



THE UNIVERSITY
of ADELAIDE

**Investigating Drugs that Enhance Imatinib
Uptake and Factors which Contribute to the
Functional Activity of OCT-1 CML Cells**

Jueqiong WANG

The Melissa White Laboratory
Department of Haematology
Centre for Cancer Biology
SA Pathology (IMVS)
Adelaide, Australia

&

Faculty of Health Sciences
Department of Medicine
The University of Adelaide
Adelaide, Australia

A thesis submitted to the University of Adelaide
in candidature for the degree of Doctor of Philosophy
November 2012

TABLE OF CONTENTS

TABLE OF CONTENTS	I
LIST OF FIGURES AND TABLES	VI
ABBREVIATIONS	X
PUBLICATIONS	XIII
<i>Manuscripts</i>	<i>xiii</i>
<i>Conference Abstracts</i>	<i>xiii</i>
SCHOLARSHIPS & AWARDS	XV
ACKNOWLEDGEMENTS	XVI
ABSTRACT	XVIII
DECLARATION	XX
CHAPTER I INTRODUCTION	1
1.1. CHRONIC MYELOID LEUKAEMIA	2
1.1.1. <i>Clinical features of CML</i>	<i>2</i>
1.1.2. <i>Pathophysiology of CML</i>	<i>3</i>
1.2. CML THERAPIES IN PRE-TKI ERA	5
1.3. CML THERAPIES IN THE ERA OF TYROSINE KINASE INHIBITORS	8
1.3.1. <i>First generation TKI: imatinib</i>	<i>8</i>
1.3.2. <i>Resistance to imatinib</i>	<i>12</i>
1.3.3. <i>Second generation TKIs: dasatinib and nilotinib</i>	<i>15</i>
1.3.4. <i>Third generation TKI: ponatinib</i>	<i>18</i>
1.3.5. <i>Transplantation in the era of TKIs</i>	<i>19</i>
1.4. DRUG TRANSPORTERS FOR IMATINIB	20
1.4.1. <i>Efflux transporters: ABCB1 and ABCG2</i>	<i>20</i>
1.4.2. <i>Influx transporters for imatinib: OCT-1</i>	<i>20</i>
1.5. DRUG INTERACTIONS WITH IMATINIB VIA TRANSPORTERS	24
1.6. HYPOTHESIS AND AIMS	26
CHAPTER II MATERIALS AND METHODS	28

2.1.	COMMONLY USED REAGENTS	29
2.2.	SOLUTIONS, BUFFERS AND MEDIA	31
2.2.1.	<i>Cell culture media</i>	31
2.2.2.	<i>Complete (25x stock)</i>	31
2.2.3.	<i>Flow cytometry fixative (FACS Fix)</i>	31
2.2.4.	<i>Hanks balanced salt solution (HBSS)</i>	31
2.2.5.	<i>Freeze Mix</i>	31
2.2.6.	<i>Laemmli's buffer</i>	32
2.2.7.	<i>1 x Red Cell Lysis buffer:</i>	32
2.2.8.	<i>MACS buffer</i>	32
2.2.9.	<i>TBS</i>	33
2.2.10.	<i>TBST</i>	33
2.2.11.	<i>Membrane blocking solution (2.5%)</i>	33
2.2.12.	<i>Pefabloc stock solutions</i>	33
2.2.13.	<i>PhosSTOP (10x stock)</i>	33
2.2.14.	<i>Prazosin hydrochloride</i>	33
2.2.15.	<i>RIPA Buffer</i>	34
2.2.16.	<i>SDS-Polyacrylamide Gel</i>	34
2.2.17.	<i>2xSDS load buffer</i>	34
2.2.18.	<i>Thaw solution</i>	35
2.2.19.	<i>Tyrosine kinase inhibitors</i>	35
2.2.20.	<i>White cell fluid</i>	36
2.3.	CELL LINES	36
2.3.1.	<i>K562</i>	36
2.3.2.	<i>KU812</i>	36
2.3.3.	<i>HL60</i>	36
2.3.4.	<i>HeLa</i>	36
2.4.	PRIMARY CELLS FROM CML PATIENTS OR HEALTHY DONORS	37
2.5.	GENERAL TECHNIQUES	37
2.5.1.	<i>Bradford protein assay</i>	37
2.5.2.	<i>Cell counts and cell viability determination</i>	38
2.5.3.	<i>Cryopreservation of cells</i>	38
2.5.4.	<i>Tissue culture</i>	38
2.5.5.	<i>Thawing cells</i>	39
2.5.6.	<i>Lymphoprep isolation of mononuclear cells (MNC)</i>	39

2.6.	SPECIALISED TECHNIQUES	40
2.6.1.	<i>Imatinib intracellular uptake and retention (IUR) assay</i>	40
2.6.2.	<i>Western blot for phosphorylated Crkl (p-Crkl) and IC50</i>	41
2.6.3.	<i>Real time quantitative PCR (RQ-PCR)</i>	42
2.6.4.	<i>Gene expression profiling analysis</i>	45
2.6.5.	<i>Enzyme immunoassays for prostaglandin E2 (PGE2)</i>	46
2.6.6.	<i>Enzyme immunoassays for 15-deoxy-Δ12,14-PGJ2 (15d-PGJ2)</i>	47
2.6.7.	<i>Western blotting for protein of interests</i>	48
2.6.8.	<i>Preparation of nuclear extract and cytoplasmic fraction</i>	49
2.6.9.	<i>Peroxisome proliferator agonist receptor γ transcription factor assay</i>	50
2.6.10.	<i>CD14⁺ and CD15⁺ cells isolation</i>	51
2.7.	STATISTICS	54

CHAPTER III IDENTIFICATION AND VALIDATION OF OCT-1 ACTIVITY

ENHANCERS BY BIOINFORMATICS AND EXPERIMENTAL APPROACHES.....55

3.1.	INTRODUCTION	56
3.2.	METHODS	59
3.2.1.	<i>Selection of candidate drugs as functional OA enhancers</i>	59
3.2.2.	<i>Drugs preparation</i>	60
3.2.3.	<i>Validation of functional OA enhancer candidates</i>	60
3.3.	RESULTS	63
3.3.1.	<i>The Connectivity Map (CMAP) as a tool to select candidate drugs as functional OA enhancers</i>	63
3.3.2.	<i>Experimental validation of OCT-1 enhancer candidates with IUR assay</i>	71
3.3.3.	<i>Effects of CMAP candidates on IUR and OA in K562 cell lines</i>	71
3.3.4.	<i>Effects of NSAIDs on IUR of imatinib and OA in K562 cell lines</i>	73
3.3.5.	<i>Effects of clinically applicable drugs on IUR of imatinib and OA in K562 cell lines</i>	75
3.3.6.	<i>Effects of clinically applicable drugs on IUR and OA in KU812 cell lines</i>	75
3.3.7.	<i>Effects of temperature on the regulation of IUR by diclofenac and ibuprofen in KU812 cells</i>	78
3.3.8.	<i>Effects of diclofenac and ibuprofen on IC50^{imatinib} of BCR-ABL-positive cell lines</i>	80
3.3.9.	<i>Effects of diclofenac and ibuprofen on viable cell counts when co-administrated with imatinib in BCR-ABL-positive cells</i>	82

3.3.10. <i>Effects of diclofenac and ibuprofen on the OA of primary cells</i>	84
3.4. DISCUSSION	87
CHAPTER IV DICLOFENAC REGULATES OCT-1 ACTIVITY AT THE TRANSCRIPTIONAL LEVEL.....	93
4.1. INTRODUCTION	94
4.2. METHODS	96
4.2.1. <i>IUR and OA assessment in the presence and absence of Actinomycin D</i>	96
4.2.2. <i>Gene expression profiling analysis</i>	97
4.3. RESULTS	97
4.3.1. <i>Effect of transcriptional inhibition on the ability of diclofenac to increase OA in KU812 cell line</i>	97
4.3.2. <i>Identification of candidate gene for OA regulation by diclofenac using gene expression profiling with GeneChip Human gene 1.0ST Array</i>	98
4.4. DISCUSSION	119
CHAPTER V THE ASSOCIATION BETWEEN OCT-1 ACTIVITY AND COX-2 IN CP-CML PATIENTS	124
5.1. INTRODUCTION	125
5.2. METHODS	130
5.2.1. <i>Examination of PTGS2 gene expression in primary samples</i>	130
5.2.2. <i>Examination of plasma levels of PGE2 and 15d-PGJ2</i>	131
5.3. RESULTS	131
5.3.1. <i>PTGS2 expression in CML patients</i>	131
5.3.2. <i>PGE2 and 15d-PGJ2 levels in CML patients</i>	133
5.3.3. <i>Correlation of PTGS2 expression and PGE2/15d-PGJ2 plasma levels</i>	135
5.3.4. <i>Correlation of OCT-1 Activity with PTGS2 expression, PGE2 and 15d-PGJ2 levels</i>	135
5.3.5. <i>Correlation of Molecular Response to imatinib and PTGS2 expression, PGE2 and 15d-PGJ2</i>	143
5.4. DISCUSSION	143
CHAPTER VI THE ASSOCIATION BETWEEN OCT-1 ACTIVITY AND PPARγ IN CP-CML PATIENTS	151
6.1. INTRODUCTION	152
6.2. METHODS	156

6.2.1.	<i>PPARγ expression levels after treatment with ligands in CML cell line</i>	156
6.2.2.	<i>PPARγ transcriptional activity in CML MNC samples</i>	156
6.2.3.	<i>PPARγ in isolated CD14⁺ and CD15⁺ cells from CML patients and healthy donors.</i>	158
6.3.	RESULT	158
6.3.1.	<i>The IUR assay in the presence of PPARγ ligands</i>	158
6.3.2.	<i>PPARγ expression and its subcellular localization in CML cells by Western Blotting.</i>	162
6.3.3.	<i>Correlation of OCT-1 activity with PPARγ transcriptional activity in CP-CML MNCs.</i>	166
6.3.4.	<i>Correlation between PPARγ transcriptional activity in MNCs and 15d-PGJ2 plasma levels in CP-CML samples</i>	171
6.3.5.	<i>Isolation of CD14⁺ cells and CD15⁺ cells from CML patients and normal donors...</i>	171
6.3.6.	<i>Protein and transcriptional activity levels of PPARγ in isolated CD14⁺ and CD15⁺ isolated cells</i>	176
6.4.	DISCUSSION	179
CHAPTER VII DISCUSSION		185
7.1.	INTRODUCTION	186
7.2.	MAJOR FINDINGS	187
7.2.1.	<i>Diclofenac and ibuprofen have opposite effects on functional OCT-1 activity in CML cells.</i>	187
7.2.2.	<i>Diclofenac regulates OA at the transcriptional level but not directly via SLC22A1 gene expression.</i>	188
7.2.3.	<i>The role of the COX pathway in OA regulation</i>	189
7.2.4.	<i>PPARγ negatively regulates OA in CML cells.</i>	190
7.3.	SUMMARY	191
7.4.	FUTURE DIRECTIONS	192
7.5.	CONCLUSION	194
APPENDIX A PUBLICATION ARISING FROM THIS THESIS		195
APPENDIX B SUPPLEMENTARY FIGURES AND TABLES		204
REFERENCES		211

List of Figures and Tables

Figure 1.1	The Philadelphia Chromosome and BCR-ABL oncogenes	4
Figure 1.2	BCR-ABL signal transduction pathways	6
Figure 1.3	Mechanism of imatinib.....	10
Figure 1.4	Mechanisms of imatinib resistance	13
Figure 1.5	Mechanism of action of nilotinib and dasatinib	16
Figure 1.6	Secondary structure and alignment of OCT1 with coding region SNPs.....	22
Figure 1.7	OCT-1 activity is associated with molecular response to imatinib therapy	25
Figure 3.1	Detailed results for one instance in Connectivity Map	64
Figure 3.2	Summary of Connectivity Map Compounds identified with different criteria to enhance OCT-1 activity	66
Figure 3.3	Bar plot for eight candidate drugs selected from Connectivity Map	69
Figure 3.4	Intracellular uptake of imatinib and OCT-1 activity in K562 cells in the absence or presence of CMAP candidates	72
Figure 3.5	OCT-1 activity in K562 cells in the absence or presence of NSAIDs	74
Figure 3.6	OCT-1 activity in K562 cells in the absence or presence of 11 clinically applicable drugs	76
Figure 3.7	Intracellular uptake of imatinib and OCT-1 activity in KU812 cells in the absence or presence of diclofenac, fenbufen, ibuprofen and paromomycin.....	77
Figure 3.8	The effects of temperature on the IUR of imatinib in KU812 cells and the OA in the presence of diclofenac and ibuprofen	79
Figure 3.9	The $IC_{50}^{imatinib}$ results in the presence or absence of diclofenac or ibuprofen	81
Figure 3.10	The effects of diclofenac and ibuprofen on the number of viable cells after 72 hours co-incubation with imatinib.	83
Figure 3.11	The effects of diclofenac or ibuprofen on OCT-1 activity in primary MNCs	85
Figure 4.1	Intracellular uptake of imatinib and OCT-1 activity in untreated and ActD pre-treated KU812 cells in the absence or presence of diclofenac	99
Figure 4.2	Quality control of RNA and microarray samples.....	100
Figure 4.3	Unsupervised clustering for the KU812 cells treated with imatinib and/or diclofenac for 2 hours	103

Figure 4.4	<i>PTGS2</i> mRNA expression levels in KU812 after treatment with imatinib and/or diclofenac for 2 hours	110
Figure 4.5	<i>SLC22A1</i> expression levels in KU812 after treatment with imatinib and/or diclofenac for 2 hours	111
Figure 4.6	Identification of candidate genes based on fold change greater than 1.5 and <i>p</i> -value less than 0.05 in KU812 cells	114
Figure 4.7	Gene expression levels of five candidate genes after the treatment with KU812 cells with imatinib and/or diclofenac for 2 hours	117
Figure 4.8	Gene expression levels of four candidate genes in KU812 after the treatment with imatinib and/or diclofenac for 2 hours.....	120
Figure 5.1	An overview of Cyclooxygenase pathway.....	126
Figure 5.2	<i>PTGS2</i> expressions in normal donors and CP-CML patients at diagnosis	132
Figure 5.3	Plasma levels of PGE2 and 15d-PGJ2 in normal donors and CP-CML patients at diagnosis	134
Figure 5.4	Correlation of <i>PTGS2</i> expression and plasma levels of PGE2/15d-PGJ2 in CP-CML patients at diagnosis	136
Figure 5.5	Relationship between OCT-1 activity and <i>PTGS2</i> expression levels in total white cells in CP-CML patients at diagnosis.....	138
Figure 5.6	Relationship between OCT-1 activity and <i>PTGS2</i> expression levels in mononuclear cells in CP-CML patients at diagnosis.....	140
Figure 5.7	Relationship between PGE2 and OCT-1 activity in CP-CML patients at diagnosis	141
Figure 5.8	Relationship between 15d-PGJ2 and OCT-1 activity in CP-CML patients at diagnosis	142
Figure 5.9	Relationship between the molecular response and <i>PTGS2</i> expression, PGE2 and 15d-PGJ2.....	144
Figure 6.1	The <i>PPARG</i> gene expression in cancer cell lines.....	157
Figure 6.2	Pre-treatment with PPAR γ antagonists diclofenac and GW9662 resulted in an increase in Imatinib IUR and OA in KU812 cells	160
Figure 6.3	Pre-treatment with PPAR γ agonists GW1929, rosiglitazone, or troglitazone resulted in a decrease in Imatinib IUR and OA in KU812 cells.....	161
Figure 6.4	<i>PPARG</i> expression levels remained unchanged after treatment with PPAR γ ligands in CML cell lines.....	163
Figure 6.5	PPAR γ ligands did not affect PPAR γ protein expression in KU812 cells after a 3-hour incubation.....	164

Figure 6.6	No significant change observed in the subcellular localization of PPAR γ protein after the treatment with PPAR γ ligands for 3 hours in KU812 cells.....	167
Figure 6.7	PPAR γ transcriptional activity in mononuclear cells was negatively associated with OCT-1 activity in CP-CML patients at diagnosis	169
Figure 6.8	PPAR γ activity in CP-CML MNCs had no correlation with the 15d-PGJ2 plasma levels.....	172
Figure 6.9	Procedure for isolation procedures for of neutrophils and monocytes	174
Figure 6.10	Purity of isolated neutrophils, monocytes and lymphocytes.....	175
Figure 6.11	PPAR γ protein expression and activity in CD14 ⁺ cells and CD15 ⁺ from normal donors and CP-CML patients at diagnosis	177
Figure 6.12	PPAR γ activity in CP-CML MNCs had no correlation with the PPAR γ activity of CD14 ⁺ cells or CD15 ⁺	178
Figure 6.13	Postulated interactions between OCT-1 and diclofenac or other NSAIDs in CP-CML MNCs.....	182
Figure 7.1	Working model of OCT-1 activity regulation by PPAR γ in CP-CML patients	193
Supplementary Figure 1	The heat map of SLC superfamily transporters in KU812 after treatment with imatinib and/or diclofenac for 2 hours	207
Supplementary Figure 2	The heat map of ABC superfamily transporters in KU812 after treatment with imatinib and/or diclofenac for 2 hours	208
Supplementary Figure 3	The heat map showing genes with fold change greater than 1.5 in dual treatment compared with imatinib only in KU812 cells.....	209
Supplementary Figure 4	The heat map showing genes with <i>p</i> -value<0.05 in KU812 cells	210

Table 3.1	NSAID agents selected in this study	61
Table 3.2	Clinically applicable drugs selected in this study.....	62
Table 3.3	Top 25 Connectivity Map compounds identified using an input signature weighted to OCT-1 as the primary up-regulated gene.....	68
Table 3.4	Candidate compounds selected from CMAP.....	70
Table 4.1	The ratio of 28S:18S rRNA and RNA integrity number (RIN) value of the samples for the Affymetrix GeneChip® Human gene 1.0 ST array.....	102
Table 4.2	Top 15 up-regulated or down-regulated genes ranked based on the fold change between the control and imatinib alone in KU812 cells.....	104
Table 4.3	Top 15 up-regulated or down-regulated genes ranked based on the fold change between the control and diclofenac alone in KU812 cells	105
Table 4.4	Top 15 up-regulated or down-regulated genes ranked based on the fold change between the control and co-administration of imatinib and diclofenac in KU812 cells	106
Table 4.5	Top 15 up-regulated or down-regulated genes ranked based on the fold change between imatinib alone and co-administration of imatinib and diclofenac in KU812 cells	107
Table 4.6	Top 25 genes ranked based on the <i>p</i> -value when comparing the conditions of control and imatinib alone to the conditions of diclofenac alone and dual treatment of two drugs	108
Table 4.7	List of candidate genes that meet the criteria of fold change >1.5 and <i>p</i> -value<0.05	116
Table 4.8	Direct target genes of PPAR γ identified using Chip Enrichment Analysis.	118
Supplementary Table 1	Primer sequences for RQ-PCR.....	205

ABBREVIATIONS

15d-PGJ2	15-deoxy- Δ 12,14-PGJ2
ABCB1/ABCG2	ATP binding cassette (ABC) transporter proteins B1 and G2
ABL	Abelson kinase
ACD	Anticoagulant Citrate Dextrose Solution Formula A
AP	Accelerated phase
ATP	Adenosine triphosphate
BC	Blast crisis
BCR	Breakpoint cluster region
BCR-ABL	Breakpoint cluster region-Abelson kinase fusion transcript/protein
BM	Bone marrow
BSA	Bovine serum albumin
C	Celcius
CCyR	Complete cytogenetic response
CD	Cluster of differentiation
cDNA	Complementary deoxyribonucleic acid
CHIP	Microarray chip
CHR	Complete haematologic response
CMAP	Connectivity map
CML	Chronic myeloid leukaemia
CMR	Complete molecular response
COX-1/2	Cyclooxygenase-1/2
CP	Chronic phase
Crkl	Crk-like protein
DDI	Drug-drug interaction
DEPC	Diethyl pyrocarbonate
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid
dNTPs	Deoxynucleotide triphosphates
DTT	Dithiothreitol
EDTA	Ethylene diamine tetraacetate
ERK	Extracelullar signal-regulated kinase
FBS	Foetal Bovine Serum
FC	Fold change
FITC	Fluorescein isothiocyanate
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
GUSB	Beta-glucuronidase
HBSS	Hanks Balanced Salt Solution

HCL	Hierarchical Clustering
IC50	50% inhibitory concentration
IFN-α	Interferon alpha
IgG	Immunoglobulin G
IRIS	International randomised study of interferon versus STI571
IUR	Intracellular uptake and retention
kD	Kilo Dalton
L	Litre
M	Molar
MACS	Magnetically activated cell sorting
MAPK	Mitogen activating protein Kinase
MCyR	Major cytogenetic response
mM	Milli Molar (10^{-3} Molar)
MMR	Major molecular response
MNC	Mononuclear cells
mRNA	Messenger ribonucleic acid
NF-κB	Nuclear factor of kappa light polypeptide gene enhancer in B-cells
ng	Nano gram (10^{-9} gram)
nM	Nano molar (10^{-9} Molar)
NSAIDs	Non-steroidal anti-inflammatory drugs
OA	OCT-1 activity
OAT-1	Organic anion transporter-1
OCT-1	Organic cation transporter 1
PB	Peripheral blood
PBS	Phosphate Buffered Saline
PCR	Polymerase chain reaction
p-CrkI	Phosphorylated Crk-like protein
PE	Phycoerythrin
PGE2	Prostaglandin E2
Ph	Philadelphia chromosome
PI3K	Phosphatidylinositol3 kinase
PPARγ	Peroxisome Proliferator Agonist Receptor
PTGS	Prostaglandin-endoperoxide synthase (COX)
p-value	Probability value
PVDF	Polyvinylidene fluoride
RANKL	Receptor activator of nuclear factor kappa-b ligand
RNA	Ribonucleic acid
ROC	Receiver operating characteristic
RPMI	Roswell Park Memorial Institute (media)

RQ-PCR	Real-time quantitative polymerase chain reaction
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide
SEM	Standard error of the mean
STI571	Signal transduction inhibitor 571 (imatinib)
TBP	TATA-binding protein
TBS	Tris buffered saline
TBST	Tris buffered saline with 0.1% Tween20
TKI	Tyrosine kinase inhibitor
TWC	Total white cells
U	Units
v/v	Volume per volume
vs.	versus
w/v	Weight per unit volume
µg	Micro gram (10^{-6} gram)
µM	Micro molar (10^{-6} molar)

PUBLICATIONS

Manuscripts

Wang J, Hughes TP, Kok CH, Saunders VA, Frede A, Groot-Obbink K, et al. Contrasting effects of diclofenac and ibuprofen on active imatinib uptake into leukaemic cells. *British Journal of Cancer*. 2012;106(11):1772-8 **Impact Factor 5.04**

Conference Abstracts

Wang J, Hughes TP, Kok CH, White DL, et al. Screening Potential OCT-1 Activity Enhancers may Improve the Outcome of Imatinib Therapy in CML. *New Directions in Leukaemia Research*, March 2010. Sunshine Coast, Australia. Poster Presentation.

Wang J, Hughes TP, Kok CH, White DL, et al. PPAR γ ligands modulate OCT-1 activity in BCR-ABL+ cell lines and primary CML cells. *Postgraduate Research Conference in the Faculty of Health Sciences*, August 2011. Adelaide, Australia. Poster Presentation.

Wang J, Hughes TP, Kok CH, White DL, et al. PPAR γ ligands modulate functional OCT-1 activity in a cell-lineage dependent way. *Haematology Society of Australia and New Zealand Annual Meeting*, October 2011. Sydney, Australia. Oral Presentation

Wang J, Hughes TP, Kok CH, White DL, et al. Non-Steroidal Anti-Inflammatory Drugs and imatinib; drug interactions that may impact efficacy. *American Society of Haematology Annual Meeting*, December 2011. San Diego, USA. Poster Presentation

Wang J, Hughes TP, Kok CH, White DL, et al. Dissecting the mechanism of OCT-1 Activity enhancement by diclofenac in CML cells. *New Directions in Leukaemia Research*, March 2012. Sunshine Coast, Australia. Poster Presentation.

Wang J, Hughes TP, Kok CH, White DL, et al. Manipulation to Enhance the Functional Activity of OCT-1 Influx Pump to Increase the Intracellular Concentration of Imatinib in Target Cells. *Postgraduate Research Conference in the Faculty of Health Sciences*, August 2012. Adelaide, Australia. Poster Presentation.

SCHOLARSHIPS & AWARDS

Poster Prize, Postgraduate Research Conference in the Faculty of Health Sciences. 2011.

For the abstract entitled “PPAR γ ligands modulate OCT-1 activity in BCR-ABL+ cell lines and primary CML cells”, Adelaide, August 2011.

Poster Prize, Postgraduate Research Conference in the Faculty of Health Sciences. 2012.

For the abstract entitled “Manipulation to Enhance the Functional Activity of OCT-1 Influx Pump to Increase the Intracellular Concentration of Imatinib in Target Cells”, Adelaide, August 2011.

Non-member Travel Grant, Haematology Society of Australia and New Zealand Annual Meeting, 2011.

For the abstract entitled “PPAR γ ligands modulate functional OCT-1 activity in a cell-lineage dependent way”, Sydney, October 2011.

PhD Scholarship, The Chinese Scholarship Council-the University of Adelaide Joint Scholarship. 2008-2012.

To provide support for the educational and professional development of Chinese researchers and other professionals undertaking a PhD in the University of Adelaide.

ACKNOWLEDGEMENTS

It is my great honor to study as a PhD candidate in the Melissa White Laboratory. Doing a PhD is a very challenging journey especially for an oversea student with English as a second language like me. Without the help and support from a large number of individuals, I would not be able to finish my research work and fulfill the whole procedure.

I would like to thank my fantastic supervisors, Professor Timothy P Hughes, Associate Professor Deborah L White and Associate Professor Richard D'Andrea for their generous help throughout my PhD candidature. Their patience, encouragement, and their understanding are essential for all of my achievements. Without their extremely rapid feedback, I would not be able to finish the thesis writing in such a short period.

I would like to thank Dr. Chung Hoow Kok for selflessly sharing his bioinformatics knowledge and laboratory techniques with me. Thanks to the entire lab team, from whom I have learnt a lot of valuable skills. Whenever I need help in the lab, I know they will be there for me. Thanks also go to all the students, past and present. The friendship established in our PhD student office is such a unique one that I would never ever forget. Also, I must thank Carine and Verity, who spent their precious time helping me with most tedious grammar mistakes in my writing and also gave me numerous constructive comments for my thesis.

I would like to acknowledge The Chinese Scholarship Council and The University of Adelaide for the financial support by providing me the CSC Joint PhD Scholarship.

I would like to thank my parents and my family members for their endless support as well as their understandings. I am also so lucky to have good friends who would listen to my complaints and help me regain my confidence. There is no doubt that it is the courage rooting from my family and friends that keeps me moving forward.

Lastly, my deepest thanks to my dear Ting XIA. Four years ago, it was you who helped me make the decision to study abroad, for which I have never felt regret. With your company, my days in Adelaide were much more colourful. Thank you for all the joy you have brought into my life.

ABSTRACT

In CML cells, the functional OCT-1 activity (OA) in mononuclear cells (MNC) from *de novo* chronic myeloid leukaemia patients in chronic phase (CP-CML) is significantly associated with imatinib-mediated *in vitro* tyrosine kinase inhibition and is a strong indicator of imatinib response.

Here we identified candidate drugs as potential OA enhancers using Connectivity Map analysis (CMAP). Their effects on OA were extensively validated in CML cell lines and patient samples, together with 12 NSAIDs and 11 commonly prescribed drugs. A significant enhancement of OA was observed after the treatment with diclofenac. Importantly this increase in OA translated to a significant increase in BCR-ABL kinase inhibition. Additionally, the long-term co-administration of diclofenac sensitized CML cells to imatinib on cell proliferation. In CML patients' mononuclear (MNC) samples, diclofenac significantly increased OA especially in patients with low OA. In contrast, ibuprofen significantly decreased the OA in CML cell lines and primary samples. This effect on OA also translated into a reduction in BCR-ABL kinase inhibition and an increase in cell growth in the presence of imatinib. Unlike diclofenac, the inhibitory effect of ibuprofen is also observed in normal cells most likely in an independent manner from OCT-1 protein.

The enhancement of OA by diclofenac could be eliminated in the presence of Actinomycin D (a transcription inhibitor), indicating that diclofenac regulated OA at a transcriptional level. However, the expression of *SLC22A1* (*OCT-1*) remained unchanged after treatment with diclofenac. Since diclofenac is an inhibitor for cyclooxygenase-2 (COX-2), the potential involvement of COX-2 in OA regulation was investigated. The plasma concentration of 15-deoxy- Δ 12,14-PGJ2 (15d-PGJ2), a

prostaglandin product by COX-2, was significantly associated with OA in CP-CML. Given 15d-PGJ2 is a potent agonist for peroxisome proliferator-activated receptor γ (PPAR γ) and diclofenac is also a known PPAR γ ligand, whether PPAR γ had a role in OA regulation was then investigated. Significant increase in OA in KU812 cells was observed after treatment with PPAR γ antagonist GW9962, while treatment with PPAR γ agonists (GW1929, troglitazone, or rosiglitazone) significantly decreased OA. In addition, there was a significant negative association between OA and the PPAR γ transcriptional activity in CP-CML MNC collected at diagnosis. Further identification of the key factor contributing to high PPAR γ activation in patients with low OA may provide better understanding of intrinsic OA interpatient variability, as well as the clinical and biological relevance of PPAR γ in CML.

DECLARATION

This thesis contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published work contained within this thesis (as listed below) resides with the copyright holder(s) of those works.

Wang J *et al.* British Journal of Cancer. 2012. (Appendix A)

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue, the Australasian Digital Theses Program (ADTP) and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Jueqiong Wang

November 2012