

Investigating chromosome pairing in bread wheat using *ASYNAPSIS I*

by

Scott Andrew Boden

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Table of Contents

Table of Contents.....	II
List of Figures.....	IX
List of Tables.....	XI
Abstract	XII
Declaration	XIV
Acknowledgements.....	XV
Glossary of Abbreviations.....	XVII
Chapter 1 - Literature Review	1
1.1 - Meiosis.....	1
1.1.1 - Stages of meiosis	1
1.1.2 - Prophase I.....	3
1.1.3 - Metaphase I to telophase II	4
1.2 - Molecular analysis of prophase I.....	5
1.2.1 - Recombination – from structure to functional analysis	6
1.2.2 – Alignment and recognition of homologous chromosomes	8
1.2.3 - Synaptonemal Complex – structure and function.....	10
1.3 - Meiosis in bread wheat – a hexaploid acting as a diploid.....	12
1.4 - <i>ASY1</i> – a gene required for successful chromosome synapsis.....	16
1.4.1 - <i>ScHOP1</i> : A gene essential for homologous chromosome pairing in yeast	17
1.4.2 - <i>AtASY1</i> , <i>BoASY1</i> and <i>OsPAIR2</i> : three genes required for synapsis of homologous chromosomes.....	19
1.5 – RNA interference	21
1.6 – Rationale of current study.....	23
Chapter 2 - Genetic and Transcriptional Analyses of <i>TaASY1</i>	25
2.1 – Introduction	25
2.2 – Materials and methods	26

2.2.1 – Plant growth conditions.....	26
2.2.2 – Staging meiotic material.....	27
2.2.3 – Isolation of <i>TaASY1</i> cDNA.....	28
2.2.3.1 – RNA isolation and gel electrophoresis.....	28
2.2.3.2 – cDNA synthesis	28
2.2.3.3 – Isolation of full-length <i>TaASY1</i> open reading frame.....	29
2.2.3.4 – Cloning of PCR products.....	29
2.2.3.4.1 – Purification of DNA from agarose gels.....	29
2.2.3.4.2 – DNA ligation	29
2.2.3.4.3 – Bacteria transformation and colony PCR.....	30
2.2.3.5 – Sequencing of <i>TaASY1</i> clones	31
2.2.4 – <i>TaASY1</i> genome location.....	31
2.2.4.1 – DNA isolation.....	31
2.2.4.2 – Identifying genome location.....	32
2.2.4.2.1 – Southern blot analysis	32
2.2.4.2.2 – Mutant PCR analysis.....	34
2.2.5 – <i>TaASY1</i> genomic DNA and native promoter isolation.....	35
2.2.5.1 – Genomic clone isolation and sequencing	35
2.2.5.2 – Native promoter isolation.....	36
2.2.5.2.1 – Production of genome walking libraries.....	36
2.2.5.2.2 – PCR based genome walking.....	37
2.2.5.2.3 – Cloning and sequencing of genome walking fragments.....	38
2.2.5.2.4 – Confirmation of promoter specificity to <i>TaASY1</i>	38
2.2.6 – Sequence analysis	40
2.2.7 – <i>TaASY1</i> transcriptional analysis.....	41
2.2.7.1 – RNA isolation and gel electrophoresis.....	41
2.2.7.2 – Northern blot analysis	42

2.2.7.3 – Microarray analysis.....	42
2.2.7.4 – Quantitative real time PCR (Q-PCR) analysis.....	42
2.2.8 – Digital image capture	44
2.3 – Results	44
2.3.1 – Isolation and analysis of the <i>TaASY1</i> cDNA clone.....	44
2.3.2 – <i>TaASY1</i> gene structure and genome location.....	47
2.3.3 – Isolation of the native <i>TaASY1</i> promoter sequence.....	49
2.3.4 – Transcript expression analysis of <i>TaASY1</i>	54
2.4 – Discussion.....	57
2.4.1 – The genetics of <i>TaASY1</i>	57
2.4.2 – <i>TaASY1</i> transcripts are highly abundant in meiotic tissue.....	60
Chapter 3 - Investigation of the <i>TaASY1</i> Protein.....	64
3.1 – Introduction	64
3.2 – Materials and methods	65
3.2.1 - Production of <i>TaASY1</i> protein.....	65
3.2.1.1 - Development of protein expression vectors	65
3.2.1.2 - <i>TaASY1</i> protein production in <i>E. coli</i>	66
3.2.1.3 – Purification and gel electrophoresis of recombinant <i>TaASY1</i> protein.....	66
3.2.1.4 - MS/MS analysis of recombinant <i>TaASY1</i> protein	67
3.2.2 - Production of anti- <i>TaASY1</i> antibodies.....	68
3.2.2.1 - Production of mouse anti- <i>TaASY1</i> antibody	68
3.2.2.2 - Production of rabbit polyclonal anti- <i>TaASY1</i> antibody	68
3.2.3 - <i>TaASY1</i> western blot analysis	69
3.2.3.1 - Plant protein extraction	69
3.2.3.2 - Gel electrophoresis and protein transfer to membrane	71
3.2.3.3 – Western blot analysis and protein detection	71
3.2.3.4 - Image capture	72

3.2.4 - Immuno-localisation of <i>TaASY1</i> by transmission electron microscopy (TEM).....	72
3.2.4.1 - Anther fixation and sample preparation	72
3.2.4.2 - Immunisation and counter-staining of sectioned tissue	73
3.2.4.3 - Sample visualisation using TEM and image capture	73
3.2.5 - Immuno-localisation of <i>TaASY1</i> by fluorescence microscopy.....	74
3.2.5.1 - Anther fixation	74
3.2.5.2 – Acrylamide embedding of meiocytes.....	74
3.2.5.3 - Immunisation and counter-staining of poly-acrylamide pads	75
3.2.5.4 - Sample visualisation and image capture	76
3.3 – Results	76
3.3.1 – Production of <i>TaASY1</i> recombinant protein	76
3.3.2 – Western blot analysis	78
3.3.3 – Immuno-gold localisation of <i>TaASY1</i> using TEM.....	78
3.3.4 – Immuno-fluorescence localisation of <i>TaASY1</i>	82
3.4 – Discussion.....	85
3.4.1 – Using localisation studies to understand the role of <i>TaASY1</i>	85
Chapter 4 - Investigating Meiosis in <i>Taasy1</i> and <i>ph1b</i> Mutants.....	91
4.1 – Introduction	91
4.2 – Materials and methods	93
4.2.1 – Plant material and growth conditions.....	93
4.2.2 – Generating <i>Taasy1</i> mutant plants.....	95
4.2.2.1 – Construction of the RNA interference (RNAi) vector	95
4.2.2.2 – Transformation of Bread Wheat (Bob White MPB26)	96
4.2.3 – Genotype analysis	96
4.2.3.1 – Identification of positive transgenic plants <i>via</i> PCR	96
4.2.3.1.1 – Identification of positive T ₁ transgenic plants	96
4.2.3.1.2 – Identification of positive T ₂ transgenic plants	97

4.2.3.2 – Southern blot hybridisation	97
4.2.3.2.1 – DNA extraction.....	98
4.2.3.2.2 – Genomic DNA digestion and transfer to membrane.....	98
4.2.3.2.3 – Southern blot analysis	98
4.2.4 – RNA and protein analysis.....	99
4.2.4.1 – Collection of meiotic tissue from <i>Taasy1</i> mutants.....	99
4.2.4.2 – Collection of meiotic tissue from <i>ph1b</i>	99
4.2.4.3 – RNA and protein extraction.....	100
4.2.5 – <i>TaASY1</i> expression analysis in <i>Taasy1</i> and <i>ph1b</i>	100
4.2.5.1 – Q-PCR analysis of <i>Taasy1</i> mutants.....	100
4.2.5.2 – Q-PCR analysis of <i>ph1b</i>	101
4.2.5.3 – Western blot analysis of <i>ph1b</i> mutants.....	101
4.2.6 – Fertility analysis of <i>Taasy1</i> mutants.....	102
4.2.6.1 – Analysis of whole plant morphology	102
4.2.6.2 – Analysis of pollen viability.....	102
4.2.6.3 – Analysis of seed set in <i>Taasy1</i> mutants	103
4.2.6.3.1 – Seed collection from T ₁ and T ₂ generation plants.....	103
4.2.6.4 – Data analysis.....	103
4.2.7 – Analysis of chromosome morphology during meiosis	103
4.2.7.1 – Collection of staged anthers	104
4.2.7.2 – Analysis of chromosome morphology using Feulgen's stain.....	104
4.2.7.3 – Image capture.....	105
4.2.8 – Immuno-localisation of <i>TaASY1</i> in <i>Taasy1</i> and <i>ph1b</i> mutants.....	105
4.2.8.1 – Collection of staged anthers	105
4.2.8.2 – Sample preparation and immunisation.....	105
4.2.8.3 – Sample visualisation and image capture.....	106
4.3 – Results	106

4.3.1 – <i>Taasy1</i> mutant analysis – T ₁ generation	106
4.3.1.1 – Genotype analysis	106
4.3.1.2 – Q-PCR analysis of T ₁ generation <i>Taasy1</i> mutants	108
4.3.1.3 – T ₁ generation <i>Taasy1</i> plants display reduced fertility	109
4.3.2 – <i>Taasy1</i> mutant analysis – T ₂ generation	110
4.3.2.1 – Genotype analysis	110
4.3.2.2 – Q-PCR analysis of T ₂ generation <i>Taasy1</i> mutants	112
4.3.2.3 – <i>Taasy1</i> mutants display significant reduction in fertility	113
4.3.2.4 – Meiotic chromosome morphology is disrupted in the <i>Taasy1</i> mutants	116
4.3.2.5 – <i>TaASY1</i> localisation in the <i>Taasy1</i> mutants.....	119
4.3.3 – Investigation of <i>TaASY1</i> in <i>ph1b</i>	122
4.3.3.1 – Q-PCR analysis of meiotic candidate genes in <i>ph1b</i>	122
4.3.3.2 – Western blot analysis of <i>TaASY1</i> in <i>ph1b</i>	125
4.3.3.3 – Localisation of <i>TaASY1</i> in <i>ph1b</i>	125
4.4 – Discussion.....	128
4.4.1 – Genetic analysis of the <i>Taasy1</i> mutants	129
4.4.2 – Expression analysis of <i>Taaasy1</i> mutants.....	131
4.4.3 – Whole plant morphology of <i>Taasy1</i> mutants.....	132
4.4.4 – Investigating meiosis of the <i>Taasy1</i> mutants.....	133
4.4.5 – Investigation of <i>TaASY1</i> in the <i>ph1b</i> mutant	137
Chapter 5 - General Discussion and Future Directions	141
5.1 – General discussion	141
5.1.1 – Investigating <i>TaASY1</i> in wild-type wheat	141
5.1.2 – Using <i>TaASY1</i> RNA interference mutants to define the role of <i>TaASY1</i>	146
5.1.3 – Using <i>TaASY1</i> to solve the mystery of <i>Ph1</i>	148
5.1.4 – A model for the involvement of chromosome pairing in bread wheat.....	152
5.2 – Future directions	154

5.2.1 – Further investigations of <i>TaASY1</i> in bread wheat.....	154
5.2.2 – Analysis of <i>Taasy1</i> mutants.....	155
5.2.3 – Further analysis of <i>Ph1</i>	157
5.2.4 – The bigger picture	158
References	160
Appendix A	183
Appendix B	187
Appendix C	192
Appendix D.....	200

List of Figures

Figure 1.1 – Meiotic divisions I and II as observed in <i>Secale cereale</i> microsporocytes.....	3
Figure 2.1 - Alignment and comparison of the deduced amino acid sequences within the HORMA domains of <i>TaASY1</i> , <i>OsPAIR2</i> (<i>O. sativa</i>), <i>AtASY1</i> (<i>A. thaliana</i>), <i>BoASY1</i> (<i>B. oleracea</i>) and <i>ScHOP1</i> (<i>S. cerevisiae</i>)	46
Figure 2.2 - Schematic representation of <i>TaASY1</i> gene structure with exons and introns..	47
Figure 2.3 - Chromosomal location of <i>TaASY1</i> determined by Southern blot	48
Figure 2.4 - Chromosomal location of <i>TaASY1</i> determined by PCR Analysis	49
Figure 2.5 – Isolation of the <i>TaASY1</i> promoter by genome walking.	51
Figure 2.6 – Diagrammatic representation of products amplified via genome walking..	52
Figure 2.7 – Tissue series and meiosis stage specific northern blot analysis of <i>TaASY1</i>	54
Figure 2.8 – Meiosis staged microarray analysis of <i>TaASY1</i> ..	56
Figure 2.9 - Meiosis staged and vegetative tissue Q-PCR analysis of <i>TaASY1</i>	56
Figure 3.1 – Production of recombinant <i>TaASY1</i> in <i>E coli</i> ..	77
Figure 3.2 – Western blot analysis of <i>TaASY1</i>	78
Figure 3.3 – Immuno-gold localisation of <i>TaASY1</i> via electron microscopy.....	79
Figure 3.4 – Schematic diagrams of <i>TaASY1</i> localisation as detected by electron microscopy.....	81
Figure 3.5 – Immuno-fluorescence localisation using anti- <i>TaASY1</i> immune sera detected <i>TaASY1</i> only in meiocytes during prophase I.	83
Figure 3.6 – Immuno-fluorescence localisation of <i>TaASY1</i> during pre-meiotic interphase and throughout prophase I.	83
Figure 4.1 – Schematic diagram of the construct used to generate the <i>Taasy1</i> RNAi lines.....	95
Figure 4.2 – Genotype analysis of T ₁ generation <i>Taasy1</i> mutants.....	108
Figure 4.3 – Q-PCR analysis of T ₁ generation <i>Taasy1</i> mutants.	109
Figure 4.4 – Genotype analysis of T ₂ generation <i>Taasy1</i> mutants via PCR.	111
Figure 4.5 – Q-PCR analysis of T ₂ generation <i>Taasy1</i> mutants.	112

Figure 4.6 - Phenotype of <i>Taasy1</i> mutants (T ₂ generation) compared to wild-type wheat during vegetative and reproductive development.....	115
Figure 4.7 – Alexander stain shows increased pollen abortion in <i>Taasy1</i> mutants (T ₂ generation). ...	116
Figure 4.8 – Feulgen's-stained chromosome preparation of <i>Taasy1</i> mutants (T ₂ generation) and <i>ph1b</i>	118
Figure 4.9 – Feulgen's-stained chromosome spreads of <i>Taasy1</i> mutants displaying various anaphase I phenotypes.....	119
Figure 4.10 – Feulgen's-stained metaphase I chromosome spreads of independent transgenic lines (T ₂ generation).....	119
Figure 4.11 – Immuno-fluorescence localisation of <i>TaASY1</i> during prophase I of <i>Taasy1</i> mutants..	122
Figure 4.12 – Merged images of <i>Taasy1</i> cells lacking any detectable ASY1 protein.....	122
Figure 4.13 – Q-PCR analysis of <i>TaASY1</i> and meiotic candidate genes during meiosis in <i>ph1b</i>	123
Figure 4.14 – Western blot analysis of <i>TaASY1</i> protein during meiosis in <i>ph1b</i> and wild-type wheat (cv. Chinese Spring).....	125
Figure 4.15 – Immuno-fluorescence localisation of <i>TaASY1</i> during prophase I of <i>ph1b</i>	128
Figure 5.1 - A proposed model for the control of homologous chromosome pairing in bread wheat developed using the results from this study.....	153

List of Tables

Table 2.1 – The list of primers used to isolate the <i>TaASY1</i> genomic DNA sequence.....	36
Table 2.2 – List of primers used during genome walking experiments to isolate the <i>TaASY1</i> promoter.	
.....	38
Table 2.3 – List of primers used to confirm that the obtained promoter sequence was specific to <i>TaASY1</i>	39
Table 2.4 - Percentage identity between the full-length <i>TaASY1</i> protein sequence and characterised asynaptic proteins from other species.....	45
Table 2.5 - Percentage identity between the HORMA domain sequence of <i>TaASY1</i> and the HORMA domains of characterised asynaptic proteins from other species.....	46
Table 2.6 – A summary of the three main groups of <i>cis</i> -elements identified within the 2355 bp <i>TaASY1</i> promoter sequence.	53
Table 4.1 – A summary of selected T ₁ <i>Taasy1</i> mutant plant lines.....	94
Table 4.2 – A summary of selected T ₂ <i>Taasy1</i> mutant plant lines.....	94
Table 4.3 – A summary of genotype analysis performed on <i>Taasy1</i> mutants from the T ₁ generation.	107
Table 4.4 – A summary of genotype analysis performed on <i>Taasy1</i> mutants from the T ₂ generation.	111
Table 4.5 - Statistics of the number of seeds per floret, seeds per head and pollen abortion in RNAi <i>Taasy1</i> transgenic wheat plants (T ₂) compared to wild-type wheat plants (Bob White MPB26).	115

Abstract

Pairing and synapsis of homologous chromosomes are required for normal chromosome segregation and the exchange of genetic material during meiosis. Pairing is defined as the recognition and alignment of chromosomes that occurs either pre-meiotically or during early prophase I to ensure that associations *via* synapsis and recombination occur only between homologues. Synapsis is the intimate juxtaposition of homologous chromosomes that is complete at pachytene following formation of a tri-partite proteinaceous structure known as the synaptonemal complex (SC). In yeast, HOP1 is an essential component of the SC that localises along chromosome axes during prophase I and promotes homologous chromosome interactions. Homologues in *Arabidopsis* (*AtASY1*), *Brassica* (*BoASY1*) and rice (*OsPAIR2*) have been isolated through analysis of mutants that display decreased fertility due to severely reduced synapsis of homologous chromosomes. Analysis of these genes has indicated that they play a similar role to HOP1 in pairing and formation of the SC through localisation to axial/lateral elements of the SC. In this study, we have characterised the bread wheat homologue of *HOP1*, *TaASY1*, and its encoded protein.

The full length cDNA and genomic DNA clones of *TaASY1* have been isolated, sequenced and characterised. *TaASY1* is located on chromosome group 5 and the open reading frame displays significant similarity to *OsPAIR2* (84%) and *AtASY1* (63%). In addition to *OsPAIR2* and *AtASY1*, the deduced amino acid sequence also displays sequence similarity to *ScHOP1*, with all four proteins containing a HORMA domain. Transcript and protein analysis showed that expression is largely restricted to meiotic tissue, with elevated levels during the stages of prophase I when pairing and synapsis of homologous chromosomes occurs.

Antibodies specific to *TaASY1* were used in immuno-fluorescence microscopy and immuno-gold transmission electron microscopy to investigate the localisation of *TaASY1* in

meiotic cells. Immuno-fluorescence analysis initially detected ASY1 in pollen mother cells (PMCs) during meiotic interphase as foci randomly distributed over the chromatin. The ASY1 signal became increasingly continuous during leptotene, reflecting the changes occurring in chromosome morphology. Throughout zygotene, the signal became progressively more continuous, localising along the entire length of the axial elements as chromosomes synapsed. This signal appeared to persist until pachytene, before disappearing from the chromatin as the SC disassociated through late pachytene and early diplotene. The immuno-gold based electron microscopy displayed that *TaASY1* localises to chromatin that is associated with both axial elements before SC formation as well as chromatin of lateral elements within formed SCs.

Analysis of RNAi *Taasy1* mutants was performed to further define the role of ASY1 in bread wheat meiosis. ASY1 localisation was disrupted in these mutants, with a diffuse and non-continuous signal observed through leptotene and zygotene. Feulgen staining of meiotic chromosomes displayed reduced synapsis during prophase I, as well as multivalents at metaphase I and abnormal chromosome segregation during anaphase I. These observations are consistent with the presence of homoeologous chromosome interactions. *TaASY1* expression and localisation was also investigated in the bread wheat pairing mutant, *ph1b*. Quantitative real-time PCR (Q-PCR) revealed that *TaASY1* is significantly up-regulated in *ph1b*, with greater than 20-fold expression compared to wild-type Chinese Spring, while maintaining the same pattern of expression as wild-type through progressive stages of meiosis. ASY1 localisation was significantly disrupted in *ph1b*, with irregular loading on axial elements during mid to late zygotene, indicative of abnormal chromatin remodelling and multiple axial element associations that have previously been reported in *ph1b*. Taken together, these results indicate that *TaASY1* is essential for promoting homologous chromosome interactions during meiosis, and that impairment of ASY1 function in bread wheat meiosis results in reduced restriction of chromosome associations to homologues.

Declaration

I declare that the work presented in this thesis contains no material which has been accepted for the award of any other degree or diploma in any University or other tertiary institution. To the best of my knowledge and belief, this thesis does not contain any material previously written or published by another person, except where due reference is made in the text.

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Glossary of Abbreviations

Abbreviation	Full term
3'	three prime
5'	five prime
9mer	9 base pair nucleotide
α -dCTP	alpha-deoxycytidine triphosphate
°C	degrees Celcius
<i>Amp</i>	<i>Ampicillin</i>
<i>ASYI</i>	<i>ASYnapsis 1</i>
<i>At</i>	<i>Arabidopsis thaliana</i>
<i>bar</i>	bialaphos resistance gene
BCIP/NBT	5-bromo-4-chloro-3-indolyl phosphate/nitro blue tetrazolium
BLAST	Basic Local Alignment and Search Tool
<i>Bo</i>	<i>Brassica oleracea</i>
bp	base pair
BSA	Bovine Serum Albumin
BW26	Bob White 26 cultivar of bread wheat
<i>Cdk</i>	<i>Cyclin Dependent Kinase</i>
cDNA	complimentary deoxyribonucleic acid
<i>Ce</i>	<i>Caenorhabditis elegans</i>
CT	cycle threshold
cv.	cultivar
DABCO	diazabicyclo-[2,2,2] octane

DAPI	4',6-diamidino-2-phenylindole
<i>DMC1</i>	<i>Disrupted Meiotic cDNA 1</i>
DNA	deoxyribonucleic acid
DPSS	diode-pumped solid state (laser)
dNTP	deoxynucleoside triphosphate
DTT	dithiothreitol
EDTA	ethylene diamine tetra-acetic acid
<i>EFA</i>	<i>Elongation Factor 1 Alpha</i>
EGTA	ethylene glycol tetra-acetic acid
EST	expressed sequence tag
FISH	fluorescent <i>in situ</i> hybridisation
g	gram
g	relative centrifugal force
<i>GAPdH</i>	<i>GlycerAldehyde-3-Phosphate DeHydrogenase</i>
h	hour(s)
<i>HIM-3</i>	<i>High Incidence of Males 3</i>
<i>His</i>	histidine
HOP1	<i>HOmologous Pairing 1</i>
HORMA	Hop1p, Rev7p, MAd2
IgG	immunoglobulin G
IPTG	isopropyl-1-thio-β-D-galactoside
kb	kilobase
kDa	kilo Dalton
KLH	keyhole limpet hemocyanin
LASER	Light Amplification by Stimulated Emission of Radiation

LB	Luria Bertani
M	molar
mg	milli gram
mM	milli molar
mCi mL ⁻¹	milli Curie per milli litre
<i>MECI</i>	<i>Mitosis Entry Checkpoint 1</i>
min	minute(s)
<i>MLH1/3</i>	<i>Mut L Homologue 1/3</i>
mRNA	messenger ribonucleic acid
MPB CRC	Molecular Plant Breeding Co-operative Research Centre
<i>MRE11</i>	<i>Meiotic REcombination 11</i>
<i>MSH4/5</i>	<i>Mut S Homologue 4/5</i>
MS/MS	tandem mass spectrometry
N/A	not applicable
NCBI	National Center of Biotechnology Information
n.d.	not determined
ng	nano gram
nm	nano metre
NT	nullisomic-tetrasomic
ORF	open reading frame
<i>Os</i>	<i>Oryza sativa</i>
<i>P</i>	probability
<i>PAIR2</i>	<i>homologous Pairing Aberration In Rice meiosis 2</i>
PBS	phosphate buffered saline
PCR	polymerase chain reaction

PLACE	PLAnt <i>Cis</i> -acting Regulatory DNA Elements
PMC	pollen mother cell
PVDF	polyvinylidene difluoride
PVP	polyvinyl pyrrolidone
PVPP	polyvinyl polypyrrolidone
Q-PCR	quantitative real-time PCR
Q-TOF ²	quadruple time of flight squared
R40	40 µg µL ⁻¹ RNase in 1X TE
<i>RAD51</i>	<u><i>RA</i>diation sensitive 51</u>
<i>REC8</i>	<u><i>RE</i>Combination 8</u>
<i>RED1</i>	<u><i>RE</i>ductional division 1</u>
RNA	ribonucleic acid
RNase	ribonuclease
RNAi	RNA interference
rpm	revolutions per minute
<i>Sc</i>	<i>Saccharomyces cerevisiae</i>
SC	synaptonemal complex
sec	second(s)
SDS	sodium dodecyl sulphate
SDS-PAGE	SDS - polyacrylamide gel electrophoresis
SNP	single nucleotide polymorphism
SSC	standard saline citrate
<i>SPO11</i>	<u><i>SP</i>Oulation-deficient 11</u>
<i>Ta</i>	<i>Triticum aestivum</i>
<i>Taq</i>	<i>Thermus aquaticus</i>

TBS	tris-buffered saline
T-DNA	transfer DNA
TE	Tris EDTA solution
<i>TEL1</i>	<i>TELomere 1</i>
TEM	transmission electron microscopy
T _m	melting temperature
Tris	tris(hydroxymethyl)aminomethane
U	units
µL	micro litre
µg	micro gram
µM	micro molar
UV	ultra-violet
V	volts
v/v	volume/volume
w/v	weight/volume
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside