rsq	F	Z	P
<i>procD.pgls</i>							
Log (size)	1	0.04	0.04	0.10	2.52	2.16	0.016
Species	4	0.08	0.02	0.19	1.23	0.60	0.270
Log (size): species	4	0.20	0.05	0.47	3.00	2.51	0.005
Residuals	6	0.10	0.02	0.24			
Total	15	0.42					
<i>procD.lm</i>							
Log (size)	1	0.03	0.03	0.17	8.20	4.27	<0.001
Species	4	0.09	0.02	0.58	6.87	4.87	<0.001
Log (size): species	4	0.02	0.00	0.12	1.48	1.50	0.065
Residuals	6	0.02	0.00	0.13			
Total	15	0.15					



**Fig. 8** **A** Phylomorphospace of mean shape of hatchling specimens. **B** Phylomorphospace of mean shape of adult specimens. Different colours represent different species. White circles represent shape of hypothetical most recent common ancestor of the clade based on maximum likelihood estimation

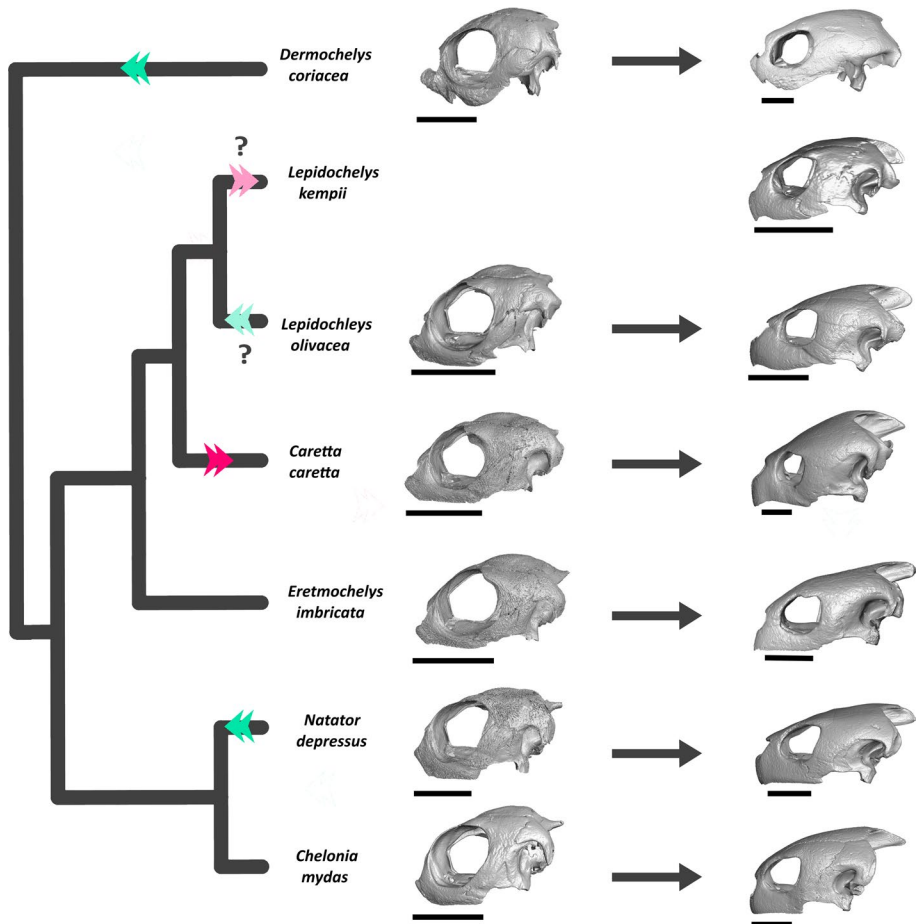
for the entire dataset (relative size of the orbits and posterior part of the skull) whereas while PC2 is similar to PC3 of the complete dataset (posterior part of the skull width and depth).

## Discussion

Our results show that much of the diversity of the skull of modern sea turtles can be explained by variation in size, including both evolutionary and ontogenetic patterns of allometry. We find that hatchling sea turtles have skull shapes that reflect the phylogenetic relationships identified and established by molecular studies (Naro-Maciel et al. 2008; Duchene et al. 2012). From this starting point, differential growth appears to be an important factor in the determination of the adult skull shape. We found that shape and phylogenetic relationships become less obviously linked and diet becomes at least as important a predictor of adult skull shape as phylogeny. This result does not have high statistical power because the number of diet categories and species are similar, but a pattern is evident. In contrast to Myers (2007), we do not find that *Lepidochelys olivacea* has a highly divergent skull shape from other sea turtles. Instead, our results are consistent with previous studies on individual sea turtle species that infer ontogenetic changes in diet are associated with changes in skull shape (Nishizawa et al. 2010; Lunardon et al. 2020). Variations in ontogenetic changes in skull shape are associated with variation in diet such that multiple instances of paedomorphosis and peramorphosis may be due to divergences in adult diet (Fig. 9).

We find that diversity in adult skull shape corresponds to diversity in diet. The relationship appears to be stronger than found in some other groups of amniotes (Maestri et al. 2016; Bright et al. 2016; Gray et al. 2019). The two sea turtle species with the most divergent diets, *Chelonia mydas* and *Eretmochelys imbricata*, show relatively clear adaptations associated with their dietary category of herbivore and sponge, respectively. The short rounded snout of *Ch. mydas* is similar to other aquatic grazing amniotes such as sirenians (Marshall et al. 2012; Aragonés et al. 2012) and marine iguanas (Wikeleski and Trillmich 1994). In contrast, the lack of other spongivorous amniotes makes comparisons with *E. imbricata* difficult. However, its long and narrow snout appears as a likely adaptation to reaching into narrow crevices in coral reefs where sponges often grow (Hill 1998; Figgenger et al. 2019), analogous to selective terrestrial browsers which tend to have narrow snouts (Solounias and Moelleken 1993; Dompierre and Churcher 1996). Neither of these morphotypes are associated with size and instead appear to be adaptations to highly specialised diets. The other carnivorous cheloniids have skull shapes characterised by features strongly associated with ontogenetic allometry.

Shape variation accompanying increased body size in sea turtles is most strongly associated with a relative expansion of structures associated with jaw musculature (Jones et al. 2012). The clearest example of this is *Caretta caretta*, in which allometric skull changes include enlarged temporal regions and jaw muscles that provide greater bite force allowing for diets consisting largely of hard shelled invertebrates (Claude et al. 2004; Huber et al. 2005; Jones et al. 2012; Marshall et al. 2012; Figueirido et al. 2013; Figueirido et al. 2013). This pattern of increased size and development of the jaw muscles and associated structures in comparison to the similarly sized *Ch. mydas* (Pritchard and Trebbau 1984) is consistent with “acceleration” peramorphosis (Klingenberg 1998), as seen in multiple other vertebrate groups (Denoel et al. 2004; Herrel and O’Reilly 2006; Chemisquy 2015; Vita et al. 2020). Within turtles, this phenotype (with large adductor chambers and deep jaws)



**Fig. 9** Cladogram of Chelonioida and the change in skull morphology from hatchling to adults for each species, with the exception of *Lepidochelys kempii* which is represented by an intermediate individual. Arrows on the tree indicate inferred heterochronic process according to our hypothesis. Green arrows on the tree represent instances of paedomorphy, pink arrows represent instances of peramorphy. Question marks represent the uncertainty of the processes involved for the two species of *Lepidochelys*. Scale bars for hatchling specimens (left) = 10mm. Scale bars for adult specimens (right) = 50mm. Specimens shown: *Dermochelys coriacea* (MV D6188, UMZC R3031); *Lepidochelys kempii* (WH 333); *Lepidochelys olivacea* (MV D5797, SAMA BM678); *Caretta caretta* (QM J73517, SAMA Unregistered); *Eretmochelys imbricata* (SAMA R14358, WAM 120,113); *Natator depressus* (SAMA R14360, WAM R112123); *Chelonia mydas* (MV D2987, SAMA Unregistered)

is also consistently associated with durophagy among unrelated species such as *Malayemys subtrijuga* and *Sternotherus odoratus* (Claude et al. 2004; Bever 2009; Parham and Pyenson 2010; Ferreira et al. 2015; Lunardon et al. 2020). The great abundance of extinct cheloniids that have independently acquired durophagous traits, such as those noted in this study as well as robust mandibles with an expanded triturating surface (Gaffney 1979; Parham and Pyenson 2010; Weems and Brown 2017) might be related to a clade-wide ability to pursue peramorphy with relative ease.

The two species of *Lepidochelys* have similar adult sizes but different skull shapes and different diets (Pritchard and Trebbau 1984; Bjørndal et al. 1997). At similar sizes *L. kempii* has a “more mature” skull shape than *L. olivacea*, meaning smaller orbits, a larger temporal region, and a longer supraoccipital crest. As in *Ca. caretta*, these differences are associated with a durophagous diet and may suggest peramorphosis. At the same time, it could be equally parsimonious to suggest that *L. olivacea* is paedomorphic in relation to *L. kempii*. The adult skull shape of *L. olivacea* is similar to that of younger individuals of *Ca. caretta*, which also have a general diet that is less reliant on benthic organisms and better suited to an oceanic rather than coastal lifestyle (Bowen et al. 1997). At present, the small sample of extant taxa and lack of available fossil evidence make it difficult to clearly state which heterochronic patterns explain the shape differences between the two species.

We find that *Dermochelys coriacea* and *Natator depressus* have a skull shape consistent with paedomorphosis, characterised by a small supraoccipital crest and relatively large orbits associated with retention of a juvenile-like diet. However, unlike the other species of sea turtles there does not appear to be a strong functional link between skull shape and diet category, except for the large orbits of *D. coriacea* which may help deep water foraging in low light (Houghton et al. 2008; Horch et al. 2008; Hall 2008; Veilleux and Kirk 2014). Both Nick (1912) and Pritchard and Trebbau (1984) found that the postcranial skeleton of *D. coriacea* also shows evidence of extensive paedomorphosis, suggesting that the paedomorphic skull may be part of a general skeletal pattern rather than the result of selection directly related to diet and the skull itself. Species develop paedomorphic morphologies in multiple evolutionary contexts, including retention of a juvenile life habit (Kordikova 2002), miniaturisation (Rieppel and Crumly 1997; Bright et al. 2016; Esquerré et al. 2017), diet (Denoël et al. 2004; Esquerré et al. 2017; Sherratt et al. 2019), and other complex factors which may be unclear (Bright et al. 2016; Morris et al. 2019; Bardua et al. 2021). Unfortunately, the selective pressures for *D. coriacea* to develop this phenotype are still unknown. The high degree of shell reduction as a consequence of paedomorphosis may assist in achieving a large size given that extensive shell reduction is common among other large sea turtles (Hirayama 1994; Cadena and Parham 2015). There may also be a link between paedomorphy and deep diving, as other deep diving amniotes such the southern elephant seal (*Mirounga leonina*) share some paedomorphic traits with *D. coriacea* such as lack of suture closure Pritchard and Trebbau 1984; Goswami et al. 2013). The paedomorphic skull of *D. coriacea* is likely an example of multiple factors influencing shape including behaviour, habitat, as well as a generally paedomorphic condition rather than just selective pressure on feeding mechanics (Figgenger et al. 2019). In *N. depressus*, paedomorphosis appears to be restricted to the skull (Zangerl et al. 1988), perhaps making paedomorphosis more likely associated with diet in this species. However, this hypothesis requires further investigation.

The oldest (*D. coriacea*-Cheloniidae) and the youngest phylogenetic divergence (*L. kempii*-*L. olivacea*) effectively demonstrate how changes in ontogenetic allometry might accompany speciation, or even act as a mechanism for morphological divergence. Dermochelyids likely diverged from cheloniids in the Late Cretaceous (Duchene et al. 2012; Thomson et al. 2021). Fossil representatives of stem chelonioids from the Cretaceous, such as *Toxochelys*, do not exhibit obvious evidence of paedomorphy (Zangerl 1980; Gentry 2017; Gentry et al. 2019). The fossil dermochelyid *Eosphargis breineri*, from the Eocene of northern continents, provides a glimpse into early dermochelyid skull shape (Nielsen 1959). The high degree of similarity between the skull morphology of *Eo. breineri* (Nielsen 1959; Hirayama 1994) to the modern *D. coriacea* and the lack of other similar

stem chelonoids suggests dermochelyids achieved a paedomorphic skull shape very early in their evolution.

The two species of *Lepidochelys* may be another example of allometric differences associated with different diets because they are sister taxa and a similar absolute size but have different skull shapes and diets. They are estimated to have diverged 2.5–3.5 Ma during the formation of the isthmus of Panama (Bowen et al. 1997; Naro-Maciel et al. 2008; Duchene et al. 2012): the most recent divergence between species of extant sea turtles (Duchene et al. 2012). As these two species appear to have distinct allometries despite diverging so recently, we suggest that heterochrony was likely a mechanism in their morphological diversification. More fossils representing possible ancestors of the *Caretta + Lepidochelys* clade are needed to better understand the exact evolutionary processes leading to the diversity we see today. Candidate fossils are available, however they are incomplete and require more extensive study (Zug and Parham 1996).

As we suggest for sea turtles, heterochrony appears to be an important mechanism for phenotypic diversification for much of Reptilia. Within closely related groups, shifts in allometric trajectories have frequently been found to be of major importance in generating adaptive diversity, whether the phenotype in question is body size (Denoel and Joly 2000; Kon and Tetsuo 2002; Sander et al. 2004; McNamara and Long 2012; Esquerré et al. 2017) or shape change (Leiberman et al. 2007; Adams and Nistri 2010; Piras et al. 2011; Morris et al. 2019; Gray et al. 2019). In other reptiles, heterochrony linked to dietary diversity is also seen in Crocodylia where shifts in allometric trajectories (i.e. deceleration, pre-displacement, acceleration) are perhaps the most significant determinants of skull shape, correlating with dietary niches (Morris et al. 2019). Similarly, in pythons, heterochrony was found to be the main mechanism in initial divergence of phenotype, with a strong adaptive link between ontogenetic allometry and ecology (Esquerré et al. 2017). Though the exact genetic mechanisms for various heterochronic patterns are varied depending on taxon and body region (Brugmann et al. 2006; Tokita et al. 2017; Bhullar et al. 2015; Ahi 2016; Morris et al. 2019), and are yet to be established for sea turtles, our study has shown that changes to the timing and extent of development have played a major role in generating sea turtle diversity.

Though our data are highly suggestive of allometry being an important factor in phenotypic diversification in modern sea turtles, the high dietary but low taxonomic diversity as well as small sample size make our results less certain. The almost one to one ratio of dietary specialisation to species is one of the most interesting aspects of sea turtle evolution. However, this condition make establishing any ties between skull shape and diet with any statistical power difficult (Blomberg et al. 2003; Adams 2014b). The challenges of collecting large sample size for some species, in particular *L. kempii*, also means that our findings are preliminary and require more extensive investigation.

## Conclusion

Heterochrony appears to play a significant role in shaping the sea turtle skull. We infer there to be at least two instances of paedomorphy and one instance of peramorphy among extant sea turtles. These instances are associated with differences in diet, suggesting that shifts in dietary niche may be an agent of selection: robust peramorphic skulls with large muscle chambers are associated with hard shelled prey whereas paedomorphic skulls with large orbits are associated with soft bodied prey. However, the paedomorphic skull shape



of *D. coriacea* is part of an overall paedomorphic body plan. Therefore, it may not reflect selective pressures to feed on soft bodied prey per se. Regardless of whether dietary differences are a driver of skull shape change in sea turtles or an incidental consequence, heterochrony is likely to be an important mechanism for facilitating such changes. Our study provides another example among Craniata of the importance of ontogenetic allometry as a mechanism for generating morphological variation.

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## Declarations

**Conflict of interest** The authors declare that they have no conflict of interest.

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## References

- Adams DC (2014) A generalized K statistic for estimating phylogenetic signal from shape and other high-dimensional multivariate data. *Syst Biol* 63:685–697
- Adams DC (2014b) A method for assessing phylogenetic least squares models for shape and other high-dimensional multivariate data. *Evolution* 68:2675–2688
- Adams DC, Collyer ML (2015) Permutation tests for phylogenetic comparative analyses of high-dimensional shape data: what you shuffle matters. *Evolution* 69:823–829
- Adams DC, Nistri A (2010) Ontogenetic convergence and evolution of foot morphology in European cave salamanders (Family: Plethodontidae). *BMC Evol Biol* 10:1–10
- Adams DC, Rohlf FJ, Slice DE (2013) A field comes of age: geometric morphometrics in the 21st century. *Hystrix* 24:7
- Adams D, Collyer M, Kaliontzopoulou A (2020) Geometric morphometric analyses of 2D/3D landmark data

- Ahi EP (2016) Signalling pathways in trophic skeletal development and morphogenesis: insights from studies on teleost fish. *Dev Biol* 420:11–31
- Anderes B, Uchida I (1994) Study of the hawksbill turtle (*Eretmochelys imbricata*) stomach content in Cuban waters. *Study of the Hawksbill Turtle in Cuba*
- Aragones LV, Lawler I, Marsh H et al (2012) The role of sirenians in aquatic ecosystems. *Sirenian conservation: issues and strategies in developing countries*. University Press of Florida, Gainesville, FL, pp 4–11
- Babcock HL (1938) The Sea-Turtles of the Bermuda Islands, with a Survey of the present state of the Turtle Fishing Industry. In: *Proceedings of the zoological society of london*. Wiley Online Library, p 595–602
- Bardua C, Fabre A-C, Clavel J et al (2021) Size, microhabitat, and loss of larval feeding drive cranial diversification in frogs. *Nat Comm* 12:2503
- Bever GS (2009) Postnatal Ontogeny of the Skull in the Extant North American Turtle *Sternotherus odoratus* (Cryptodira: Kinosternidae). *Bull Am Mus Nat Hist* 330:1–97
- Bhullar B-aS, Morris ZS, Sefton EM et al (2015) A molecular mechanism for the origin of a key evolutionary innovation, the bird beak and palate, revealed by an integrative approach to major transitions in vertebrate history. *Evolution* 69:1665–1677
- Bhullar BS, Hanson M, Fabbri M et al (2016) How to make a bird skull: major transitions in the evolution of the avian cranium, paedomorphosis, and the beak as a surrogate hand. *Integr Comp Biol* 56:389–403
- Bjorndal KA (1985) Nutritional ecology of sea turtles. *Copeia* 1985:736–751
- Bjorndal KA, Lutz P, Musick J (1997) Foraging ecology and nutrition of sea turtles. *Biol Sea Turtles* 1:199–231
- Bleakney JS (1965) Reports of marine turtles from New England and eastern Canada. *Can Field-Nat* 79:120–128
- Blomberg SP, Garland T Jr, Ives AR (2003) Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* 57:717–745
- Bolten AB (2003) Variation in sea turtle life history patterns: neritic vs. oceanic developmental stages. In: Lutz PL, Musick JA, Wyneken J (eds) *The biology of sea turtles*, vol 2. CRC Press, Boca Raton, Florida, pp 243–257
- Bowen BW, Clark AM, Abreu-Grobois FA et al (1997) Global phylogeography of the ridley sea turtles (*Lepidochelys* spp.) as inferred from mitochondrial DNA sequences. *Genetica* 101:179–189
- Boyle MC, Limpus CJ (2008) The stomach contents of post-hatchling green and loggerhead sea turtles in the southwest Pacific: an insight into habitat association. *Mar Biol* 155:233–241
- Brandt LA (1991) Growth of juvenile alligators in Par Pond, Savannah River Site, South Carolina. *Copeia* 1991:1123–1129
- Bright JA, Marugán-Lobón J, Cobb SN et al (2016) The shapes of bird beaks are highly controlled by nondietary factors. *Proc Nat Acad Sci* 113:5352–5357
- Brugmann SA, Kim J, Helms JA (2006) Looking different: understanding diversity in facial form. *Am J Med Gen A* 140A:2521–2529
- Burke VJ, Standora EA, Morreale SJ (1993) Diet of juvenile Kemp's ridley and loggerhead sea turtles from Long Island, vol 1993. *Copeia*, New York, pp 1176–1180
- Burke VJ, Morreale SJ, Standora EA (1994) Diet of the Kemp's ridley sea turtle, *Lepidochelys kempii*, in New York waters. *Fish Bull* 92:26–32
- Cadena EA, Parham JF (2015) Oldest known marine turtle? A new protostegid from the Lower Cretaceous of Colombia. *PaleoBios* 32:1–42
- Chemisquy MA (2015) Peramorphic males and extreme sexual dimorphism in *Monodelphis dimidiata* (Didelphidae). *Zoomorphology* 134:587–599
- Claude J, Pritchard PCH, Tong HY et al (2004) Ecological correlates and evolutionary divergence in the skull of turtles: a geometric morphometric assessment. *Syst Biol* 53:933–948
- Coelho VF, Domit C, Broadhurst MK et al (2018) Intra-specific variation in skull morphology of juvenile *Chelonia mydas* in the southwestern Atlantic Ocean. *Mar Biol* 165:1–12
- Colman LP, Sampaio CLS, Weber MI et al (2014) Diet of olive ridley sea turtles, *Lepidochelys olivacea*, in the waters of Sergipe, Brazil. *Chelonian Con Biol* 13:266–271
- Collyer M, Adams D, Collyer MM (2020) Package 'RRPP&#8217
- Den Hartog J (1979) Notes on the food of sea turtles: *Eretmochelys imbrica* Ta (Linnaeus) and *Dermochelys Coriacea* (Linnaeus). *Neth J Zool* 30:595–611
- Denoël M, Andreone F (2003) Trophic habits and aquatic microhabitat use in gilled immature, paedomorphic and metamorphic Alpine newts (*Triturus alpestris apuanus*) in a pond in central Italy. *Belg J Zool* 133:95–102

- Denoël M, Joly P (2000) Neoteny and progenesis as two heterochronic processes involved in paedomorphosis in *Triturus alpestris* (Amphibia: Caudata). *Proc Roy Soc B* 267:1481–1485
- Denoël M, Schabetsberger R, Joly P (2004) Trophic specialisations in alternative heterochronic morphs. *Naturwissenschaften* 91:81–84
- Dodd CKJ (1988) Synopsis of the biological data on the loggerhead sea turtle *Caretta caretta* Linnaeus 1758. U S Fish and Wildlife Serv Biol Rep 88:I-VIII,1–110
- Dompierre H, Churcher CS (1996) Premaxillary shape as an indicator of the diet of seven extinct late Cenozoic new world camels. *J Vertebr Paleontol* 16:141–148
- Duchene S, Frey A, Alfaro-Nunez A et al (2012) Marine turtle mitogenome phylogenetics and evolution. *Mol Phylogenet Evol* 65:241–250
- Eckert SA, Eckert KL, Ponganis P et al (1989) Diving and foraging behavior of leatherback sea turtles (*Dermodochelys coriacea*). *Can J Zool* 67:2834–2840
- Erickson GM, Lappin AK, Vliet KA (2003) The ontogeny of bite-force performance in American alligator (*Alligator mississippiensis*). *J Zool* 260:317–327
- Esquerré D, Sherratt E, Keogh JS (2017) Evolution of extreme ontogenetic allometric diversity and heterochrony in pythons, a clade of giant and dwarf snakes. *Evolution* 71:2829–2844
- Evers SW, Barrett PM, Benson RBJ (2019) Anatomy of *Rhinochelys pulchriiceps* (Protostegidae) and marine adaptation during the early evolution of chelonoids. *PeerJ* 7:e6811
- Ferreira GS, Rincón AD, Solórzano A et al (2015) The last marine pelomedusoids (Testudines: Pleurodira): a new species of *Bairdemys* and the paleoecology of Stereogenyina. *PeerJ* 3:e1063
- Figgeneger C, Bernardo J, Plotkin PT (2019) Beyond trophic morphology: stable isotopes reveal ubiquitous versatility in marine turtle trophic ecology. *Biol Rev* 94:1947–1973
- Figueirido B, Tseng ZJ, Martín-Serra A (2013) Skull shape evolution in durophagous carnivorans. *Evolution* 67:1975–1993
- Foth C, Rabi M, Joyce WG (2017) Skull shape variation in extant and extinct Testudinata and its relation to habitat and feeding ecology. *Acta Zool* 98:310–325
- Fowler de Neira LE, Johnson MK (1985) Diets of giant tortoises and feral burros on Volcan Alcedo, Galapagos. *J Wildl Management* 49:165–169
- Frazier J, Margartoulis D, Muldoon K et al (1985) Epizoan communities on marine turtles: I. Bivalve and Gastropod mollusks. *Mar Ecol* 6:127–140
- Frederich B, Adriaens D, Vandewalle P (2008) Ontogenetic shape changes in Pomacentridae (Teleostei, Perciformes) and their relationships with feeding strategies: a geometric morphometric approach. *Biol J Linn Soc* 95:92–105
- Frick MG, Kopitsky K, Bolten AB et al (2011) Sympatry in grapsoid crabs (genera *Planes* and *Plagusia*) from olive ridley sea turtles (*Lepidochelys olivacea*), with descriptions of crab diets and masticatory structures. *Mar Biol* 158:1699–1708
- Furrer SC, Hatt JM, Snell H et al (2004) Comparative study on the growth of juvenile Galapagos giant tortoises (*Geochelone nigra*) at the Charles Darwin Research Station (Galapagos Islands, Ecuador) and Zoo Zurich (Zurich, Switzerland). *Zoo Biol* 23:177–183
- Gaffney ES (1979) Comparative cranial morphology of Recent and fossil turtles. *Bull Am Mus Nat Hist* 164:65–376
- Gentry AD (2017) New material of the Late Cretaceous marine turtle *Ctenochelys acris* Zangerl, 1953 and a phylogenetic reassessment of the ‘toxocheilyd’-grade taxa. *J Syst Palaeontol* 15:675–696
- Gentry AD, Ebersole JA, Kiernan CR (2019) *Asmodochelys parhami*, a new fossil marine turtle from the Campanian Demopolis Chalk and the stratigraphic congruence of competing marine turtle phylogenies. *R Soc Open Sci* 6:191950
- Gibson CWD, Hamilton J (1983) Feeding ecology and seasonal movements of giant tortoises on Aldabra atoll. *Oecologia* 56:84–92
- Goswami A, Foley L, Weisbecker V (2013) Patterns and implications of extensive heterochrony in carnivoran cranial suture closure. *J Evol Biol* 26:1294–1306
- Gray JA, Sherratt E, Hutchinson MN et al (2019) Changes in ontogenetic patterns facilitate diversification in skull shape of Australian agamid lizards. *BMC Evol Biol* 19:7
- Hall MI (2008) Comparative analysis of the size and shape of the lizard eye. *Zoology* 111:62–75
- Herrel A, O’reilly JC (2006) Ontogenetic scaling of bite force in lizards and turtles. *Phys Biochem Zool* 79:31–42
- Hill MS (1998) Spongivory on Caribbean reefs releases corals from competition with sponges. *Oecologia* 117:143–150
- Hirayama R (1994) Phylogenetic systematics of chelonoid sea turtles. *Isl Arc* 3:270–284

- Horch KW, Gocke JP, Salmon M et al (2008) Visual spectral sensitivity of hatchling loggerhead (*Caretta caretta* L.) and leatherback (*Dermochelys coriacea* L.) sea turtles, as determined by single-flash electroretinography. *Mar Freshw Behav Physiol* 41:107–119
- Houghton JDR, Doyle TK, Davenport J et al (2008) The role of infrequent and extraordinary deep dives in leatherback turtles (*Dermochelys coriacea*). *J Exp Biol* 211:2566–2575
- Huber DR, Eason TG, Hueter RE et al (2005) Analysis of the bite force and mechanical design of the feeding mechanism of the durophagous horn shark *Heterodontus francisci*. *J Exp Biol* 208:3553–3571
- Jones MEH, Werneburg I, Curtis N et al (2012) The head and neck anatomy of sea turtles (Cryptodira: Chelonioida) and skull shape in Testudines. *PLoS ONE* 7:e47852
- Jones TT, Hastings MD, Bostrom BL et al (2011) Growth of captive leatherback turtles, *Dermochelys coriacea*, with inferences on growth in the wild: implications for population decline and recovery. *J Exp Mar Biol Ecol* 399:84–92
- Kamezaki N, Matsui M (1995) Geographic-variation in skull morphology of the green turtle, *Chelonia mydas*, with a taxonomic discussion. *J Herpetol* 29:51–60
- Kamezaki N, Matsui M (1997) Allometry in the loggerhead turtle, *Caretta caretta*. *Chelonian Conserv Biol* 2:421–424
- Kamezaki N, Bolten A, Witherington B (2003) What is a loggerhead turtle? The morphological perspective. In: Bolten AB, Witherington BE (eds) *Loggerhead sea turtles*. Smithsonian Books, Washington, USA, pp 28–43
- Klingenberg CP (1998) Heterochrony and allometry: the analysis of evolutionary change in ontogeny. *Biol Rev* 73:79–123
- Kon T, Yoshino T (2002) Extremely early maturity found in Okinawan gobioid fishes. *Ichthyoll Res* 49:224–228
- Kordikova EG (2002) Heterochrony in the evolution of the shell of Chelonia. Part 1: terminology, Cheloniidae, Dermochelyidae, Trionychidae, Cyclanorbidae and Carettochelyidae. *Neues Jahrbuch für Geologie und Paläontologie-Abhandlungen*:343–417
- Leach G, Delaney R, Fukuda Y (2009) Management program for the saltwater crocodile in the Northern Territory of Australia, 2009–2014. Department of Natural Resources, Environment, the Arts and Sport Darwin, NT
- Lieberman DE, Carlo J, Ponce De León M et al (2007) A geometric morphometric analysis of heterochrony in the cranium of chimpanzees and bonobos. *J Hum Evol* 52:647–662
- Limpus CJ (1992) The hawksbill turtle, *Eretmochelys imbricata*, in Queensland: population structure within a southern Great Barrier Reef feeding ground. *Wildl Res* 19:489–505
- Limpus C, Limpus D (2007) Recruitment of *Eretmochelys imbricata* from the pelagic to the benthic feeding life history phase. The hawksbill turtle, *Eretmochelys imbricata*, in eastern and northern Australia. Queensland Parks and Wildlife Service, Brisbane
- Lunardon EA, Costa-Schmidt LE, Lenz AJ et al (2020) Skull ontogenetic variation of the coastal developmental stage of the loggerhead turtle (*Caretta caretta*) in the western South Atlantic Ocean. *Hydrobiologia* 947:1999–2019
- Maestri R, Patterson BD, Fornel R et al (2016) Diet, bite force and skull morphology in the generalist rodent morphotype. *J Evol Biol* 29:2191–2204
- Marchiori E, Cassini R, Ricci I et al (2018) Qualitative and quantitative methods for estimating Spirorchidiasis burden in sea turtles. *Int J Parasitol Parasites Wildl* 7:409–414
- Márquez R (1994) Synopsis of biological data on the Kemp's ridley turtle, *Lepidochelys kempi*, vol 1880. US Department of Commerce, National Oceanic and Atmospheric Administration, Garman
- Márquez R, Peñaflores C (1976) Sinopsis de datos biológicos sobre la tortuga golfinalepidochelys olivacea eschscholtz, 1829. In, No. F/598.13 M3
- Marshall CD, Guzman A, Narazaki T, Sato K, Kane EA, Sterba-Boatwright BD (2012) The ontogenetic scaling of bite force and head size in loggerhead sea turtles (*Caretta caretta*): implications for durophagy in neritic, benthic habitats. *J Exp Biol* 215(23):4166–4174
- McNamara KJ, Long JA (2012) The role of heterochrony in dinosaur evolution. The complete dinosaur. Life of the Past. Indiana University press, Indiana, pp 761–784
- Meylan AB (1984) Feeding ecology of the hawksbill turtle (*Eretmochelys imbricata*): spongivory as a feeding niche in the coral reef community. PhD thesis, University of Florida, Gainesville
- Meylan A (1985) The role of sponge collagens in the diet of the hawksbill turtle (*Eretmochelys imbricata*). Biology of invertebrate and lower vertebrate collagens. Springer, Berlin, pp 191–196
- Mitteroecker P, Gunz P, Bookstein FL (2005) Heterochrony and geometric morphometrics: a comparison of cranial growth in *Pan paniscus* versus *Pan troglodytes*. *Evol Dev* 7:244–258

- Montenegro S, Bernal G (1982) Análisis del contenido estomacal de *Lepidochelys olivacea*. Unpublished Thesis Profesional. ENEP Iztacala, Universidad Nacional Autonoma de Mexic
- Morris ZS, Vliet KA, Abzhanov A et al (2019) Heterochronic shifts and conserved embryonic shape underlie crocodylian craniofacial disparity and convergence. *Proc Roy Soc B* 286:20182389
- Mortimer JA (1982) Feeding ecology of sea turtles. Smithsonian Institution Press, Washington D.C.
- Myers T (2007) Osteological morphometrics of Australian chelonioid turtles. *Zool Sci* 24:1012–1027
- Naro-Maciel E, Le M, Fitzsimmons NN et al (2008) Evolutionary relationships of marine turtles: a molecular phylogeny based on nuclear and mitochondrial genes. *Mol Phylogenet Evol* 49:659–662
- Nick L (1912) Das Kopfskelet von *Dermodochelys coriacea* L. *Zool Jb Abt f Anat* 33:1–238
- Nielsen E (1959) Eocene turtles from Denmark. *Bull Geol Soc Denmark* 14:96–114
- Van Nierop MM, Den Hartog JC (1984) A study on the gut contents of live juvenile loggerhead turtles, *Caretta caretta* (Linnaeus) (Reptilia, Cheloniidae), from the south-eastern part of the North Atlantic ocean, with emphasis on coelenterate identification. *Zool Meded* 59:35–54
- Nishizawa H, Asahara M, Kamezaki N et al (2010) Differences in the skull morphology between juvenile and adult green turtles: implications for the ontogenetic diet shift. *Curr Herpetol* 29:97–101
- Paradis E, Schliep K (2019) ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* 35:526–528
- Parham JF, Pyenson ND (2010) New sea turtle from the Miocene of Peru and the iterative evolution of feeding ecomorphologies since the Cretaceous. *J Paleontol* 84:231–247
- Piras P, Salvi D, Ferrara G et al (2011) The role of post-natal ontogeny in the evolution of phenotypic diversity in *Podarcis* lizards. *J Evol Biol* 24:2705–2720
- Prevosti FJ, Turazzini GF, Ercoli MD et al (2011) Mandible shape in marsupial and placental carnivorous mammals: a morphological comparative study using geometric morphometrics. *Zool J Linn Soc* 164:836–855
- Pritchard PCH, Trebbau P (1984) The turtles of venezuela. Society for the Study of Amphibians and Reptiles, Oxford
- Püschel TA, Sellers WI (2016) Standing on the shoulders of apes: analyzing the form and function of the hominoid scapula using geometric morphometrics and finite element analysis. *Am J Phys Anthropol* 159:325–341
- Rieppel O, Crumly C (1997) Paedomorphosis and skull structure in Malagasy chamaeleons (Reptilia: Chamaeleoninae). *J Zool* 243:351–380leas
- Ross JP (1985) Biology of the green turtle, *Chelonia mydas*, on an Arabian feeding ground. *J Herpetol* 19:459–468
- Sander PM, Klein N, Buffetaut E et al (2004) Adaptive radiation in sauropod dinosaurs: bone histology indicates rapid evolution of giant body size through acceleration. *Organism Diver Evol* 4:165–173
- Sanger TJ, Sherratt E, Mcglathlin JW et al (2013) Convergent evolution of sexual dimorphism in skull shape using distinct developmental strategies. *Evolution* 67:2180–2193
- Schmid JR, Tucker AD (2018) Comparing diets of Kemp's ridley sea turtles (*Lepidochelys kempii*) in mangrove estuaries of southwest Florida. *J Herpetol* 52:252–225
- Seminoff JA, Resendiz A, Nichols JW (2002) Diet of East Pacific green turtles (*Chelonia mydas*) in the central Gulf of California, Mexico. *J Herpetology* 36:447–453
- Seney EE, Musick JA (2005) Diet analysis of Kemp's ridley sea turtles (*Lepidochelys kempii*) in Virginia. *Chelonian Conserv Biology* 4:864–871
- Seney EE, Musick JA (2007) Historical diet analysis of loggerhead sea turtles (*Caretta caretta*) in Virginia. *Copeia* 2007:478–489
- Shaver DJ (1991) Feeding ecology of wild and head-started Kemp's ridley sea turtles in South Texas waters. *J Herpetol* 25:327–334
- Sherratt E, Coutts FJ, Rasmussen AR, Sanders KL (2019) Vertebral evolution and ontogenetic allometry: the developmental basis of extreme body shape divergence in microcephalic sea snakes. *Evol Dev* 21:135–144
- Solounias N, Moelleken SMC (1993) Dietary adaptation of some extinct ruminants determined by pre-maxillary shape. *J Mammal* 74:1059–1971
- Thomson RC, Spinks PQ, Shaffer HB (2021) A global phylogeny of turtles reveals a burst of climate-associated diversification on continental margins. *Proc Natl Acad Sci USA* 118:e2012215118
- Tokita M, Yano W, James HF et al (2017) Cranial shape evolution in adaptive radiations of birds: comparative morphometrics of Darwin's finches and Hawaiian honeycreepers. *Philos T R Soc B* 372:20150481
- Urošević A, Ljubisavljević K, Ivanović A (2013) Patterns of cranial ontogeny in lacertid lizards: morphological and allometric disparity. *J Evol Biol* 26:399–415

- Veilleux CC, Kirk EC (2014) Visual acuity in mammals: effects of eye size and ecology. *Brain Behav Evol* 83:43–53
- Vita G, Zanata AM, Datovo A (2020) Anatomy and ontogenetic changes of the facial and gular musculature of the tetra *Astyanax brucutu*: A remarkable case of adaptation to durophagy. *J Anat* 237:1136–1150
- Wang B, Zelditch M, Badgley C (2021) Geometric morphometrics of mandibles for dietary differentiation of Bovidae. *Artiodactyla, Mammalia. Current Zoology*
- Weems RE, Brown KM (2017) More-complete remains of *Procolpochelys charlestonensis* (Oligocene, South Carolina), an occurrence of *Euclastes* (upper Eocene, South Carolina), and their bearing on Cenozoic pancheloniid sea turtle distribution and phylogeny. *J Paleontol* 91:1228–1243
- Wikelski M, Trillmich F (1994) Foraging strategies of the Galapagos Marine Iguana (*Amblyrhynchus cristatus*): adapting behavioral rules to ontogenetic size change. *Behaviour* 128:255–279
- Wiley DF, Landmark Editor 3.6 (IDAV, University of California, Davis, 2007).[https://www.cs.ucdavis.edu/~amenta/LandmarkDoc\\_v3\\_b6.pdf](https://www.cs.ucdavis.edu/~amenta/LandmarkDoc_v3_b6.pdf)
- Wilson LaB, Sánchez-Villagra MR (2011) Evolution and phylogenetic signal of growth trajectories: the case of chelid turtles. *J Exp Zool Part B: Mol Dev Evol* 316B:50–60
- Zangerl R (1980) Patterns of phylogenetic differentiation in the toxochelyid and cheloniid sea turtles. *Am Zool* 20:585–596
- Zangerl R, Hendrickson LP, Hendrickson JR (1988) A redescription of the Australian flatback sea turtle, *Natator depressus*. Bishop Museum Pr, Honolulu, Hawaii
- Zug GR, Parham JF (1996) Age and growth in leatherback turtles, *Dermodochelys coriacea* (Testudines: Dermochelyidae): a skeletochronological analysis. *Chelonian Conserv Biol* 2:244–249

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