Mesenchymal Stem Cells for the Treatment of Myocardial Infarction-Induced Ventricular Dysfunction.

Dr James David Richardson

MBBS (Hons) MRCP

Department of Medicine
Faculty of Health Sciences
The University of Adelaide, South Australia

&

Cardiovascular Research Centre
The Royal Adelaide Hospital, South Australia

&

Bone and Cancer Research Laboratories

Division of Haematology

Institute of Medical and Veterinary Science

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Declaration

Declaration

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James David Richardson

December 2013

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- Cardiac magnetic resonance, transthoracic and transoesophageal echocardiography: a comparison of in vivo assessment of ventricular function in rats. Richardson JD, Bertaso AG, Frost L, Psaltis PJ, Carbone A, Koschade B, Wong DT, Nelson AJ, Paton S, Williams K, Azarisman S, Worthley MI, Teo KS, Gronthos S, Zannettino AC, Worthley SG. Lab Anim. 2013 Oct; 47(4):291-300. doi: 10.1177/0023677213494373
- Impact of timing and dose of mesenchymal stromal cell therapy in a preclinical model of acute myocardial infarction. Richardson JD, Bertaso AG, Psaltis PJ, Frost L, Carbone A, Paton S, Nelson AJ, Wong DT, Worthley MI, Gronthos S, Zannettino AC, Worthley SG. J Card Fail. 2013 May;19(5):342-53. doi: 10.1016/ j.cardfail.2013.03.011. PMID: 23663817 [PubMed - in process]
- 4. Incremental Benefits of Repeated Mesenchymal Stromal Cell Administration Compared to Solitary Intervention in a Preclinical Model of Myocardial Infarction. JD Richardson, PJ Psaltis, L Frost, A Carbone, S Paton, AG Bertaso, AJ Nelson, DT Wong, MI Worthley, S Gronthos, A Zannettino, SG Worthley. Cytotherapy 2013

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- 2. **Richardson JD**, Paton S, Frost F, Carbone A, Bertaso AG, Nelson AJ, Psaltis PJ, Wong DT, Worthley MI, Gronthos S, Zannettino ACW, Worthley SG. Prospectively Isolated, Hypoxic-Preconditioned Mesenchymal Stem Cells Significantly Attenuate Myocardial Infarction-Induced Ventricular Dysfunction In Rats. Heart, Lung & Circulation 2012; S21: S1-S142.
- 3. **Richardson JD**, Frost F, Bertaso AG, Carbone A, Paton S, Nelson AJ, Psaltis PJ, Wong DT, Worthley MI, Gronthos S, Zannettino ACW, Worthley SG. Sequential Mesenchymal Stem Cell Interventions Produce Greater Myocardial Repair Than Solitary Treatment in Rats After Acute Myocardial Infarction. Heart, Lung & Circulation 2012; S21: S1-S142.
- 4. **Richardson JD**, Bertaso AG, Koschade B, Wong DT, Williams K, Frost F, Carbone A, Paton S, Nelson AJ, Psaltis PJ, Worthley MI, Teo KS, Gronthos S, Zannettino ACW, Worthley SG. Cardiac Magnetic Resonance, Transthoracic and Transoesphageal Echocardiography: A Comparison of In Vivo Ventricular Function Assessment in Rats. Heart, Lung & Circulation 2012; S21: S143-S316.

5. **Richardson JD**, Bertaso AG, Wong DT, Frost F, Carbone A, Nelson AJ, Psaltis PJ, Paton S, Koschade B, Williams K, Worthley MI, Teo KS, Gronthos S, Zannettino ACW, Worthley SG. Assessment of Regional Myocardial Function in Rats using 1.5T Cardiac Magnetic Resonance Imaging. Heart, Lung & Circulation 2012; S21: S143-S316.

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- Cardiac Society (CSANZ) 2012 Oral Presentations:
 - o "Immediate Mesenchymal Stem Cell Therapy Provides Greater Attenuation of Myocardial Injury Than Deferred Treatment in Rats After Acute Myocardial Infarction."
 - o "Sequential Mesenchymal Stem Cell Interventions Produce Greater Myocardial Repair Than Solitary Treatment in Rats After Acute Myocardial Infarction"
- Cardiac Society (CSANZ) 2012 Poster Presentations:
 - o "Prospectively Isolated, Hypoxic-Preconditioned Mesenchymal Stem Cells Significantly Attenuate Myocardial Infarction-Induced Ventricular Dysfunction In Rats".
 - o "Cardiac Magnetic Resonance, Transthoracic and Transoesphageal Echocardiography: A Comparison of In Vivo Ventricular Function Assessment in Rats".
 - "Assessment of Regional Myocardial Function in Rats using 1.5T Cardiac Magnetic Resonance Imaging"
- Ralph Reader Young Investigator Award, Cardiac Society Australia and New Zealand 2012 (Runner-up)
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- Genesis Research Award 2012 (Winner)

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Abbreviations

medium μg Microgram FACS Fluorescence-activated cell sorting μL Microlitre μm Micrometre μm Micromolar ANOVA Analysis of Variance β FS Fractional Shortening ANOVA Analysis of Variance β Gram AUC Area under the curve β FP Green fluorescent protein BM Bone marrow HBSS Hanks' balanced salt solution BMMNC Bone marrow mononuclear cells CD Cluster of differentiation CFU-F Colony forming units-fibroblast CI Confidence interval CI Confidence interval CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide DNA Deoxyribonucleic acid EC Endothelial cells EDD End-diastolic dimension EF Ejection Fraction EIC Endothelial progenitor cells V/V Volume per volume ESD End-systolic dimension W/V Weight per volume ESD End-systolic dimension WV Weight per volume	αΜΕΜ	Alpha modification of Eagle's	ESV	End-systolic volume
μL Microlitre FCS Foetal calf serum μm Micrometre FTC Fluorescein isothiocyanate μM Micromolar FS Fractional Shortening ANOVA Analysis of Variance g Gram AUC Area under the curve GFP Green fluorescent protein BM Bone marrow HBSS Hanks' balanced salt solution BMMNC Bone marrow mononuclear cells HGF Hepatocyte growth factor CD Cluster of differentiation HR Heart rate CDNA Complementary IGF Insulin-like growth factor cDNA Complementary IGF Insulin-like growth factor CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor I (SDF-1) LV Left ventricle (or left ventricular) DMEM Dulbecco's modification of Eagle's medium MACS Magnetic-activated cell sorting DMSO Dimethy				
μm Micrometre FITC Fluorescein isothiocyanate μM Micromolar FS Fractional Shortening ANOVA Analysis of Variance g Gram AUC Area under the curve GFP Green fluorescent protein BM Bone marrow HBSS Hanks' balanced salt solution BMMNC Bone marrow mononuclear cells HGF Hepatocyte growth factor CD Cluster of differentiation HR Heart rate cDNA Complementary deoxyribonucleic acid IGF Insulin-like growth factor CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) LV Left ventricle (or left ventricular) DMEM Dulbecco's modification of Eagle's medium MACS Magnetic-activated cell sorting DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyr	μg	Microgram	FACS	
MM Micromolar ANOVA Analysis of Variance BM Bone marrow BONE Marro	μL	Microlitre	FCS	Foetal calf serum
ANOVA Analysis of Variance g Gram AUC Area under the curve GFP Green fluorescent protein BM Bone marrow HBSS Hanks' balanced salt solution BMMNC Bone marrow mononuclear cells CD Cluster of differentiation HR Heart rate cDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ECC Endothelial progenitor cells v/v Volume per volume	μm	Micrometre	FITC	Fluorescein isothiocyanate
AUC Area under the curve GFP Green fluorescent protein BM Bone marrow Mononuclear cells CD Cluster of differentiation HR Heart rate CDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	μΜ	Micromolar	FS	Fractional Shortening
BM Bone marrow HBSS Hanks' balanced salt solution BMMNC Bone marrow mononuclear cells CD Cluster of differentiation HR Heart rate CDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	ANOVA	Analysis of Variance	g	Gram
BMMNC Bone marrow mononuclear cells CD Cluster of differentiation HR Heart rate CDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	AUC	Area under the curve	GFP	Green fluorescent protein
cells CD Cluster of differentiation HR Heart rate cDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast IHD Ischaemic heart disease CI Confidence interval IL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	BM	Bone marrow	HBSS	Hanks' balanced salt solution
CDNA Complementary deoxyribonucleic acid CFU-F Colony forming units-fibroblast CI Confidence interval CMR Cardiac magnetic resonance CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide DNA Deoxyribonucleic acid EC Endothelial cells EDD End-diastolic dimension EDV End-diastolic volume EF Ejection Fraction EDC Endothelial progenitor cells EPC Endothelial progenitor cells VI Interleukin IL Interleukin It Interleukin Interleukin Interleukin Interleukin Interleukin	BMMNC		HGF	Hepatocyte growth factor
deoxyribonucleic acid CFU-F Colony forming units-fibroblast IIL Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDV End-diastolic volume EF Ejection Fraction EC Endothelial progenitor cells V/V Volume per volume EPC Endothelial progenitor cells V/V Volume per volume	CD	Cluster of differentiation	HR	Heart rate
CI Confidence interval II Interleukin CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	cDNA	1	IGF	Insulin-like growth factor
CMR Cardiac magnetic resonance PBS Phosphate buffered saline CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	CFU-F	Colony forming units-fibroblast	IHD	Ischaemic heart disease
CXCL12 Stromal cell-derived factor 1 (SDF-1) DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	CI	Confidence interval	IL	Interleukin
DMEM Dulbecco's modification of Eagle's medium DMSO Dimethyl sulphoxide mg Milligram DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells v/v Volume per volume	CMR	Cardiac magnetic resonance	PBS	Phosphate buffered saline
Eagle's medium DMSO Dimethyl sulphoxide DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells V/v Volume per volume	CXCL12		LV	Left ventricle (or left ventricular)
DNA Deoxyribonucleic acid MI Myocardial Infarction EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	DMEM		MACS	Magnetic-activated cell sorting
EC Endothelial cells SD Standard deviation EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	DMSO	Dimethyl sulphoxide	mg	Milligram
EDD End-diastolic dimension MPC Mesenchymal precursor cells EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	DNA	Deoxyribonucleic acid	MI	Myocardial Infarction
EDTA Ethylenediaminetetraacetic acid MSC Mesenchymal stromal/stem cells EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	EC	Endothelial cells	SD	Standard deviation
EDV End-diastolic volume n Sample number EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	EDD	End-diastolic dimension	MPC	Mesenchymal precursor cells
EF Ejection Fraction TGF Transforming growth factor ELISA Enzyme-linked immunosorbent assay VEGF Vascular endothelial growth factor EPC Endothelial progenitor cells v/v Volume per volume	EDTA	Ethylenediaminetetraacetic acid	MSC	Mesenchymal stromal/stem cells
ELISA Enzyme-linked immunosorbent assay EPC Endothelial progenitor cells VEGF Vascular endothelial growth factor VegF Vascular endothelial growth factor	EDV	End-diastolic volume	n	Sample number
assay factor EPC Endothelial progenitor cells v/v Volume per volume	EF	Ejection Fraction	TGF	Transforming growth factor
	ELISA		VEGF	
ESD End-systolic dimension w/v Weight per volume	EPC	Endothelial progenitor cells	v/v	Volume per volume
	ESD	End-systolic dimension	w/v	Weight per volume

Thesis Abstract

Despite current treatment options, cardiac failure after myocardial infarction (MI) is associated with significant morbidity and mortality so highlighting a compelling clinical need for novel therapeutic approaches. Based on promising pre-clinical data, stem cell therapy has been suggested as a possible therapeutic strategy. Early studies largely utilised autologous bone marrow cells with only modest benefits observed in clinical trials. Of the alternative candidate cell types evaluated, mesenchymal stromal/stem cells (MSCs) have shown promise, however their clinical application for mainstream cardiovascular use is currently hindered by several important limitations. Consequently, this has prompted intense efforts to advance the therapeutic properties of MSCs through cell optimisation strategies.

Allogeneic sources of MSC appear to hold several important advantages over autologous bone marrow/BM mononuclear cells (BMMNC); (1) MSC can be derived from young, healthy donors thereby enhancing the absolute yield and functional biology of MSCs; (2) The cell product could be prepared well ahead of time, so making very early MSC treatment feasible, e.g. after primary percutaneous intervention, when myocardium remains viable; (3) MSC could be optimised to potentially advance their therapeutic efficacy.

The studies described in this thesis utilised all of the above features to address the primary aims of:

- 1. Reviewing the literature and writing a review regarding the optimisation of the cardiovascular therapeutic properties of MSC.
- Develop an allogeneic MSC population optimised by the novel combination of prospective-isolation enrichment and hypoxic preconditioning. Furthermore, evaluate the in vivo function of optimised MSC compared to conventional plasticadherent isolation of MSC (PA-MSC).

- 3. Develop a reliable non-invasive assessment of rat ventricular function using 1.5T cardiac magnetic resonance and evaluate this modality against conventional methods (transthoracic echocardiography) and novel modalities in rats (transoesophageal echocardiography).
- 4. Explore the impact of the timing of MSC intervention and cell dose after MI, now that immediate cell intervention is feasible clinically and these factors have not previously been investigated.
- 5. Explore the potential benefits of immediate and deferred MSC treatment after MI, two very different time points a novel concept.

An allogeneic source of MPCs was derived from donor rat bone marrow. In contrast to conventional plastic-adherent isolation of MSC, an enriched and optimised MSC population prepared by prospective isolation of immature MPCs (via a CD45 immunodepletion step) and hypoxic preconditioning was established. In cell-based experiments, optimised MSC were compared to same-donor plastic-adherence isolated MSC and demonstrated superior in-vitro differentiation and colony forming capacity than PA-MSC.

To evaluate the effects of MSC treatment after MI in rats, highly accurate and reproducible imaging techniques are required. Cardiac magnetic resonance (CMR) is widely regarded as the gold standard modality, however the use of standard 1.5T "clinical" MR scanners in rodents has only been achieved by a handful of investigators worldwide and none have used contemporary MR techniques. CMR was then evaluated against conventional imaging modalities (transthoracic echocardiography) and novel methods in rats (transoesophageal echocardiography).

Allogeneic MSC permits immediate treatment, previously impossible with autologous stem cells, therefore this potentially important variable (timing) was assessed. Myocardial infarction was induced by ligation of the left anterior descending artery in rats. Optimised

MSC were then injected into the myocardium either immediately after MI or one week later, at one of two cell doses. This study provided an innovative comparison of these clinically relevant time points and demonstrated value at both times. Furthermore, greater efficacy was observed with immediate treatment, which displayed high sensitivity to MSC dose, with benefits largely localised to the infarct territory. Deferred treatment, though less effective, was not dose dependant and primarily influenced non-infarct myocardium.

Given the disparate, yet beneficial effects, of immediate and deferred MSC intervention the benefit of combining MSC treatment at both time points was investigated. Again, this was undertaken in the rat model of MI, with CMR determination of ventricular function. This novel study showed clinically relevant improvements in LV function and confirmed the differential distribution of MSC repair according to timing of cell intervention.

In summary, the studies described in this thesis provide new evidence outlining the merits of prospective isolation and hypoxic preconditioning of MSC. Furthermore they demonstrate the reparative effects of these cells and provide novel insights into the significance of timing of MSC intervention on efficacy and mode/distribution of effect, which can be further augmented through treatment both time points after MI.