Salinity detection and control of sodium transport in *Arabidopsis thaliana*

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List of Abbreviations

3' three prime, of nucleic acid sequence

5' five prime, of nucleic acid sequence number

% percent

approximately

× times

°C degrees Celsius

aa amino acid

ACPFG Australian Centre for Plant Functional Genomics

AGRF Australian Genome Research Facility

ANOVA analysis of variance
Arabidopsis Arabidopsis thaliana
At Arabidopsis thaliana

AtHKT1;1-C24 AtHKT1;1 allele from C24

AtHKT1;1-Col-0 AtHKT1;1 allele from Col-0

AtHKT1;1-C24 AtHKT1;1 protein form C24

AtHKT1;1-Col-0 AtHKT1;1 protein from Col-0

AVP Arabidopsis vacuolar pyrophosphatase

BAC bacterial artificial chromosome

BLAST basic local alignment search tool

bp base pairs, of nucleic acid

BSA bovine serum albumin

C24 Arabidopsis ecotype C24

Ca²⁺ calcium ion

[Ca²⁺]_{cvt} cytosolic free calcium concentration

CaCl₂ calcium chloride

CaMV cauliflower mosaic virus

Cat. # catalogue number

CBL calcineurin like-B protein

cDNA complimentary deoxyribonucleic acid

CI- chloride ion

CIPK calcineurin like-B interacting protein kinase

cm centimetre(s)

Col-0 Arabidopsis ecotype Columbia-0

Col-0×C24 mapping population with Col-0 and C24 as parents

cRNA complimentary ribonucleic acid

d day(s)

dATP deoxyadenosine triphosphate dCTP deoxycytidine triphosphate dGTP deoxyguanosine triphosphate

dH₂0 deionised water

DNA deoxyribonucleic acid

dNTP deoxynucleotide triphosphate

dS deciSiemens
DTT dithiothreitol

dTTP deoxythymidine triphosphate

ECe electrical conductivity

EDTA ethylenediaminetetraacetic acid

E_{rev} reversal potential

FAO Food and Agricultural Organization of the United Nations

g gram(s)

G conductance

g gravity

gDNA genomic deoxyribonucleic acid

GFP green fluorescent protein GUS β -glucuronidase protein

H+ hydrogen ion

H₂0 water

HCI hydrochloric acid

HKT high affinity potassium transport

hr hour(s)
I current

IDSE intron dependent spatial expression

IME intron mediated enhancement

K+ potassium ion

kb kilo base pairs, of nucleic acid

KCI potassium chloride

kg kilogram(s)

KOH potassium hydroxide

L litre(s)

Ler Arabidopsis ecotype Landsberg erecta

LB left border, of T-DNA sequence

LB media luria betani media

M molar

mg milligram(s)
Mg²⁺ magnesium ion

MgCl₂ magnesium chloride

min minute(s)

mRNA messenger ribonucleic acid

miRNA micro ribonucleic acid

mL millilitre(s)
mm millimetre(s)
mM millimolar

mol mole

mOsm milliosmole

MS-media media, Murashige and Skoog media

mV millivolt

n sample size
N/A not applicable

N₂ nitrogen
mA milliampere
Na+ sodium ion

NaCl sodium chloride NaOH sodium hydroxide

NCBI National Center for Biotechnology Information

ng nanogram(s)

NHX Na+/H+ exchanger

nm nanometre(s)
nM nanomolar

nosT bacterial nopaline synthase terminator sequence

NPK ratio of nitrogen, phosphate and potassium in fertilizer

o/n overnight

OD₆₀₀ optical density measured at 600 nm

pC24 2.7 kb promoter region upstream of AtHKT1;1 from C24

pCol-0 2.7 kb promoter region upstream of *AtHKT1;1* from Col-0

PCR polymerase chain reaction

pCR8 entry vector pCR™8/GW/TOPO Gateway®

PI propidium iodide

qRT-PCR quantitative reverse transcription polymerase chain reaction

QTL quantitative trait loci

RB right border, of T-DNA sequence RdDM RNA mediated DNA methylation

RIL recombinant inbred line

RNA ribonucleic acid
RO reverse osmosis
rpm rounds per minute
RT room temperature

RT-PCR reverse transcription polymerase chain reaction

s second(s)

Salty Salt water crocodile, mascot
Sc Saccharomyces cerevisiae
SDS sodium dodecyl sulfate

- coalam acaccy, canate

Semi-qPCR semi-quantitative polymerase chain reaction

siRNA short interfering ribonucleic acid
SNP single nucleotide polymorphism(s)

SOS salt overly sensitive

T1 primary Arabidopsis transformant containing T-DNA

T2 progeny of T1 plant
TAE tris-acetate-EDTA

T-DNA transfer deoxyribonucleic acid TE transposable element

TE transposable element
TF transcription factor

T_m melting temperature, of primers

U units

UAS upstream activation sequence

uid
A β -glucuronidase gene

UTR untranslated region

UV ultraviolet V voltage

v/v volume per volume

wk week(s)

w/v weight per volume

Xenopus Xenopus laevis

μg microgram(s)

 μL microlitre(s)

μm micrometre(s)

μM micromolar

µmol micromole(s)

Abstract of thesis

Soil salinity is a major abiotic stress, reducing crop yields and endangering global food security. With salt affected areas increasing, understanding the molecular mechanisms of salinity stress is of great importance. Plant salinity stress can be categorised into two phases, the initial shoot ion independent osmotic stress and the later ionic stress. Osmotic stress occurs as soon as the plant encounters salt in the soil and results in an immediate reduction in the shoot growth rate. Ionic stress is caused by the accumulation of ions such as Na⁺ and Cl⁻ in the cytosol of cells in the shoot and results in the inhibition of cellular processes and induces premature leaf senescence.

The two Arabidopsis thaliana ecotypes Col-0 and C24 have previously been identified as interesting candidates to study plant salinity tolerance. The Col-0 ecotype is less salt tolerant than the C24 ecotype, based on its reduction in dry weight under stressed conditions. This is despite C24 accumulating significantly more Na* in the shoot than Col-0. Interestingly, C24 also appeared to be less responsive to salt stress, as transcript levels of several key salt responsive genes are not substantially altered in response to salt stress. The AtHKT1;1 gene is one key gene found to be not up-regulated in C24 during salt stress. AtHKT1;1 encodes a protein likely to be involved in the retrieval of Na+ from the xylem thereby reducing the amount of Na⁺ translocating to the shoot. In this thesis the C24 and Col-0 HKTs are compared at the protein and transcriptional levels. Electrophysiological analysis in Xenopus oocytes and a functional assay in yeast confirm Na⁺ transport properties of both AtHKT1;1 proteins and, interestingly, indicated AtHKT1;1 from both ecotypes had the ability to transport K+. To determine the difference in expression profile between the two ecotypes, a series of AtHKT1;1promoter::GFP and AtHKT1;1promoter::AtHKT1;1cDNA constructs were tested in Arabidopsis. Results suggest that both the Col-0 and C24 AtHKT1;1 promoters are able to drive expression of the downstream genes, suggesting that differences in the promoter region are not responsible for the lack of AtHKT1;1 expression in C24. A transposable element identified in the second intron of the C24 AtHKT1;1 genomic sequence may be important in causing the lack of *AtHKT1;1* expression in roots.

Furthermore, the reduced responsiveness of C24 to salt stress is investigated in relation to how salt is initially perceived by the plant. An assay using aequorin bioluminescence was used to compare the responses in the salt stress inducible Ca^{2+} -signatures of Col-0 and C24 seedlings. Excitingly, C24 appears to be missing part of the Ca^{2+} signature observed in the salt responsive plant Col-0, suggesting that C24 may not detect the ion component of salt stress. This potentially provides a suitable screening methodology for the identification of as yet unknown components in the early stages of the salt signalling pathway. An attempt is made to develop a screening assay suitable for performing QTL analysis on an available $Col-0 \times C24$ mapping population, based on measuring changes in transcript levels of salt responsive genes.

Declaration

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution to Sandra Manuela Schmöckel and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

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Date			

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