# CELL LINEAGE, CELL MATURITY AND BCR-ABL: FACTORS WHICH INFLUENCE IMATINIB UPTAKE IN CHRONIC MYELOID LEUKAEMIA

# **Jane Engler**

The Melissa White Laboratory
Department of Haematology
Centre for Cancer Biology
SA Pathology (IMVS)
Adelaide, Australia

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Faculty of Health Sciences Department of Medicine The University of Adelaide Adelaide, Australia

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# candor dat viribus alas

(Sincerity gives wings to strength)

# ipsa scientia poleslas est (Knowledge itself is power)

# prefer et obdura; dolor hic tibi proderit olim (Be patient and tough; some day this pain will be useful to you)

# aul viam inveniam aul faciam

(I'll either find a way or make one)

per aspera ad astra (Through adversities to the stars!)

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Jane Engler

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# **PUBLICATIONS**

#### **Manuscripts**

<u>JR Engler</u>, TP Hughes & DL White. OCT-1 as a determinant of response to anti-leukemic treatment. *Clinical Pharmacology and Therapeutics*. 2011. Feb 23; Epub ahead of print. (Impact Factor: 7.586)

<u>JR Engler</u>, AC Zannettino, C Bailey, J Rasko, TP Hughes & DL White. OCT-1 function varies with cell lineage but is not influenced by BCR-ABL. *Haematologica*. 2011. Feb;96(2):213-20. (Impact Factor: 6.416)

JR Engler, A Frede, V Saunders, AC Zannettino, DL White & TP Hughes. The poor response to imatinib observed in CML patients with low OCT-1 activity is not attributable to lower uptake of imatinib into their CD34+ cells. *Blood.* 2010. Oct;116(15):2776-8. (Impact Factor: 10.555)

<u>JR Engler</u>, A Frede, V Saunders, AC Zannettino, TP Hughes & DL White. Chronic myeloid leukemia CD34+ cells have reduced uptake of imatinib due to low OCT-1 activity. *Leukemia*. 2010. Apr;24(4):765-70. (Impact Factor: 8.296)

#### **Conference Abstracts**

<u>JR Engler</u>, C Bailey, J Rasko, AC Zannettino, DL White & TP Hughes. OCT-1 function varies with cell lineage but is not influenced by BCR-ABL. *New Directions in Leukaemia Research*, March 2010. Sunshine Coast, Australia. Poster Presentation.

JR Engler, A Frede, V Saunders, AC Zannettino, DL White & TP Hughes. OCT-1 activity in CML CD34+ cells is not predictive of molecular response to imatinib treatment in CP-CML patients, despite the strong predictive value of MNC OCT-1 activity. *American Society of Hematology Annual Meeting*, December 2009. New Orleans, USA. Poster Presentation.

JR Engler, A Frede, V Saunders, AC Zannettino, R D'Andrea, TP Hughes & DL White. CML CD34+ cells have reduced uptake of imatinib due to uniformly low OCT-1 activity, which can be increased with diclofenac treatment. *Centre for Stem Cell Research Annual Meeting*, November 2009. Adelaide, Australia. Poster Presentation.

JR Engler, A Frede, V Saunders, AC Zannettino, DL White & TP Hughes. OCT-1 activity in CML CD34+ cells is not predictive of molecular response to imatinib treatment in CP-CML patients. *Haematology Society of Australia and New Zealand Annual Meeting*, October 2009. Adelaide, Australia. Oral Presentation.

JR Engler, A Frede, AC Zannettino, DL White & TP Hughes. Reduced activity of the OCT-1 protein in primitive CML cells: A likely determinant of stem cell resistance in imatinib treated CML patients. *American Society of Haematology Annual Meeting*, December 2008. San Francisco, USA. Oral Presentation.

JR Engler, A Frede, S Quinn, AC Zannettino, TP Hughes & DL White. Reduced activity of the OCT-1 protein in primitive CML cells may be a key to stem cell resistance in imatinib treated CML patients. *New Directions in Leukaemia Research*, March 2008. Sunshine Coast, Australia. Poster Presentation.

# **SCHOLARSHIPS & AWARDS**

#### Poster Prize, Centre for Stem Cell Research Annual Meeting. 2009.

For the abstract entitled "CML CD34+ cells have reduced uptake of imatinib due to uniformly low OCT-1 activity, which can be increased with diclofenac treatment", Adelaide, November 2009.

#### Student Travel Scholarship, The Leukaemia Foundation of Australia. 2008.

Support for students to attend the New Directions in Leukaemia Research Conference. Awarded on the basis of submitted abstracts for work of exceptional novelty and significance. For the abstract entitled "Reduced activity of the OCT-1 protein in primitive CML cells may be a key to stem cell resistance in imatinib treated CML patients", Sunshine Coast, March 2008.

#### PhD Scholarship, The Leukaemia Foundation of Australia. 2007-2010.

To provide support for the educational and professional development of researchers and other professionals undertaking a PhD. The award is to support basic, applied and translational research in Australia into the causes, treatment and care of people with leukaemia, lymphoma, myeloma and related blood disorders.

# **ABBREVIATIONS**

ABL Abelson kinase

ACD Anticoagulent Citrate Dextrose Solution Formula A

ALL Acute lymphoblastic leukaemia

**AML** Acute myeloid leukaemia

AP Accelerated phase

ATP Adenosine triphosphate

BC Blast crisis

BCR Breakpoint cluster region

**BM** Bone marrow

**BSA** Bovine serum albumin

**C** Celcius

**CCR** Complete cytogenetic response

**cDNA** Complementary deoxyribonucleic acid

**CFSE** 5-6-carboxyfluorescein diacetate, succinimidyl ester

**CML** Chronic myeloid leukaemia

**CMR** Complete molecular response

**CP** Chronic phase

**CPM** Counts per minute

**Crkl** Crk-like protein

**p-Crkl** Phosphorylated Crk-like protein

CV Control vector

**DEPC** Diethyl pyrocarbonate

**DMSO** Dimethyl sulphoxide

**DNA** Deoxyribonucleic acid

**dNTPs** Deoxynucleotide triphosphates

**DTT** Dithiothreitol

**EDTA** Ethylene diamine tetraacetate

**eGFP** Enhanced green fluorescence protein

FACS Fluorescence activated cell sorting

FITC Fluorescein isothiocyanate

FCS Foetal calf serum

Hanks Hanks Balanced Salt Solution

**HSC** Haematopoietic stem cell

**IC50** 50% inhibitory concentration

**IFN-**α Interferon alpha

IM Imatinib (STI571)

IRIS International randomised study of interferon versus STI571

IUR Intracellular uptake and retention

kD Kilo Dalton

**L** Litre

M Molar

mA Milli Amp (10<sup>-3</sup> Amp)

MACS Magnetically activated cell sorting

MCR Major cytogenetic response

MMR Major molecular response

**mM** Milli Molar (10<sup>-3</sup> Molar)

MNC Mononuclear cells

mRNA Messenger ribonucleic acid

μ**M** Micro Molar (10<sup>-6</sup> Molar)

**μg** Micro gram (10<sup>-6</sup> gram)

**ng** Nano gram (10<sup>-9</sup> gram)

Nil Nilotinib (AMN107)

OA OCT-1 activity

OCT-1 Organic cation transporter 1

PB Peripheral blood

PBS Phosphate Buffered Saline

PCR Polymerase chain reaction

**PE** Phycoerythrin

Ph Philadelphia chromosome

PI Propidium Iodide

PMA Phorbol-12-myristate-13-acetate

**PVDF** Polyvinylidene fluoride

RNA Ribonucleic acid

**RPM** Revolutions per minute

**RPMI** Roswell Park Memorial Institute (media)

RT Room temperature

**RT-PCR** Reverse transcription polymerase chain reaction

**RQ-PCR** Real time quantitative polymerase chain reaction

**SD** Standard deviation

SDS Sodium dodecyl sulphate

**SEM** Standard error of the mean

**S/N** Supernatant

**STI571** Signal transduction inhibitor 571 (imatinib)

**TBS** Tris buffered saline

**TBST** Tris buffered saline with 0.1% Tween20

**TKI** Tyrosine kinase inhibitor

**U** Units

**UV** Ultraviolet

v/v Volume per volume

WBC White blood cells

WCC White cell count

w/v Weight per unit volume

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## **ABSTRACT**

Despite the excellent responses observed in patients with chronic phase (CP) chronic myeloid leukaemia (CML) on imatinib therapy, approximately 25% display primary resistance or suboptimal response. The organic cation transporter 1 (OCT-1) is the major active influx pump for imatinib in CML cells. The functional OCT-1 activity in mononuclear cells (MNC) is highly variable between patients and significantly correlates with a patient's molecular response to imatinib treatment and overall survival. Given the strong predictive value of OCT-1 activity, the present study was aimed at identifying factors responsible for the variation in OCT-1 activity seen in patients.

Pure populations of neutrophils, monocytes and lymphocytes were isolated from the peripheral blood of CML patients at diagnosis. The OCT-1 activity and OCT-1 mRNA expression was found to be the highest in the neutrophil population, followed by monocytes then lymphocytes. When the surface expression of the granulocytic antigens CD15 and CD16 were examined, a significant correlation was observed between MNC OCT-1 activity and the proportion of immature myeloid cells expressing CD15+16-. Interestingly, the neutrophil OCT-1 activity was found to be similar when recovered from CML patients at diagnosis, CML patients in cytogenetic remission and normal donors, implying that BCR-ABL expression is unlikely to influence OCT-1 activity. This hypothesis was confirmed in a cell line model, in which ectopic BCR-ABL expression was not found to directly affect OCT-1 expression or function, but stimulated myeloid differentiation which, in turn, led to increased OCT-1 activity. These data suggest that the predictive MNC OCT-1 activity is most strongly related to cell lineage, particularly the proportion of immature myeloid cells, but is not directly related to BCR-ABL.

CML early progenitor cells are less sensitive to imatinib induced apoptosis and are likely contributors to disease persistence. It was found that the OCT-1 activity and OCT-1 mRNA expression was significantly lower in primitive CD34+ cells compared with mature CD34- cells recovered from CML patients. These results indicate that low imatinib accumulation in primitive CML cells may be a critical determinant of long-term disease persistence. Studies to investigate whether the MNC OCT-1 activity provides a surrogate indicator of effective targeting of the more immature CD34+ cells failed to identify a relationship between high CD34+ OCT-1 activity and the achievement of major molecular response. This is despite the confirmation of previous findings that high MNC OCT-1 activity is significantly associated with the achievement of major molecular response to imatinib treatment. These important findings suggest that kinase inhibition in these mature cells, and not the CD34+ cells, may be the key determinant of response in CML.

In conclusion, the studies outlined in this thesis have identified cell lineage as a key contributor to MNC OCT-1 activity and hence response to imatinib treatment. While primitive CD34+ cells demonstrate low OCT-1 activity, which may contribute to their persistence despite imatinib therapy, the OCT-1 activity in these cells does not correlate with patient response to treatment. Therefore, direct targeting of this primitive population may not be essential for achievement of early and deep molecular responses.